

AWARD NUMBER: W81XWH-15-1-0353

TITLE: microRNA Biomarkers to Generate Sensitivity to Abiraterone-Resistant Prostate Cancer

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REPORT DATE: September 2016

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
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REPORT DOCUMENTATION PAGE

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1. REPORT DATE September		2. REPORT TYPE Annual		3. DATES COVERED 15 Aug 2015 - 14 Aug 2016	
4. TITLE AND SUBTITLE microRNA Biomarkers to Generate Sensitivity to Abiraterone-Resistant Prostate Cancer				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-15-1-0353	
6. AUTHOR(S) Pheruza Tarapore, Sarah To, Dan Song, Shuk-mei Ho E-Mail: Pheruza.Tarapore@uc.edu				5c. PROGRAM ELEMENT NUMBER	
				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Cincinnati 2600 CLIFTON AVE CINCINNATI OH 45220-2872				5f. WORK UNIT NUMBER	
				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT We plan to develop a combination therapeutic approach, employing Abiraterone (Abi) plus RNA therapy. For this, we will use an aptamer specific for PSMA (aptPSMA) to specifically target CRPC cells. The affinity and high specificity of aptPSMA for binding human CRPC cells expressing PSMA has already been reported, as has its utility as a drug delivery system for siRNAs. However, it has not been used to deliver pre-miRNA to cells. Identification of Abi-R markers is important for designing therapeutic interventions sensitizing PCas to combination therapies and for prognostic applications to monitor and predict for disease relapse (Abi-R). Additionally, we propose to use patient derived PCa xenograft animal model (PCa-PDX mice) to identify differentially expressed microRNA (miRNA) on castration and Abi dependent tumor regression followed by regrowth/relapse. <i>Our central hypothesis is that changes in miRNA expression underlie Abi-R mechanisms and that PCa-PDX mice will be excellent surrogates to identify markers for Abi-R. We further postulate that RNA therapy (restoring or targeting miRNA) should increase sensitivity of Abi-R tumors, allowing us to prolong treatments, and hence the life of a patient.</i> Our Aims: (1) To develop RNA aptamer therapy. We will test 8 of the recently identified Abi regulated miRNAs for therapeutic utility in vitro. We will design an aptPSMA-pre-miRNA therapeutic delivery vehicle for CRPC-tissue specific delivery. The best miRNA will be used for in vivo studies (2) To generate Abi-R PDX mice and identify the differentially expressed miRNA.					
15. SUBJECT TERMS					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT UU	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT	b. ABSTRACT	c. THIS PAGE			USAMRMC
Unclassified	Unclassified	Unclassified	Unclassified	12	19b. TELEPHONE NUMBER (include area code)

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1. INTRODUCTION:

Prostate cancer (**PCa**) is the most frequent cancer occurring in men in the United States. While screening for elevated levels of prostate-specific antigen (PSA) has dramatically improved early detection of this disease (Cooperberg et al., 2004, Etzioni et al., 2002), PCa is still the second leading cause of cancer deaths in men. In 2014, it is estimated that 233,000 men will be diagnosed with and 29,480 men will die of cancer of the prostate (American Cancer Society, 2013). Of concern, a substantial proportion of patients develop an incurable disseminated disease after local therapy, even when the primary lesion was localized to the prostate when first diagnosed (Pendleton et al., 2007). Also, a large number of advanced PCas become androgen-independent, for which there is no known cure. Indeed, median survival for patients with metastatic hormone refractory PCa is still around 18 months (Hadaschik et al., 2007, Hadaschik and Gleave, 2007). Thus, there is an urgent need to come up with new treatment regimens for these patients.

In this proposal, we sought to develop a combination therapeutic approach to treat PCa, employing Abiraterone plus RNA therapy, as a promising methodological approach. A candidate cell surface receptor is PSMA (prostate specific membrane antigen). PSMA expression is associated with higher Gleason scores in tumors, and with increased aneuploidy. It is currently used as a tumor marker for diagnosis, monitoring and prognosis of prostatic carcinoma. Elevated levels of PSMA is also used as an independent marker for predicting disease relapse. We will use an RNA aptamer which binds specifically to PCa cells to deliver the miRNA. **miRNA** have an advantage over siRNA for gene silencing since they can target multiple components of the cellular networks / signaling pathways responsible for advanced disease progression.

