

UNCLASSIFIED

AD NUMBER
AD279670
NEW LIMITATION CHANGE
TO Approved for public release, distribution unlimited
FROM No Foreign
AUTHORITY
DTRA Ltr., 6 May 99

THIS PAGE IS UNCLASSIFIED

UNCLASSIFIED

AD 279 670

*Reproduced
by the*

ARMED SERVICES TECHNICAL INFORMATION AGENCY
ARLINGTON HALL STATION
ARLINGTON 12, VIRGINIA



UNCLASSIFIED

NOTICE: When government or other drawings, specifications or other data are used for any purpose other than in connection with a definitely related government procurement operation, the U. S. Government thereby incurs no responsibility, nor any obligation whatsoever; and the fact that the Government may have formulated, furnished, or in any way supplied the said drawings, specifications, or other data is not to be regarded by implication or otherwise as in any manner licensing the holder or any other person or corporation, or conveying any rights or permission to manufacture, use or sell any patented invention that may in any way be related thereto.

AD No. 279670

ASTIA FILE COPY

279670

210177

DASA 1246

TECHNICAL PROGRESS REPORT

DEFENSE ATOMIC SUPPORT AGENCY

WASHINGTON 25, D. C.

A Shock Tube Utilized to Produce Sharp-
rising Overpressures of 400 Milliseconds
Duration and Its Employment in
Biomedical Experimentation

by

D. R. Richmond
V. R. Clare
V. C. Goldizen
D. E. Pratt
R. T. Sanchez
C. S. White

Technical Progress Report
on
Contract No. DA-49-146-XZ-055

This work, an aspect of investigations dealing with the
Biological Effects of Blast from Bombs, was supported
by the Defense Atomic Support Agency of the Department
of Defense.

(Reproduction in whole or in part is permitted for any
purpose of the United States Government.)

Lovelace Foundation for Medical Education and Research
Albuquerque, New Mexico

April 7, 1961

A Shock Tube Utilized to Produce Sharp-
rising Overpressures of 400 Milliseconds
Duration and Its Employment in
Biomedical Experimentation

FORWORD

The purpose of the present study is two-fold; namely, (1) to describe "long-duration" pressure pulses simulating those produced by nuclear explosions, and (2) to present the result of an interspecies animal study when the data on six different animals are extrapolated to a biologic target the size of man.

The results are limited to single-pulses of overpressure which rise almost instantaneously to a maximum and endure for about 400 msec. As far as man is concerned, these results do not apply to very short duration overpressures (50 msec or less), to pressures having a slow or step-wise rise to a maximum, and to oscillating pressures of considerable magnitude. Also the data apply to overpressure injury under circumstances in which displacement from blast pressures and winds is minimized and blast-energized missiles do not occur. The findings are applicable to military and industrial situations involving potential exposure to explosive phenomena, e.g., nuclear weapons, high explosives, tanks containing gases or liquids under high pressure, etc.

The interspecies study is a part of research carried on since 1952 aimed at better understanding human response to the several environmental variations produced by low and high yield explosives.

ABSTRACT

A shock tube employed for blast biology studies is described along with the results of one series of experiments.

The shock tube is air-driven and utilizes Mylar plastic diaphragms. The compression chamber is 17.5 ft in length and 40.5 in. I.D.; it reduces in diameter to 23.5 in. over a 3-ft-long transition section just upstream of the diaphragm station. The expansion chamber consists of 30 ft of 23.5 in. I.D. tubing followed by 22 ft of 40.5 in. I.D. tubing. It is closed distally by a steel end-plate to generate high pressures from the reflected shock. Three vents in the expansion side of the system serve to control the duration of the overpressure and to eliminate multiple reflections by bleeding off the reflected shock as it travels upstream.

An interspecies correlation is presented based on mortality data from six species of experimental animals with an extrapolation to a 70 kg animal.

TABLE OF CONTENTS

	<u>Page</u>
Abstract	1
Introduction	4
Methods	5
Geometry of the Shock Tube	5
Instrumentation	7
Calibration of Gauges	11
Biological Material	11
Results	14
Pressure-time Records	14
Mortality	14
Interspecies Correlation Between LD ₅₀ Reflected Pressures and Species Weight	21
Discussion	23
The General Behavior of the Pressure Source	23
The Wave Form Achieved	24
Biological Implications of the Data	25

LIST OF TABLES

1. Animals Used in This Study	13
2. Mortality as Related to the Magnitude of the Incident and Reflected Shock Fronts	17
3. Summary of the Results from the Probit Analysis	19

LIST OF FIGURES

Figure No. 1 Diagram of the Shock Tube	6
Figure No. 2 View of the Compression Chamber	8
Figure No. 3 View of the Expansion Chamber	9
Figure No. 4 View of the Expansion Chamber	10
Figure No. 5 One Channel of Instrumentation	12

Table of Contents (continued)

	<u>Page</u>
Figure No. 6 Pressure-time Records taken with Quartz Piezo-electric Gauges	15
Figure No. 7 Calibration Curves Relating the Magnitude of the Incident and Reflected Shocks with the Various Compression Chamber Pressures	16
Figure No. 8 Probit Regression Lines Relating the Per Cent Mortality in Probit Units to the Log of the Reflected Shock Pressure for Six Animal Species	20
Figure No. 9 The Relation Between Body Weight and Fast- rising Overpressures of 400 msec Duration Needed to Produce 50 Per Cent Mortality . .	22

INTRODUCTION

The shock tube has proven to be a valuable tool in studying the biological effects of air blast.¹⁻⁵ By appropriate modifications, the device can be made to generate a wide variety of wave forms some of which closely resemble those recorded inside structures exposed to full-scale nuclear detonations.⁶⁻⁷ The capability of creating and reproducing at will a desired variation in environmental pressure has made it possible to initiate comparative interspecies experimentation without which there can be no clear understanding of hazardous and non-hazardous wave forms. In this light, it is well to emphasize that the air-driven shock tube has an unusual versatility and offers many other advantages. For example, it has become possible to achieve the precision in performance that is quite critical for biological experimentation. Too, with proper care, there need be no complications due to secondary missiles, hot gases, and toxic fumes which often plague explosive-driven tubes. Finally, pressure-time recording instruments, as well as those employed to monitor pathophysiological processes, can be mounted just outside the test chamber in close proximity to the animal under study.

It is well now to turn to the two main purposes of this paper; namely, first to describe a shock tube assembled to produce single pressure pulses that rise almost instantaneously to a maximum and endure for about 400 msec, which is a wave form quite comparable to those produced under certain circumstances by nuclear detonations; and second, to present mortality data on six species of animals all exposed in a similar geometry to similar pressure-time phenomena that varied among the species mostly with respect to the magnitude of the overpressure.

METHODS

Geometry of the Shock Tube

Figure 1 presents a diagram of the blast tube. The over-all length is approximately 70 ft. The compression chamber measures 17 ft 5 in. and has an internal diameter of 40.5 in. The driver reduces in diameter from 40.5 in. to 23.5 in. over a 3-ft long transition section at its diaphragm end. The expansion chamber is 53 ft 4 in. in length, of which 30 ft is 23.5 in. I.D. tubing followed by 22 ft of 40.5 in. I.D. tubing. The increase in diameter of the expansion chamber from 23.5 in. to 40.5 in. occurs rather abruptly over a span of little over 1 ft.

As noted in Fig. 1, the driver is stationary since it is "nested" in a massive reinforced concrete back-stop. Consequently, the various components on the expansion side are on casters to facilitate diaphragm replacement. The end of the tube is closed by a 2-in. thick steel plate — the end-plate.

Three vents, each 10x14x8 in., are located at the upstream end of the 40.5 in. tubing of the expansion chamber. These vents serve to "tailor" the wave form and will be discussed later.

The diaphragms employed are 40x40 in. sheets of polyester plastic film (Dupont Mylar). They have holes pre-drilled to match the bolt holes in the flanges. For their insertion they are simply bolted between the appropriate flanges. Mylar sheets of 0.010 and 0.0075 in. thickness are employed, a predetermined number being used to hold a given pressure in the driver section.

Diaphragms are ruptured by lead pellets from a sawed-off 12-gauge shotgun mounted on the tube. The gun fires straight down through the tube, through holes appropriately placed in the top and bottom of the tube. The

ARRANGEMENT A'

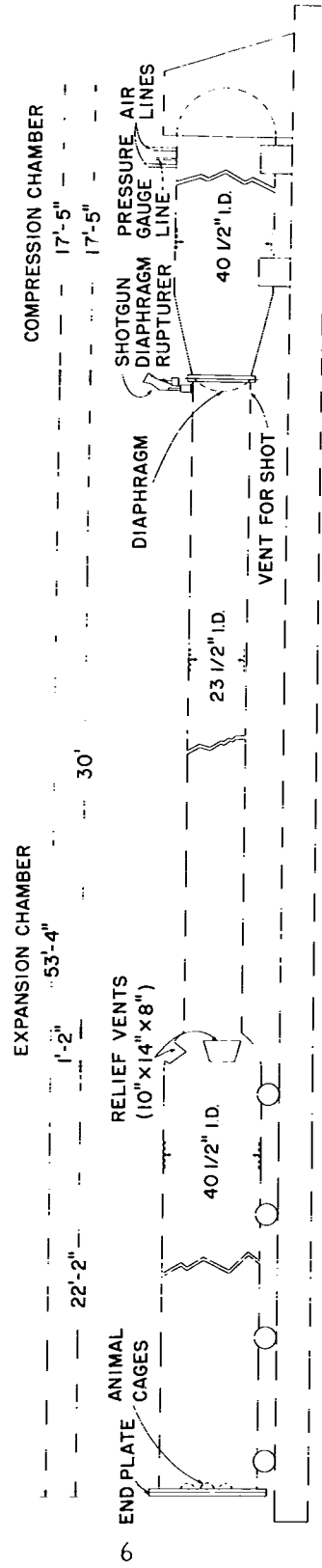


Figure 1

diaphragms under pressure obligingly bow-out in the path of the shotgun blast. Since the shot passes out of the tube through the vent in the under side, it is not carried downstream where it might interfere with the test.

Photographic views taken of the blast tube are presented in Figs. 2, 3 and 4. As seen in Fig. 3 access into the end of the tube, for animal placement, is accomplished by separating the tube at the distal most 24 in. flanges.

The shack seen near the end-plate houses the pressure-time recording instruments and the equipment necessary to operate the shock tube.

Instrumentation

In these particular experiments pressure-time variations were measured by Quartz piezo-electric gauges (Model 401)* that were shock mounted flush with the inside of the tube. Gauges were routinely placed at the end-plate and at short distances upstream from the end-plate. The signal from a gauge was fed through low noise cable with an amplifier-calibrator* into an oscilloscope (Tektronix Model 535A) having a Type L or Type 53/54C pre-amplifier plug-in unit. This oscilloscope had a single sweep circuit which prevented retriggering after the desired trace was recorded.

In using the oscilloscope, the horizontal sweep was externally triggered by the signal from another "trigger gauge" of Barium Titanate** located just upstream of the recording gauge. Pre-triggering the sweep allowed a reference trace of base line to be photographed during the time the shock travelled from the "trigger gauge" to the recording gauge. To insure sufficient voltage to trigger reliably the oscilloscope sweep, the signal from the trigger gauge was amplified. A power supply (Type 105) with amplifiers** (Type 104A) was used for that purpose.

*Purchased from Kistler Instrument Corporation, North Tonawanda, N.Y.

**Purchased from Atlantic Research Corporation, Alexandria, Va.



Figure 2



Figure 3

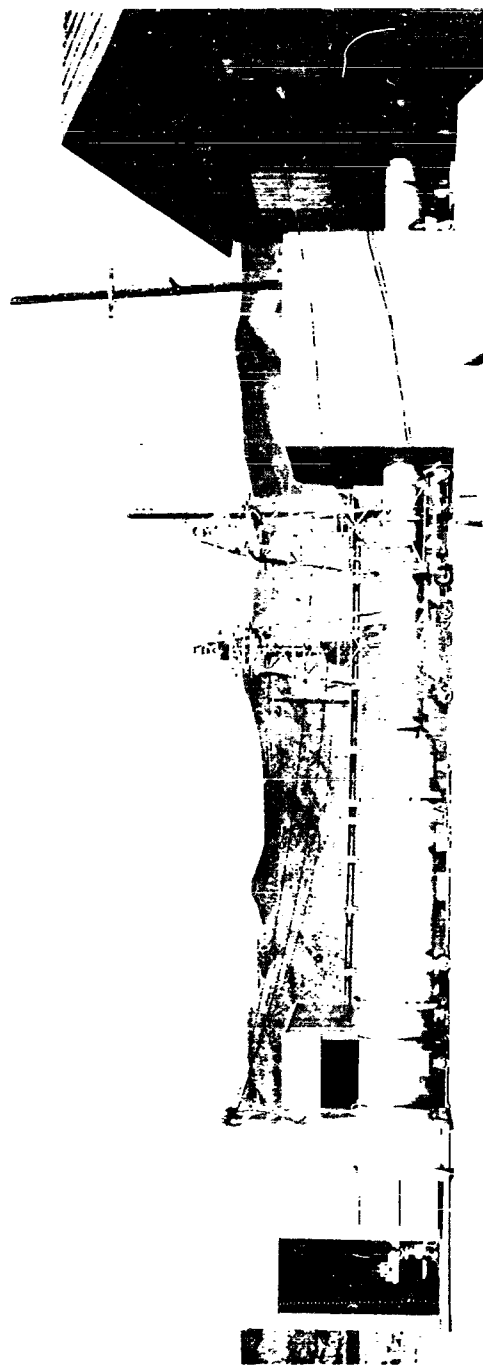


Figure 4

The sweep on the face of the oscilloscope was photographed for the permanent pressure-time record with a Polaroid Land Camera mounted in a periscope assembly. The components that make up a channel of instrumentation are shown in Fig. 5. Available at present are 8 channels of pressure-time measuring gear plus three channels for monitoring the pressures within biological systems.

Calibration of Gauges

The quartz piezo-electric gauges were statically calibrated using a small pressure vessel. Their dynamic performance was also checked on a 12-in. - diameter calibration shock tube. Barium titanate and lead zirconate crystal gauges were calibrated on the calibration shock tube by measuring their voltage out-put at various shock pressures. The latter were computed from measuring the speed of the shock with a Hewlett-Packard Electronic Counter that was started and stopped by gauges placed 18 in. apart.

Biological Material

Table 1 summarizes the number of animals from each of the 6 different species that were employed in this study, along with their body weights and ages. Of the total of 569 animals, 140 were mice; 164, rats; 96, guinea pigs; 104, rabbits; 35, dogs; and 30, goats. Their body weights ranged between 22 g for the mouse to 20 kg for the goat.

All animals were exposed to the overpressure against the end-plate. Except for a few of the mice and rats that managed to turn end-for-end in their cages, all animals were right-side-on to the incident shock front. The dogs and goats were restrained in harness - the mice, rats, guinea pigs, and rabbits in wire mesh cages as described in a previous study.⁴