We had identified miRNA induced by Abi treatment of castration resistant prostate cancer C4-2 cells using TaqMan Array Human MicroRNA Card Set v3.0 (Life Technologies). The top 7 candidates are shown in **Table 1**. From our literature search and data mining to determine known and potential gene targets for miRNA, we determined that most of these miRNA appear to be tumor suppressors (**Table 1**), inhibiting the expression of oncogenes, pro-angiogenic factors, or genes involved in cell proliferation, migration, or metastasis. The expression pattern of *miR-487a-3p*, *miR-492-5p*, *miR-510-5p* and *miR-623-5p* especially suggest that these miRNAs maybe involved in Abi-R (**Table1**, contrast the gene function with expression levels). Using mimics and inhibitors for each miRNA, we will determine which of the identified miRNA are best suited for further studies.

Since Abi targets androgen biosynthesis in the adrenals, testis, as well as has direct effects on the tumor tissues, an *in vitro* approach to identify the biomarkers of resistance to Abi can be imprecise. Our results and others (Kosaka et al., 2014) have shown that while Abi inhibits CYP17A1 activities and expression in C4-2 cells, it is not very effective in reducing cell proliferation *in vitro* at physiological levels of <14 μ M. However, it is most effective at suppressing cancer properties *in vivo* in mouse xenograft models (Bruno et al., 2011, Mostaghel et al., 2011) and in patients with CRPC (Mostaghel, 2014), indicating that system-wide androgen suppression and inhibition of AR axis is important. **Patient-derived xenograft (PDX)** tumor models retain much of the biological diversity, heterogeneity, molecular characteristics and tissue architecture of the original patient tumor. Recent advances in the development of these models and their increasing sophistication have led to their escalating use for anticancer drug research and development, and as predictive clinical models. *Hence, for the correct identification of Abi-R*

markers, we propose using PCa-PDX animal model (PDX mice, Jackson Labs). The identification of abi-R markers is important for designing therapeutic interventions sensitizing PCas to combination PCa therapies and for prognostic applications to monitor and predict for disease relapse (Abi-R).

Hypothesis: *We hypothesize that changes in miRNA expression underlie Abi resistance (Abi-R) mechanisms. We further postulate that a RNA therapy (restoring or targeting miRNA) should increase sensitivity of Abi-R tumors (Fig. 2), allowing us to prolong treatments (Table 2), and hence the life of a patient.*

2. KEYWORDS: RNA aptamer, PSMA, microRNA

3. ACCOMPLISHMENTS:

Table 1. Summary of miRNAseq data on Abi-treated C4-2 Cells		
Fold change	Functions	Target genes involved
Possible involvement in Abi-induced tumor regression		
hsa-miR-573-5p	4.8 Tumor suppressor	<i>Known Target:</i> Genes involved in cell adhesion (MCAM/ CD146/muc18). <i>Potential Targets :</i> SPARCL1, SLC25A26
hsa-miR-202-3p	4.2 Tumor suppressor	<i>Known Target:</i> Genes involved in tumor progression, epithelial–mesenchymal transition (EMT), bone metastasis (Gli2, LRP6, CALD1, GHR) <i>Potential Targets:</i> ARID3B, DICER1, MYCN, PPARGC1B, HMGA2
hsa-miR-1262-5p	1.5 ?? Tumor suppressor	<i>Potential Targets :</i> mRNA splicing GEMIN5, ECM/proliferation: FGFR1, OCM2, ITGAV, MYB
Possible involvement in Abi-Resistance		
hsa-miR-487a-3p	0.45 Tumor suppressor	<i>Known Target:</i> Genes involved in mitoxantrone (MX) resistance (BCRP/ABCG2). <i>Potential targets:</i> SP1, SP3 which upregulate PSA, AR, alpha integrin.
hsa-miR-492-5p	0.65 Anti-angiogenic	<i>Known Target:</i> Genes involved in angiogenesis (restin, MCL1, DUSP3, BRAF, MAP3K1, MMP10, SP1) <i>Potential targets :</i> ECM remodelling/metastasis CCBE1, CD44, CLDN19
hsa-miR-510-5p	5.4 Oncomir	<i>Known Target:</i> Genes involved in inhibition of cancer progression/invasion PRDX1, prostate-derived Ets factor PDEF <i>Potential Targets :</i> SPOCK1, RNA export THOC2
hsa-miR-623-5p	0.03 ?? Tumor suppressor	<i>Potential Targets:</i> oncogene SKI, cell cycle regulator CCND2