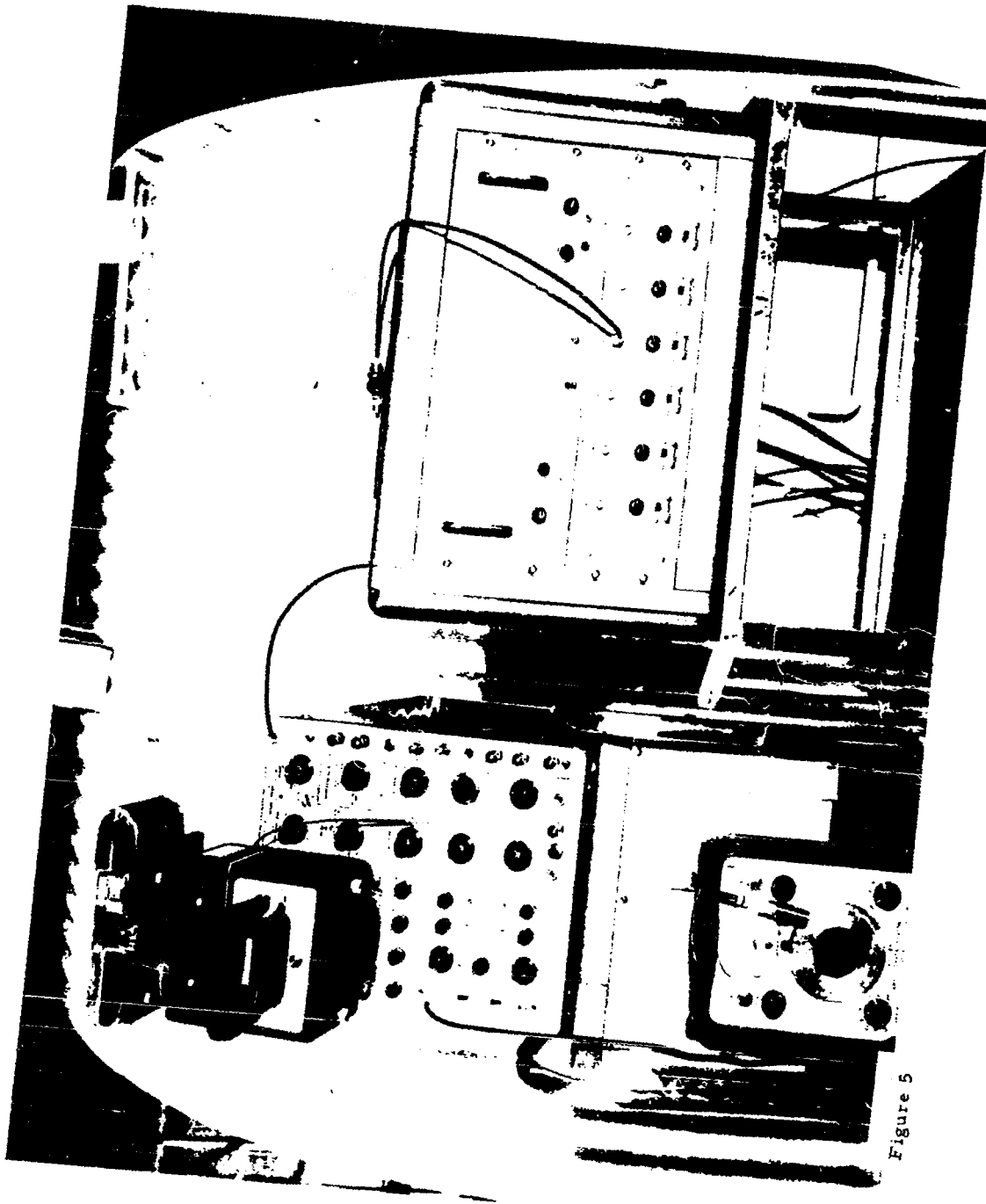


Figure 5

TABLE 1
ANIMALS USED IN THIS STUDY

Species	Number of animals	Body weight	Age, months
Mouse (Webster strain)	140	22 g $\pm$ 1.9*	1 - 1-1/2
Rat (Sprague-Dawley)	164	192 g $\pm$ 25	2 - 2-1/2
Guinea pig (English short haired)	96	445 g $\pm$ 37	3-1/2 - 4
Rabbit (New Zealand White)	104	1.97 kg $\pm$ 0.26	2-1/2 - 3
Dog (Mongrel)	35	15.1 kg $\pm$ 3.1	—
Goat (Mixed breed)	30	20.5 kg $\pm$ 3.6	4 - 5

*Mean and standard deviation

RESULTS

Pressure-time Records

Illustration of four piezo-electric gauge records are presented in Fig. 6. The upper two records were from gauges 1 and 2 located face-on flush with the inside of the end-plate, while the lower two records were from gauges 3 and 4 side-on at 3 in. and 9 in. upstream from the end-plate. As seen in the figure, the face-on gauges see the incident and reflected shock pressures as one step, whereas those gauges side-on resolve the shocks as two pressure steps. It can also be noted from the record of gauge 2 that the rise time of the pressure on the end-plate was less than that of the gauge. The oscillations seen on the records to damp out rapidly were, of course, due to the natural frequencies of the quartz crystal gauges.

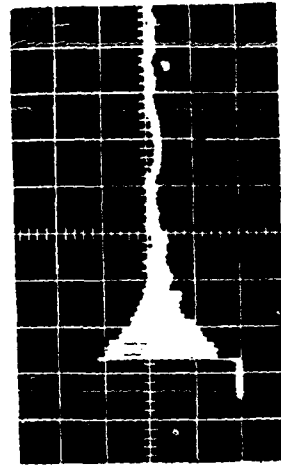
The duration of the overpressure (near 400 msec) can be read from the record of gauge 4 since the oscilloscope that recorded the out-put of that gauge was run at a fairly slow sweep-speed of 50 msec/cm.

The magnitude of the incident and reflected shock fronts obtained with a range of compression chamber pressures between 17 psi to 170 psi was plotted and is shown in Fig. 7. The reflected shock pressures from 25 psi to nearly 60 psi were sufficient for the compilation of the dose-response curves. The durations of the overpressures over that range of reflected pressures were between 350 to 412 msec.

Mortality

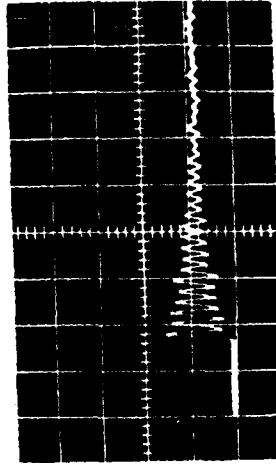
The 24-hr lethality obtained with the sample of animals exposed at the various driver pressures and the associated mean incident and reflected pressures are given in Table 2. Previous studies have shown that the mortality

PRESSURE-TIME RECORDS QUARTZ PIEZO GAUGES



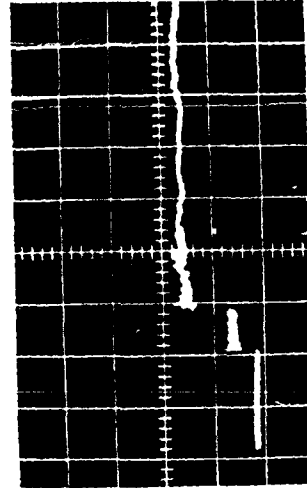
18.7
psi/cm

0.2 msec/cm
Gauge 1 Face-on at End Plate



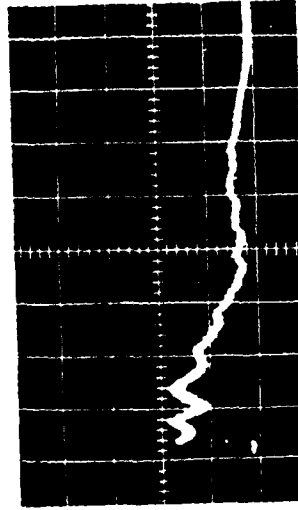
45.4
psi/cm

0.1 msec/cm
Gauge 2 Face-on at End Plate



26.0
psi/cm

0.5 msec/cm
Gauge 3 Side-on 3" from End Plate



22.6
psi/cm

50 msec/cm
Gauge 4 Side-on 9" from End Plate

Figure 6

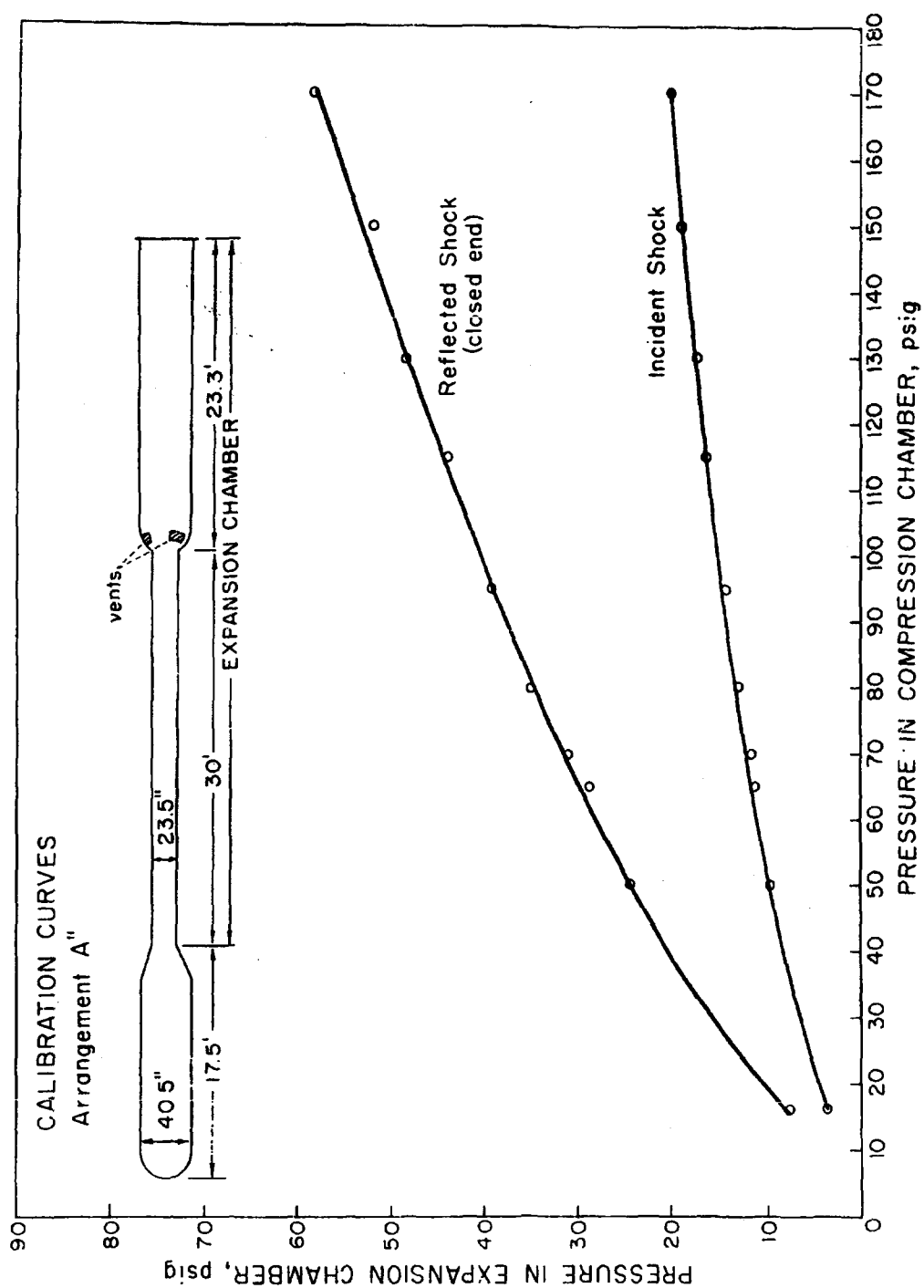


Figure 7

TABLE 2
MORTALITY AS RELATED TO THE MAGNITUDE OF THE INCIDENT
AND REFLECTED SHOCK FRONTS

Compression chamber pressure psi	Incident shock psi	Reflected shock psi	Mortality	
			No. dead Total	Per cent
<u>Mice:</u>				
58	10.2 (9.5-11.0)*	27.2 (25.4-27.9)*	7/40	17.5
65	11.2 (10.9-11.4)	28.9 (28.6-29.1)	8/30	26.7
70	10.9 (10.5-11.2)	31.6 (31.3-31.9)	18/30	60.0
80	13.3 (13.2-13.4)	35.6 (34.7-36.6)	18/20	90.0
130	18.0 (17.5-18.5)	50.8 (49.5-52.2)	20/20	100.0
<u>Rats:</u>				
70	11.4 (10.9-11.9)	30.4 (30.0-30.8)	2/30	6.7
80	13.8 (13.4-14.1)	34.4 (33.8-34.7)	12/34	35.3
95	13.6 (12.7-14.0)	38.6 (37.9-39.6)	35/50	70.0
105	15.4 (15.0-15.8)	42.2 (40.3-43.2)	35/40	87.5
130	18.0 (17.5-18.5)	50.8 (49.5-52.2)	10/10	100.0
<u>Guinea pigs:</u>				
65	10.9 (10.6-11.0)	29.0 (28.6-29.8)	1/24	4.2
80	12.0 (11.0-12.4)	33.9 (33.2-34.3)	9/24	37.5
80	13.4 (12.8-14.0)	35.8 (35.5-36.5)	18/24	75.0
95	13.5 (11.3-14.7)	39.9 (38.1-41.0)	21/24	87.5
<u>Rabbits:</u>				
48	9.2 (8.7-9.8)	22.8 (20.0-24.7)	1/20	5.0
58	10.5 (9.6-10.9)	26.9 (25.0-29.1)	10/28	35.7
75	12.2 (11.5-13.6)	32.6 (30.5-34.6)	16/24	66.7
80	13.1 (12.5-13.6)	36.6 (35.2-38.3)	17/20	85.0
95	14.3 (14.0-14.7)	40.5 (40.4-40.5)	12/12	100.0
<u>Dogs:</u>				
95	14.7 (13.9-15.1)	39.2 (39.0-39.4)	0/5	0
115	16.6 (16.0-17.4)	44.1 (42.2-44.9)	1/10	10.0
130	17.7 (16.9-18.4)	48.1 (46.8-49.6)	6/10	60.0
150	19.0 (18.2-19.9)	53.0 (50.0-55.3)	9/10	90.0
<u>Goats:</u>				
130	16.7 (15.8-17.8)	44.9 (44.0-46.2)	2/10	20.0
150	18.2 (16.2-19.3)	51.4 (47.3-56.2)	4/10	40.0
170	19.4 (19.0-20.1)	56.9 (56.6-57.4)	3/5	60.0
170	20.3 (20.0-20.7)	59.3 (58.7-60.1)	4/5	80.0

Computed LD₅₀'s: Mice, 30.7 psi; rats, 36.3 psi; guinea pigs, 34.5 psi; rabbits, 29.6 psi; dogs, 47.8 psi; goats, 53.0 psi.

*Mean and range.

was best correlated with the reflected shock pressures.⁴ Since plotting the per cent mortality against the reflected pressure produced a sigmoid curve, the probit analysis was applied to the data which transforms it into a straight line.⁸ The general form of the probit equation of this type was:

$$Y = a + b \log X$$

when Y was the per cent mortality in probit units, X was the reflected shock pressure, and a and b were the intercept and slope constants, respectively. The LD₅₀ (the pressure associated with 50 per cent mortality) was obtained by substituting the probit value of 5 (equal to 50 per cent mortality) for Y in the appropriate probit equation and solving for X. The LD₅₀'s obtained in this manner, the standard errors of the LD₅₀'s and the probit regression line equations are listed in Table 3. The LD₅₀'s in order of increasing species weights were as follows: mouse, 30.7; rat, 36.3; guinea pig, 34.5; rabbit, 29.6; dog, 47.8; and goat, 53.0 psi. The trend was for the LD₅₀'s to increase with the species size. The probit regression lines with the data points are presented graphically in Fig. 8.

The LD₅₀'s fell into three groups according to statistical tests that compared them for significant differences. The mouse and rabbit were not significantly different from one another at the 95 per cent confidence level; neither were the guinea pig and rat, nor the dog and goat. However, each pair, as grouped above, were different statistically from one another. That is, the mouse and rabbit were significantly below the rat and guinea pig who were below the dog and goat.

Appropriate statistical tests that compared the slopes of the six probit mortality curves revealed that they were essentially parallel at the 95 per cent

TABLE 3
SUMMARY OF THE RESULTS FROM THE PROBIT ANALYSIS

Species	LD ₅₀ psi	Standard error of the LD ₅₀ , psi	Probit equation constants	
			intercept, a	slope, b
Mouse	30.7	± 0.56	-23.63	19.25
Rat	36.3	± 0.61	-23.82	18.48
Guinea pig	34.5	± 0.64	-28.50	21.78
Rabbit	29.6	± 0.90	-13.63	12.67
Dog	47.8	± 1.06	-49.47	32.43
Goat	53.0	± 2.79	-16.68	12.57

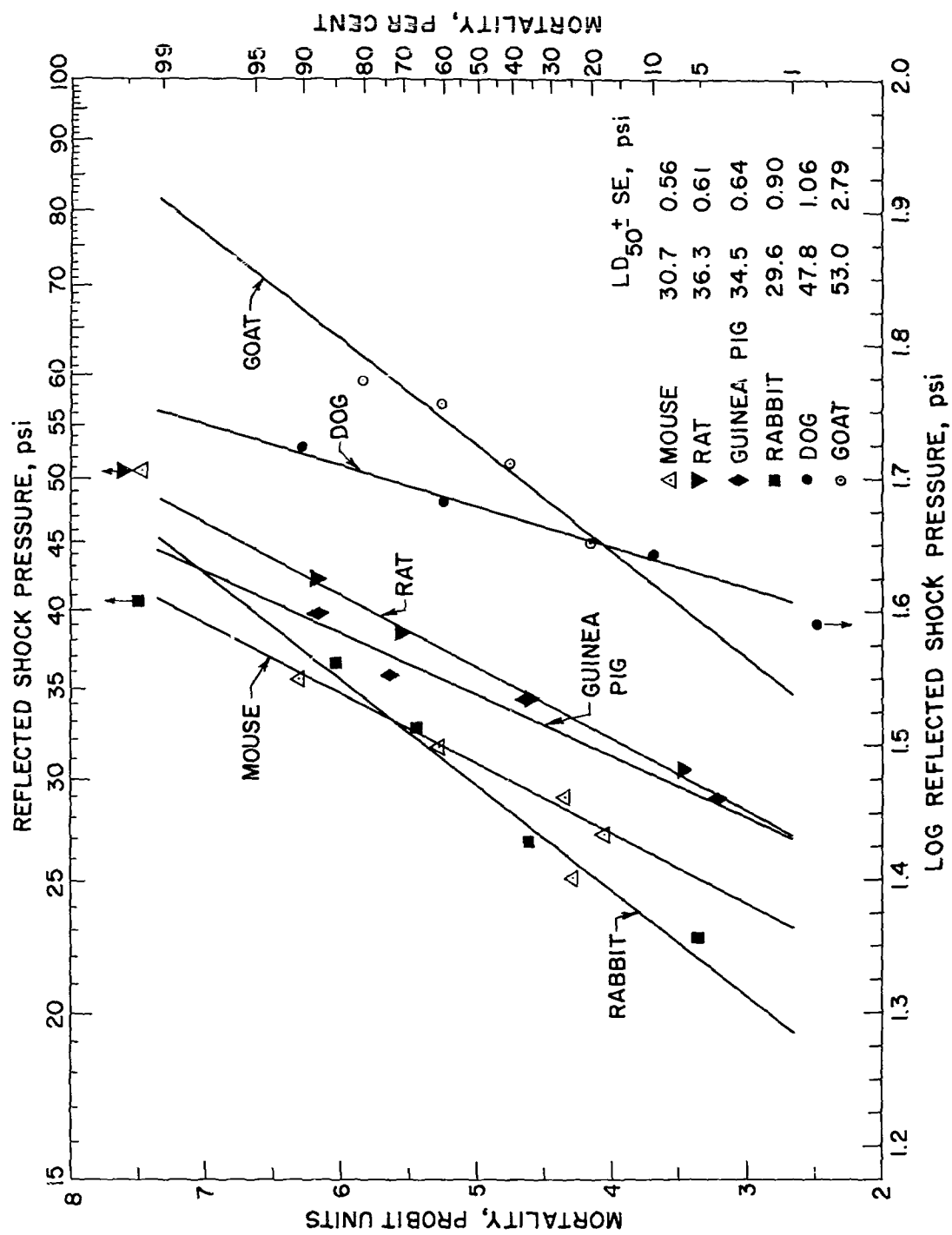


Figure 8

confidence level. This may be interpreted to mean that the same type of stimulus was instrumental in killing the animals from each species.⁸

Interspecies Correlation Between LD₅₀ Reflected Pressures and Species Weight

A log-log plot of the LD₅₀'s versus the mean body weight of each species appears in Fig. 9. The regression line equation, calculated by the method of least squares that best fit the data, was:

$$\text{Log LD}_{50} = 1.3673 + 0.06939 \log \text{BW}$$

where

LD₅₀ = the reflected pressure required for 50 per cent
lethality, psi

BW = mean body weight of the species, grams

1.3673 = the intercept constant

0.06939 = the slope constant

According to Fig. 9 there is fair agreement between the calculated line and the measured points save for the rabbit whose point was noticeably low for its weight. The standard error of the regression estimate was 0.0602 log units or 13.9 per cent.