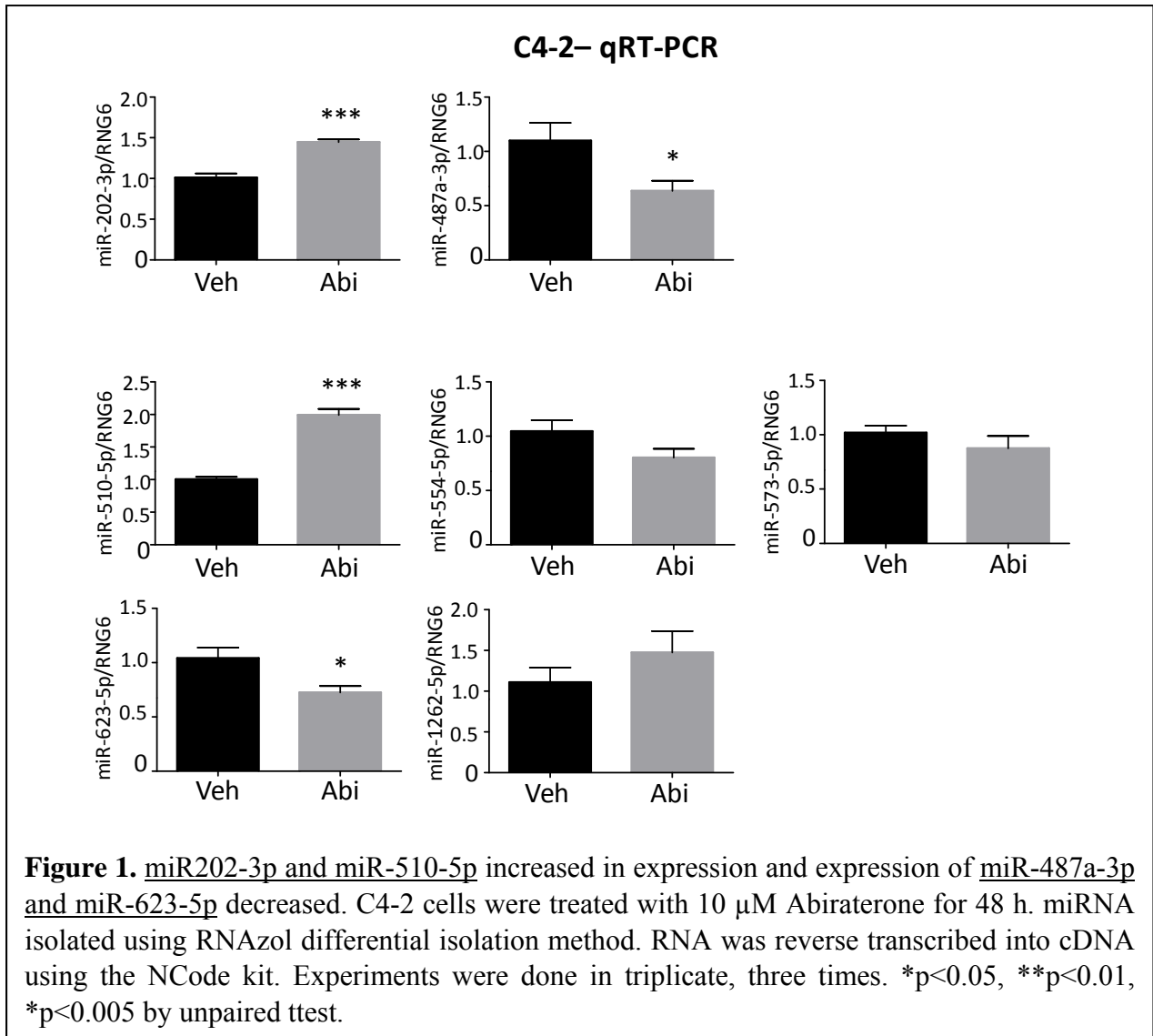
Specific Aims:

1. **To develop RNA aptamer therapy.** We will test 7 of the recently identified Abi regulated miRNAs (Table 1) for therapeutic utility *in vitro*. The best miRNA will be used for *in vivo* studies for inducing sensitivity to Abi, using the C4-2 and /or 22Rv1 CRPC cells which express PSMA. We will also identify (for miRNA), their downstream targets and pathways in CRPC cells to gain insights regarding the mechanism of resistance.

Task 1. Determine which miRNA (amongst miRNA miR202-3p, miR-487a-3p, miR-492-5p, miR-510-5p, miR-554-5p, miR-573-5p, miR-623-5p, miR-1262-5p) is best suited for PCa therapy. (months 1-9).

We had identified miRNA induced by Abi treatment of C4-2 cells using TaqMan Array Human MicroRNA Card Set v3.0 (Life Technologies). We first validated that expression of these

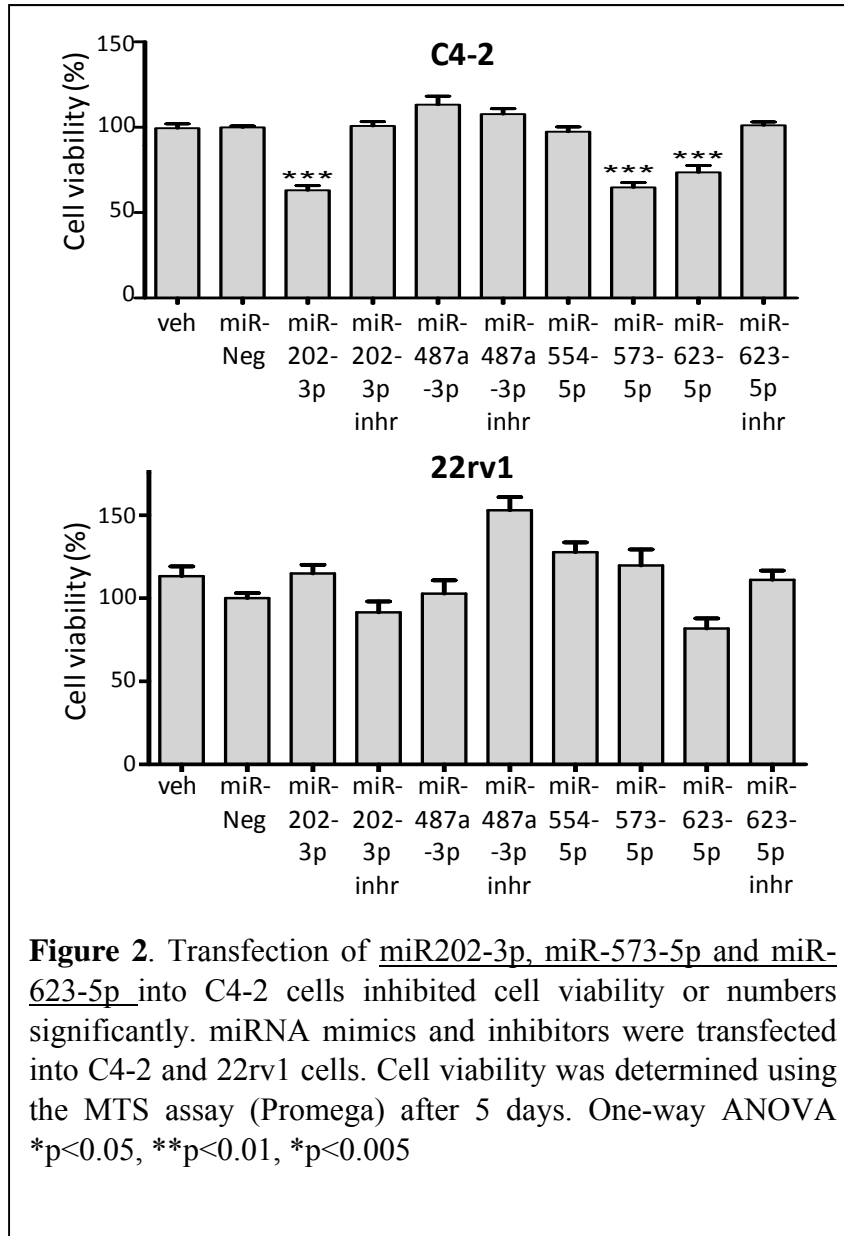
miRNA was significantly changed by qPCR (**Figure 1**). We found that the expression of 4 of the