Included in Fig. 9 is the calculated point for a 70 kg animal. It was obtained by simply solving the regression equation for a body weight of 70,000 g. In other words, this predicts 50.5 psi to be the LD₅₀ reflected pressure for a large animal the size of man exposed against a reflecting surface to a single-pulse overpressure rising almost instantaneously to near a maximum and enduring for about 400 msec.

RELATION BETWEEN BODY WEIGHT AND FAST-RISING
OVERPRESSURES OF 400 MILLISECONDS DURATION
NEEDED TO PRODUCE 50 PERCENT MORTALITY

Animals exposed side-on against the
plate closing the end of a shock tube

REGRESSION EQUATION

$$\log (LD_{50}) = 1.3673 + 0.06939 \log (BW)$$

Where LD_{50} = Pressure required for 50% mortality, psi

BW = Average body weight of the group, grams

Standard Error of Estimate: 0.06102 log units (13.9%)

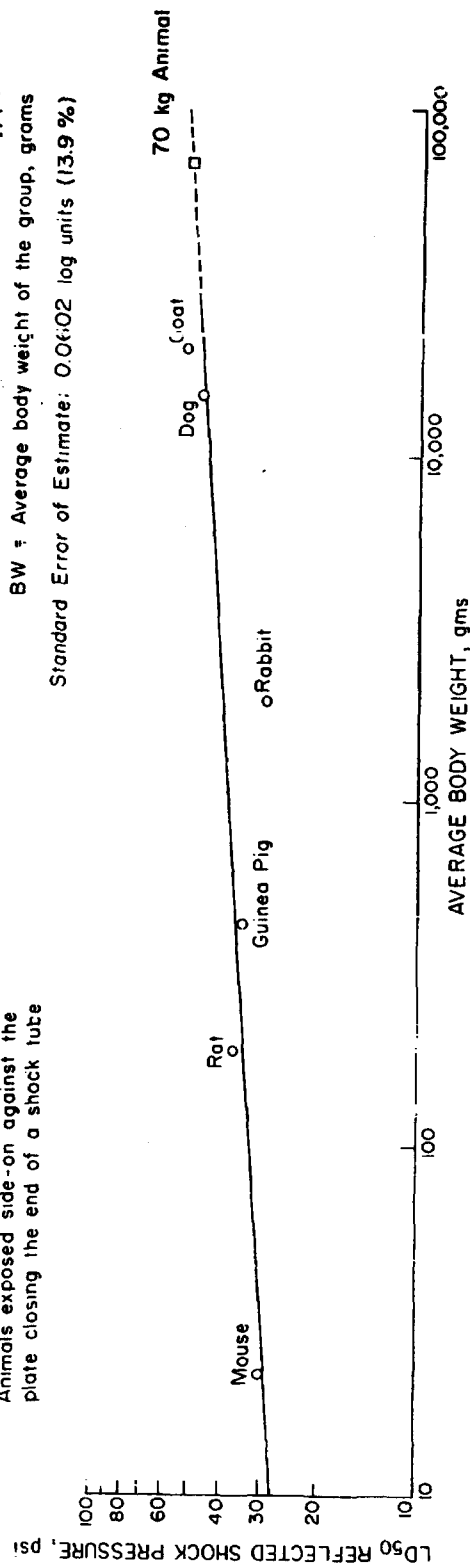


Figure 9

DISCUSSION

By way of discussion, remarks in three areas are indicated; i.e., (a) the general behavior of the pressure source, (b) the wave form achieved using the shock-tube hardware described, and (c) the biological implications of the data.

The General Behavior of the Pressure Source

A study of Table 2, which gives the mean and range of the incident and reflected shock overpressures for the several experiments, shows that the pressure source utilized did not function perfectly as far as reproducibility of overpressures were concerned. For example, the variation in incident shocks for the different pressure groupings ranged from a few tenths of a psi to a maximum of 3.4 psi (see the guinea pig data for 95 psi compression chamber pressure). In the latter case the mean incident shock pressure was 13.5 psi with a minimum 2.2 psi below and a maximum 1.2 psi above the mean, respectively. Variability for reflected shocks, as could be expected, was somewhat greater, but only ranged from a few tenths of a psi to a maximum of 8.9 psi for the goat exposure involving a compression chamber pressure of 150 psi. The maximum reflected shock was 4.1 psi above the mean while the minimum was 4.1 psi below the mean of 51.4 psi. Though this last variability represents a performance about ± 10 per cent from the mean pressure, it is indeed the maximal "misbehavior" of the shock tube. In most cases, the variability was much less as a study of the data in Table 2 will show. In fact, judging from earlier experience, the reproducibility performance noted in the present study is considered quite good indeed.

The Wave Form Achieved

Before the vent holes were added to the distal section of the shock tube, the wave form had many undesirable characteristics: first, the duration of the overpressure was longer than desired; second, there was considerable "crowning" of the overpressure after the development of the reflected shock, that is, the maximal pressure developed was considerably above the reflected shock pressure; third, marked oscillations in the overpressure occurred during the falling phase of the pulse. These were many in number and the amplitude of the oscillations was high.

After addition of the vents, all of these untoward characteristics were improved, though the wave form achieved is, from one point of view, still not exactly that desired. For example, attention is directed to the lower right-hand trace in Fig. 6 which shows the pressure pulse in its entirety on a somewhat compressed time scale. The reader will note that the maximal overpressure achieved is still somewhat above (about 5 psi) the reflected shock pressure of near 36 psi. Also, the pressure oscillations present represent a swing in pressure of about 19 psi, occurring in 20 msec; this involves a variation of about $1/2$ the magnitude of the reflected shock pressure. Also, if 20 msec is taken for the half-cycle time, the frequency of this portion of the oscillation is 25 cps.

Thus, wave forms, having the characteristics of the pattern just discussed, raises uncertainties as regards assessing the biological implications of the pressure pulse, which uncertainties would be markedly lessened if the wave forms were "clean"; e.g., the crowning and pressure oscillations were absent. This point will be pursued later.

However this may be, it can be said from another point of view that the

wave forms recorded in the present study are similar to some of those seen in full-scale operations in Nevada and are, therefore, quite desirable.⁶ In truth, there is need for knowing the biology of both "clean" and "unclean" pressure pulses and in this light the present study contributes a great deal.

Biological Implications of the Data

At least two interesting questions arise in the biological area: first, what portions of the pressure pulse contribute to injury and second, what faith may or may not be put in extrapolating interspecies data to larger animals?

With regard to the first question, it is now clear that the animal poorly tolerates very "fast"-rising overpressures compared with "slow"-rising pressure pulses of the same magnitude.⁴⁻⁶ This fact directs attention to the rising phase of the overpressure as critical and implies that damage is associated with shock loading. If indeed this is so and the animal suffers damage from the initial pressure rise, then any after-coming pressure variations might well enhance the injury. Also involved, of course, is the amplitude and frequency of the pressure oscillations, particularly as the latter may "match" the natural frequency of the thorax-abdominal system of a given species.

Also, if shock loading is one of the critical factors biologically, one would expect that any degrading of the average rate of pressure rise — all other factors being equal — would be associated with increased tolerance to overpressure. Such is the case empirically. Guinea pigs, for example, exposed at increasing distances from the end-plate of a shock tube show an increase in the P_{50} from 37 psi against the end-plate to 57 psi at 1 ft from the end-plate; e.g., the pressure rises in two rapid steps instead of one.⁴ Likewise, dogs tolerate without fatality well over 150 psi if the time to P_{max} is

30 msec or longer, even though the pulse duration is as long as 5 to 20 sec.⁵

Involved in the biological interpretation of the hazards of overpressure is the duration of a single, "fast"-rising overpressure. Data are at hand which indicate that, within limits, the duration of such a pulse of overpressure is significant; e.g., for the dog about 220 psi enduring for 1.8 msec is fatal, whereas only 75 psi is fatal when the duration is 11.8 msec; for smaller animals, duration is important only for shorter intervals like fractions of a millisecond to 1, 2, or 3 msec.⁹

Currently, data simply are not at hand to allow a more definitive and quantitative interpretation of those characteristics of pulses of overpressure which define hazard clearly. It remains for future work to systematically spell out the criticality of rates of pressure rise, overpressure duration, step loading with two or more shocks in the rising phase of a pulse, and finally the biological meaning of oscillating overpressures.

Last, there remains the question of extrapolating interspecies blast data to larger (or smaller) animals. There is little to be said except that one should approach the extrapolation of data to any given species, including man, with considerable caution. First, it should be noted that all the animals used in the work described here were mounted against a reflecting surface and any extrapolation should keep this fact in mind. Second, the shock overpressures related or correlated with the interspecies mortality were the reflected shock pressures and one should not confuse an incident or local static-free field pressure — corresponding to the incident pressures reported here — with the reflected shock. Third, exactly what the pressure reflection would be when an incident wave strikes an animal in the open is not currently clear to the authors and certainly the data presented do not bear upon this point.

Fourth, the extrapolation set forth in Fig. 9 applies strictly to the pulse form studied and to an overpressure duration of about 400 msec. Fifth, for these conditions, it is not known whether man is more or less tolerant than might be implied by the 70 kg point marked in Fig. 9. A few data do exist for the human case which relate 235 psi⁹ and 450 psi¹⁰ with human mortality, but these concern only high explosive-produced overpressures of a few msec duration and do not apply at all to the longer duration case. It would seem, therefore, that the extrapolation indicating that a 400 msec single, sharp-rising overpressure of 50.5 psi applies to as large an animal as man and might well be considered a tentative figure subject to all the conditions mentioned above. In the meantime, one must await the results of further experimental work to define more definitively man's tolerance to blast.

REFERENCES

1. B. Cassen, L. Curtis and K. Kistler, "Initial Studies of the Effect of Laboratory Produced Air Blast on Animals," J. Aviat. Med. 21:38-47 (1950).
2. H. Celander, C. Clemedson, U. Ericsson, and H. Hultman, "The Use of Compressed-air-operated Shock Tube for Physiological Blast Research," Acta Physiol. Scand. 32:6-13 (1955).
3. D. R. Richmond and R. V. Taborrelli, "Some Results of a Shock Tube for Biomedical Investigation," pp. 56-69, Proceedings of Second Shock Tube Symposium, Report SWR-TM-58-3, Air Force Special Weapons Center, Air Research and Development Command, Kirtland Air Force Base, New Mexico, March 5-6, 1958.

4. D. R. Richmond, R. V. Taborelli, F. Sherping, M. B. Wetherbe, R. T. Sanchez, V. C. Goldizen, and C. S. White, "Shock Tube Studies of the Effects of Sharp-rising, Long-duration Overpressures on Biological Systems," pp. 171-194, Proceedings of Third Shock Tube Symposium, Report SWR-TM-59-2, Hqs. Air Force Special Weapons Center, Kirtland Air Force Base, New Mexico, March 10-12, 1959. Also published as TID-6056, U. S. Atomic Energy Commission, Washington, D. C., dated March 10, 1959.
5. D. R. Richmond, M. B. Wetherbe, R. V. Taborelli, T. L. Chiffelle, and C. S. White, "The Biologic Response to Overpressure. I. Effects on Dogs of Five to Ten-second Duration Overpressures Having Various Times of Pressure Rise," J. Aviat. Med. 28:447-460, 1957.
6. C. S. White, T. L. Chiffelle, D. R. Richmond, W. H. Lockyear, I. G. Bowen, V. C. Goldizen, H. W. Merideth, D. E. Kilgore, B. B. Longwell, J. T. Parker, F. Sherping, and M. E. Cribb, "The Biological Effects of Pressure Phenomena Occurring Inside Protective Shelters Following Nuclear Detonation," Operation Teapot Report, WT-1179, U. S. Atomic Energy Commission, October 1956.
7. D. R. Richmond, R. V. Taborelli, I. G. Bowen, T. L. Chiffelle, F. G. Hirsch, B. B. Longwell, J. G. Riley, C. S. White, F. Sherping, V. C. Goldizen, J. D. Ward, M. B. Wetherbe, V. R. Clare, M. L. Kuhn, and R. T. Sanchez, "Blast Biology - A Study of the Primary and Tertiary Effects of Blast in Open Underground Protective Shelters," Operation Plumbbob Report, WT-1467, U. S. Atomic Energy Commission, June 30, 1959.
8. D. J. Finney, Probit Analysis. A Statistical Treatment of the Sigmoid

Response Curve, 2nd Edition, Cambridge University Press, New York, 1952.

9. H. Desaga, "Blast Injuries," Chap. XIV-D, German Aviation Medicine, World War II, Vol. II, pp. 1274-1293, U. S. Government Printing Office, Washington, D.C., 1950.
10. R. B. Fisher, P. L. Krohn, and S. Zuckerman, "The Relationship Between Body Size and the Lethal Effects of Blast," Ministry of Home Security Report R.D. 284, Oxford University, Oxford, England, 1941.

DISTRIBUTION

ARMY AGENCIES

Dep Chief of Staff for Mil Ops., DA, Washington 25, DC Attn: Dir of SW&R	1
Chief of Research & Develop, DA, Washington 25, DC Attn: Atomic Division	1
Assistant Chief of Staff, Intelligence, DA, Washington 25, DC	1
Chief Chemical Officer DA, Washington 25, DC	2
Chief of Engineers DA, Washington 25, DC ATTN: ENGNB	1
Chief of Engineers DA, Washington 25, DC Attn: ENGEB	1
Chief of Engineers DA, Washington 25, DC Attn: ENGTE	1
Chief of Ordnance, DA, Washington 25, DC Attn: ORDTN	2
Chief Signal Officer, DA, Comb Dev & Ops Div Washington 25, DC Attn: SIGCO-4	1
Chief of Transportation, DA, Office of Planning & Intel. Washington 25, DC	1
The Surgeon General, DA, Washington 25, DC Attn: MEDNE	2
Commander-in-Chief, U.S. Army Europe, APO 403, New York, N.Y. Attn: OPOT Div, Weapons Branch	1

Commanding General U.S. Continental Army Command Ft. Monroe, Va.	3
Director of Special Weapons Development Office, HQ CONARC, Ft. Bliss, Texas Attn: Capt Chester I. Peterson	1
President U.S. Army Artillery Board Ft. Sill, Okla	1
President U.S. Army Aviation Board Ft. Rucker, Alabama Attn: ATBG-DG	1
Commandant U.S. Army C&GS College Ft. Leavenworth, Kansas Attn: Archives	1
Commandant U.S. Army Air Defense School Ft. Bliss, Texas Attn: Command & Staff Dept	1
Commandant U.S. Army Armored School Ft. Knox, Kentucky	1
Commandant U.S. Army Arty & Missile Sch Ft. Sill, Oklahoma Attn: Combat Dev Dept	1
Commandant U.S. Army Infantry School Ft. Benning, Ga. Attn: C.D.S.	1
Commandant Quartermaster School, US Army Ft. Lee, Va. Attn: Ch, QM Library	1
Commanding General Chemical Corps Training Comd Ft. McClellan, Ala.	1

Commandant US Army Chemical Corps CBR Weapons School Dugway Proving Ground Dugway, Utah	1
Commandant US Army Signal School Ft. Monmouth, N. J.	1
Commandant US Army Transport School Ft. Eustis, Va. Attn: Security & Info Off.	1
Commanding General The Engineer Center Ft. Belvoir, Va. Attn: Asst. Cmdt Engr School	1
Commanding General Army Medical Service School Brooke Army Medical Center Ft. Sam Houston, Texas	1
Commanding Officer 9th Hospital Center APO 180, New York, N.Y. Attn: CO, US Army Nuclear Medicine Research Det, Europe	1
Director Armed Forces Institute of Path. Walter Reed Army Med. Center 625 16th St. NW Washington 25, D.C.	1
Commanding Officer Army Medical Research Lab. Ft. Knox, Ky	1
Commandant, Walter Reed Army Inst of Res. Walter Reed Army Med Center Washington 25, D.C.	1
Commanding General QM R&D Comd, QM R&D Center Natick, Mass. Attn: CBR Liaison Officer	2

Commanding General QM Research & Engr. Comd, USA Natick, Mass (For reports from Opn HARDTACK only)	1
Commanding General US Army Chemical Corps Research & Development Comd. Washington 25, DC	2
Commanding Officer Chemical Warfare Lab Army Chemical Center, Md. Attn: Tech Library	2
Commanding General Engineer Research & Dev Lab Ft. Belvoir, Va. Attn: Ch, Tech Support Branch	1
Director Waterways Experiment Station PO Box 631 Vicksburg, Miss. Attn: Library	1
Commanding General Aberdeen Proving Ground Aberdeen Proving Ground, Md. Attn: Ballistic Research Lab, Dir. BRL	2
Commander Army Ballistic Missile Agency Redstone Arsenal, Alabama Attn: ORDAB-HT	1
Commanding General US Army Electronic Proving Ground Ft. Huachuca, Arizona Attn: Tech Library	1
Director Operations Research Office Johns Hopkins University 6935 Arlington Road Bethesda 14, Md.	1

DISTRIBUTION

NAVY AGENCIES

Chief of Naval Operations D/N, Washington 25, D.C. ATTN: OP03EG	1
Chief of Naval Operations D/N, Washington 25, D.C. ATTN: OP-75	1
Chief of Naval Operations D/N, Washington 25, D.C. ATTN: OP-922G2	1
Chief of Naval Operations D/N, Washington 25, D.C. ATTN: OP-91	1
Chief of Naval Personnel D/n, Washington 25, D.C.	1
Chief of Naval Research D/N, Washington 25, D.C. ATTN: Code 811	2
Chief Bureau of Medicine & Surgery D/N, Washington 25, D.C. ATTN: Special Wpns Def Div	1
Chief, Bureau of Ships D/N, Washington 25, D.C. ATTN: Code 423	1
Chief Bureau of Yards & Docks D/N, Washington 25, D.C. ATTN: D-440	1
Director U.S. Naval Research Laboratory Washington 25, D.C. ATTN: Mrs. Katherine H. Cass	1
Commander U.S. Naval Ordnance Lab White Oak, Silver Spring, Maryland	2
Director Material Laboratory (Code 900) New York Naval Shipyard Brooklyn 1, N.Y.	1

Commanding Officer
 U.S. Naval Mine Defense Lab
 Panama City, Fla 1

Commanding Officer
 U.S. Naval Radiological Defense
 Laboratory, San Francisco
 California, ATTN: Tech Info Div 4

Commanding Officer & Director
 U.S. Naval Civil Engineering
 Lab., Port Hueneme, California
 ATTN: Code L31 1

Commanding Officer, U.S. Naval
 Schools Command, U.S. Naval
 Station, Treasure Island,
 San Francisco, California 1

Superintendent
 U.S. Naval Postgraduate School
 Monterey, California 1

Commanding Officer, Nuclear
 Weapons Training Center,
 Atlantic, U.S. Naval Base,
 Norfolk 11, Va., ATTN:
 Nuclear Warfare Dept 1

Commanding Officer, Nuclear
 Weapons Training Center,
 Pacific, Naval Station,
 San Diego, California 1

Commanding Officer, U.S. Naval
 Damage Control Tng Center,
 Naval Base, Philadelphia 12, Pa
 ATTN: ABC Defense Course 1

Commanding Officer
 U.S. Naval Air Development
 Center, Johnsville, Pa
 ATTN: NAS, Librarian 1

Commanding Officer, U.S. Naval
 Medical Research Institute,
 National Naval Medical Center,
 Bethesda, Maryland 1

Officer in Charge, U.S. Naval
Supply Research & Development
Facility, Naval Supply Center,
Bayonne, New Jersey 1

Commandant
U.S. Marine Corps
Washington 25, D.C.
ATTN: Code AO3H 1

DISTRIBUTION

AIR FORCE AGENCIES

Air Force Technical Application
Center, Hq USAF,
Washington 25, D.C. 1

Hq USAF, ATTN: Operations
Analysis Office, Vice
Chief of Staff,
Washington 25, D.C. 1

Air Force Intelligence Center
Hq USAF, ACS/1 (AFOIN-3VI)
Washington 25, D.C. 2

Assistant Chief of Staff
Intelligence, HQ USAFE, APO
633, New York, N.Y. ATTN:
Directorate of Air Targets 1

Director of Research &
Development, DCS/D, Hq USAF,
Washington 25, D.C.
ATTN: Guidance & Weapons
Division 1

Commander
Tactical Air Command
Langley AFB, Virginia
ATTN: Doc Security Branch 1

The Surgeon General
Hq USAF, Washington 25, D.C.
ATTN: Bio-Def Pre Med Div 1

Commander Tactical Air Command Langley AFB, Virginia ATTN: Doc Security Branch	1
Commander Air Defense Command Ent AFB, Colorado, ATTN: Assistant for Atomic Energy, ADLDC-A	1
Commander, HQ Air Research & Development Command, Andrews AFB, Washington 25, D.C. ATTN: RDRWA	1
Commander, Air Force Ballistic Missile Division Hq ARDC, Air Force Unit Post Office, Los Angeles 45, California ATTN: WDSOT	1
Commander-in-Chief, Pacific Air Forces, APO 953, San Francisco, California, ATTN: PFCIE-MB, Base Recovery	1
Commander, AF Cambridge Research Center, L.G. Hanscom Field, Bedford, Massachusetts, ATTN: CRQST-2	2
Commander, Air Force Special Weapons Center, Kirtland AFB, Albuquerque, New Mexico, ATTN: Tech Info & Intel Div	5
Directory Air University Library Maxwell AFB, Alabama	2
Commander Lowry AFB, Denver, Colorado Attn: Dept of Sp Wpns Tng	1
Commandant, School of Aviation Medicine, USAF Aerospace Med- ical Center (ATC) Brooks AFB Tex ATTN: Col Gerrit L. Hekhuis	2

Commander 1009th Sp Wpns Squadron Hq USAF, Washington 25, D.C.	1
Commander Wright Air Development Center Wright-Patterson AFB, Ohio ATTN: WCOSI	2
Director, USAF Project Rand, VIA:US Air Force Liaison Office The Rand Corporation, 1700 Main Street, Santa Monica, California	2
Commander, Air Defense Systems Integration Division, L.G. Hanscom Field, Bedford, Mass ATTN: SIDE-S	1
Commander, Air Technical Intell- igence Center, USAF, Wright- Patterson Air Force Base, Ohio ATTN: AFCIN-4Bla, Library	1
DISTRIBUTION	
OTHER AGENCIES	
Director of Defense Research and Engineering, Washington 25, D.C. ATTN: Tech Library	1
Director, Weapons Systems Evaluation Group, Room IE880 The Pentagon Washington 25, D.C.	1
U.S. Documents Officer Office of the United States National Military Representa- tive-SHAPE APO 55, NY., N.Y.	1
Chief Defense Atomic Support Agency Washington 25, D.C. ATTN: Document Library *Reduce to 3 cys for all FWE reports	4*

Commander, Field Command DASA, Sandia Base, Albuquerque, New Mexico	1
Commander, Field Command DASA, Sandia Base Albuquerque, New Mexico ATTN: FCTG	1
Commander, Field Command DASA, Sandia Base Albuquerque, New Mexico ATTN: FCWT	2
Administrator, National Aeronautics & Space Adminis- tration, 1520 "H" Street N.W. Washington 25, D.C., ATTN: Mr. R.V. Rhode	1
Commander-in-Chief Strategic Air Command Offutt AFB, Nebraska ATTN: OAWS	1
Commandant U.S. Coast Guard 1300 E. Street, NW Washington 25, D.C. ATTN: (OIN)	1
Staff Medical Officer British Naval Staff Room 4922 Main Navy Bldg. Washington 25, D.C.	1
SPECIAL DISTRIBUTION	
U.S. Atomic Energy Commission Washington 25, D.C. ATTN: Chief, Civil Effects Branch Division of Biology and Medicine	450
Aberdeen Proving Ground, Md, Ballistic Research Laboratories Terminal Ballistics Attn: Mr. Robert U. Clark, Physicist Mr. William J. Taylor, Physicist	2

Airborne Instruments Laboratory
 Department of Medicine and Biological Physics
 Deer Park, Long Island, New York
 Attn: Mr. W. J. Carberry 1

Air Force Special Weapons Center
 Kirtland Air Force Base
 Albuquerque, N.M
 Attn: Mr. R. R. Birukoff, Research Engineer 1

Air Research & Development Command Hqs.
 Andrews Air Force Base
 Washington 25, D.C.
 Attn: Brig. Gen. Benjamin Strickland
 Deputy Director of Life Sciences 1

AiResearch Manufacturing Company
 9851-9951 Sepulveda Blvd.
 Los Angeles 25, California
 Attn: Mr. Frederick H. Green, Assistant Chief,
 Preliminary Design
 Dr. James N. Waggoner, Medical Director 2

AeResearch Manufacturing Company
 Sky Harbor Airport
 402 South 36th Street
 Phoenix, Arizona
 Attn: Delano Debaryshe
 Leighton S. King 2

American Airlines, Inc.
 Medical Services Division
 La Guardia Airport Station
 Flushing 71, N.Y.
 Attn: Dr. Kenneth L. Stratton, Medical Director 1

Brooks Air Force Base
 United States Air Force Aerospace Medical Center (ATC)
 School of Aviation Medicine
 Brooks Air Force Base, Texas
 Attn: Brig. Gen. Theodore C. Bedwell, Jr., Commandant
 Col. Paul A. Campbell, Chief, Space Medicine
 Dr. Hubertus Strughold, Advisor for Research &
 Professor of Space Medicine 3

The Boeing Company 3
P. O. Box 3707
Seattle 24, Washington
Attn: Dr. Thrift G. Hanks, Director of Health & Safety
Dr. Romney H. Lowry, Manager, Space Medicine Branch

Dr. F. Werner, Jr., Space Medicine Section
P.O. Box 3915

Chance Vought Astronautics 5
P. O. Box 5907
Dallas 22, Texas
Attn: Dr. Charles F. Gell, chief Life Sciences
Dr. Lathan
Mr. Ramon McKinney, Life Sciences Section
Mr. C. O. Miller
Mr. A. I. Sibila, Manager Space Sciences

Chemical Corps Research & Development Command 2
Chemical Research & Development Laboratories
Army Chemical Center, Md.
Attn: Dr. Fred W. Stemler
Dr. R. S. Anderson

Civil Aeromedical Research Institute 1
Oklahoma City, Oklahoma
Attn: Director of Research

Professor Carl-Johan Clemenson 1
Department of Hygiene
University of Göteborg
Fjärde Langgatan 7B,
Göteborg, Sweden

Convair Division, General Dynamics Corp. 2
Fort Worth, Texas
Attn: Mr. H. A. Bodely
Mr. Schreiber

Convair - General Dynamics Corporation 8
Mail Zone 1-713
P. O. Box 1950
San Diego 12, California
Attn: Dr. R. C. Armstrong, Chief Physician
Dr. J. C. Clark, Assistant To Vice-President Engineering
Mr. James Dempsey
Dr. L. L. Lowry, Chief Staff Systems Evaluation Group
Mr. M. H. Thiel, Design Specialist

Dr. R. A. Nau (Mail Zone 6-104)

Mr. W. F. Rector, III (Mail Zone 580-40), P.O. Box 1128

Mr. R. C. Sebold, Vice-President Engineering
Convair General Offices

Defense Atomic Support Agency Department of Defense Field Command Sandia Base, New Mexico Attn: Col. S. W. Cavender, Surgeon	1
Department of National Defence Defence Research Board Suffield Experimental Station Ralston, Alberta, Canada Attn: Mr. M. K. McPhail, Head of Physiology Section Mr. E. G. Brigden	2
The Dikewood Corporation 4805 Menaul Blvd., N.E. Albuquerque, New Mexico	1
Douglas Aircraft Company, Inc. El Segundo Division El Segundo, California Attn: Mr. Harvey Glassner Dr. E. B. Konecni	2
Federal Aviation Agency Washington 25, D.C. Attn: Dr. James L. Goddard, Civil Air Surgeon	1
Goodyear Aircraft Corporation Department 475, Plant H 1210 Massillon Road Akron 15, Ohio Attn: Dr. A. J. Cacioppo	1
Harvard School of Public Health Harvard University 695 Huntington Avenue Boston 15, Mass. Attn: Dr. Ross A. McFarland, Associate Professor of Industrial Hygiene	1
Mr. Kenneth Kaplan Physicist Broadview Research Corporation 1811 Trousdale Drive Burlingame, Calif.	1
Lockheed Aircraft Company Suite 302, First National Bank Bldg. Burbank, California Attn: Dr. Charles Barron	1

Lockheed Aircraft Corporation
Lockheed Missile and Space Division
Space Physics Department (53-23)
Sunnyvale, California
Attn: Dr. W. Kellogg, Scientific Assistant to
Director of Research
Dr. Heinrich Rose

Lockheed Aircraft Corporation
1122 Jagels Road
Palo Alto, California
Attn: Dr. L. Eugene Root, Missile Systems Director

Lovelace Foundation for Medical Education and Research
4800 Gibson Blvd., SE
Albuquerque, N.M.
Attn: Dr. Clayton S. White, Director of Research

The Martin Company
Denver, Colorado
Attn: Dr. James G. Gaume, Chief, Space Medicine

McDonnell Aircraft Company
Lambert Field
St. Louis, Missouri
Attn: Mr. Henry F. Creel, Chief Airborne Equipment
Systems Engineer
Mr. Bert North

National Aeronautics and Space Administration
1520 "H" Street, N.W.
Washington 25, D.C.
Attn: Brig. Gen. Charles H. Roadman, Acting Director,
Life Sciences Program

Naval Medical Research Institute
Bethesda, Md.
Attn: Dr. David E. Goldman, MSC, Commander

Department of the Navy
Bureau of Medicine & Surgery
Washington 25, D.C.
Attn: Capt. G. J. Duffner, Director, Submarine Medical Division

Dr. Arne Nelson
Research Institute of National Defense
Sundbyberg 4, Sweden

4
North American Aviation
International Airport
Los Angeles 45, Calif.
Attn: Scott Crossfield
Dr. Toby Freedman, Flight Surgeon
Mr. Fred A. Payne, Manager Space Planning,
Development Planning
Mr. Harrison A. Storms

1
Office of the Director of Defense Research & Engineering
Pentagon
Washington 25, D.C.
Attn: Col. John M. Talbot, Chief, Medical Services Division,
Room 3D1050 Office of Science

2
The Ohio State University
410 West 10th Avenue
Columbus 10, Ohio
Attn: Dr. William F. Ashe, Chairman, Department of
Preventive Medicine
Dean Richard L. Meiling

2
The RAND Corporation
1700 Main Street
Santa Monica, Calif.
Attn: Dr. H. H. Mitchell, Physics Division
Dr. Harold L. Brode

3
Republic Aviation Corporation
Applied Research & Development
Farmingdale, Long Island, N.Y.
Attn: Dr. Alden R. Crawford, Vice-President
Life Sciences Division
Dr. William H. Helvey, Chief, Life Sciences Division
Dr. William J. O'Donnell, Life Sciences Division

5
Sandia Corporation
P. O. Box 5800
Albuquerque, New Mexico
Attn: Dr. C. F. Quate, Director of Research
Dr. S. P. Bliss, Medical Director
Dr. T. B. Cook, Manager, Department 5110
Dr. M. L. Merritt, Manager, Department 5130
Mr. L. J. Vortman, 5112

1
System Development Corporation
Santa Monica, California
Attn: Dr. C. J. Roach

United Aircraft Company Denver, Colorado Attn: Dr. George J. Kidera, Medical Director	1
Laboratory of Nuclear Medicine & Radiation Biology School of Medicine University of California, Los Angeles 900 Veteran Avenue Los Angeles 24, California Attn: Dr. G. M. McDonnel, Associate Professor Dr. Benedict Cassen	2
University of Illinois Chicago Professional Colleges 840 Wood Street Chicago 12, Illinois Attn: Dr. John P. Marbarger, Director, Aeromedical Laboratory	1
University of Kentucky School of Medicine Lexington, Kentucky Attn: Dr. Loren D. Carlson, Professor of Physiology & Biophysics	1
University of New Mexico Albuquerque, New Mexico Attn: Library	1
U. S. Naval Ordnance Laboratory White Oak, Maryland Attn: Capt. Richard H. Lee, MSC Mr. James F. Moulton	2
U. S. Naval School of Aviation Medicine U. S. Naval Aviation Medical Center Pensacola, Florida Attn: Capt. Ashton Graybiel, Director of Research	1
Dr. Shields Warren Cancer Research Institute New England Deaconess Hospital 194 Pilgrim Road Boston 15, Mass.	1
Wright Air Development Center Aeromedical Laboratory Wright-Patterson Air Force Base, Ohio Attn: Commanding Officer Dr. Henning E. vonGierke, Chief, Bioacoustics Laboratory	2

Sir Solly Zucker 1
6, Carpenter Roman
Edgbaston Road,
Birmingham 15, England

Dr. Eugene Zwoyer 1
Director, Shock Tube Laboratory
P. O. Box 188
University Station
Albuquerque, New Mexico

Armed Services Technical Information Agency 20
Arlington Hall Station
Arlington 12, Virginia



Defense Threat Reduction Agency

45045 Aviation Drive
Dulles, VA 20166-7517

CPWC/TRC

May 6, 1999

MEMORANDUM FOR DEFENSE TECHNICAL INFORMATION CENTER
ATTN: OCQ/MR WILLIAM BUSH

SUBJECT: DOCUMENT REVIEW

The Defense Threat Reduction Agency's Security Office
has reviewed and declassified or assigned a new
distribution statement:

- AFSWP-1069, AD-341090, STATEMENT A - *RD waiting for reply*
- DASA-1151, AD-227900, STATEMENT A
- DASA-1355-1, AD-336443, STATEMENT A
- DASA-1298, AD-285252, STATEMENT A ✓
- DASA-1290, AD-444208, STATEMENT A ✓
- DASA-1271, AD-276892, STATEMENT A ✓
- DASA-1279, AD-281597, STATEMENT A
- DASA-1237, AD-272653, STATEMENT A
- DASA-1246, AD-279670, STATEMENT A
- DASA-1245, AD-419911, STATEMENT A
- DASA-1242, AD-279671, STATEMENT A
- DASA-1256, AD-280809, STATEMENT A
- DASA-1221, AD-243886, STATEMENT A - *FRD*
- DASA-1390, AD-340311, STATEMENT A
- DASA-1283, AD-717097, STATEMENT A *OK*
- DASA-1285-5, AD-443589, STATEMENT A
- DASA-1714, AD-473132, STATEMENT A
- DASA-2214, AD-854912, STATEMENT A
- DASA-2627, AD-514934, STATEMENT A
- DASA-2651, AD-514615, STATEMENT A
- ~~DASA-2536, AD-876697, STATEMENT A~~
- DASA-2722T-V3, AD-518506, STATEMENT A
- DNA-3042F, AD-525631, STATEMENT A
- DNA-2821Z-1, AD-522555, STATEMENT A

If you have any questions, please call me at 703-325-1034.

Ardith Jarrett

ARDITH JARRETT

Chief, Technical Resource Center