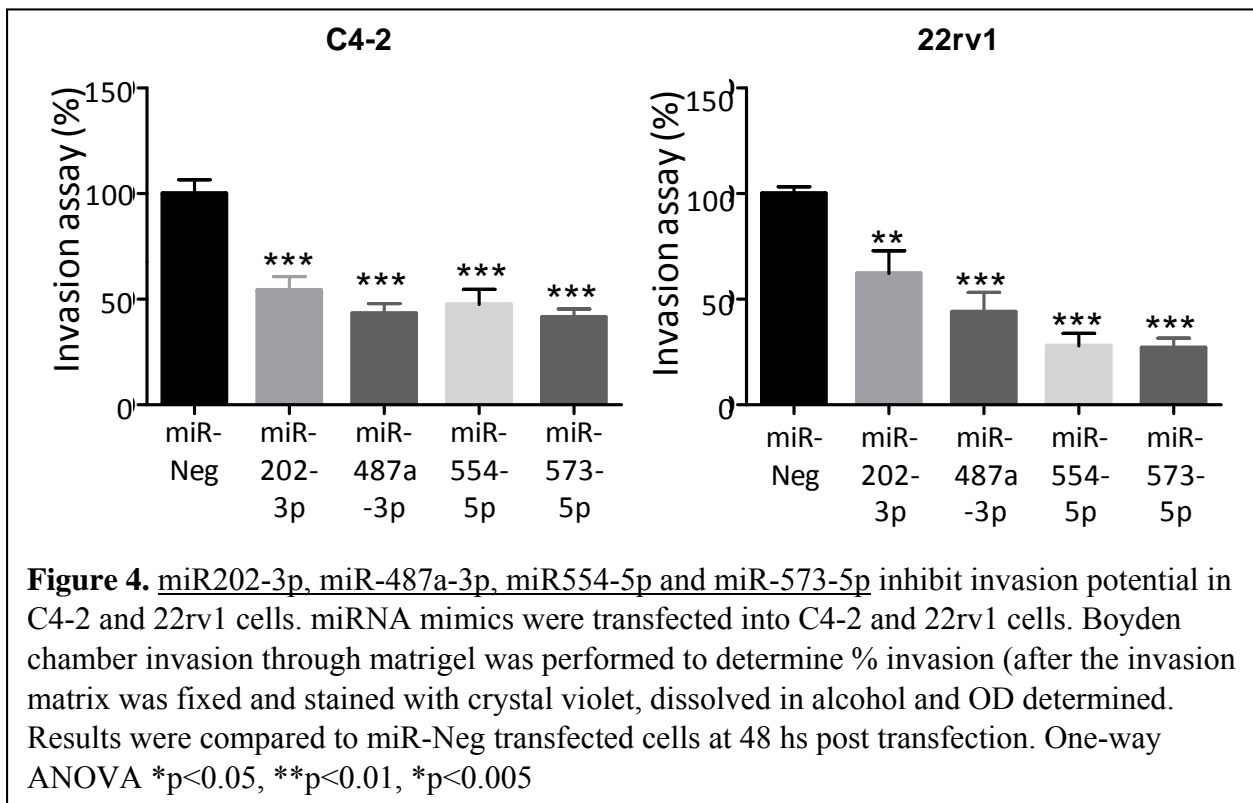
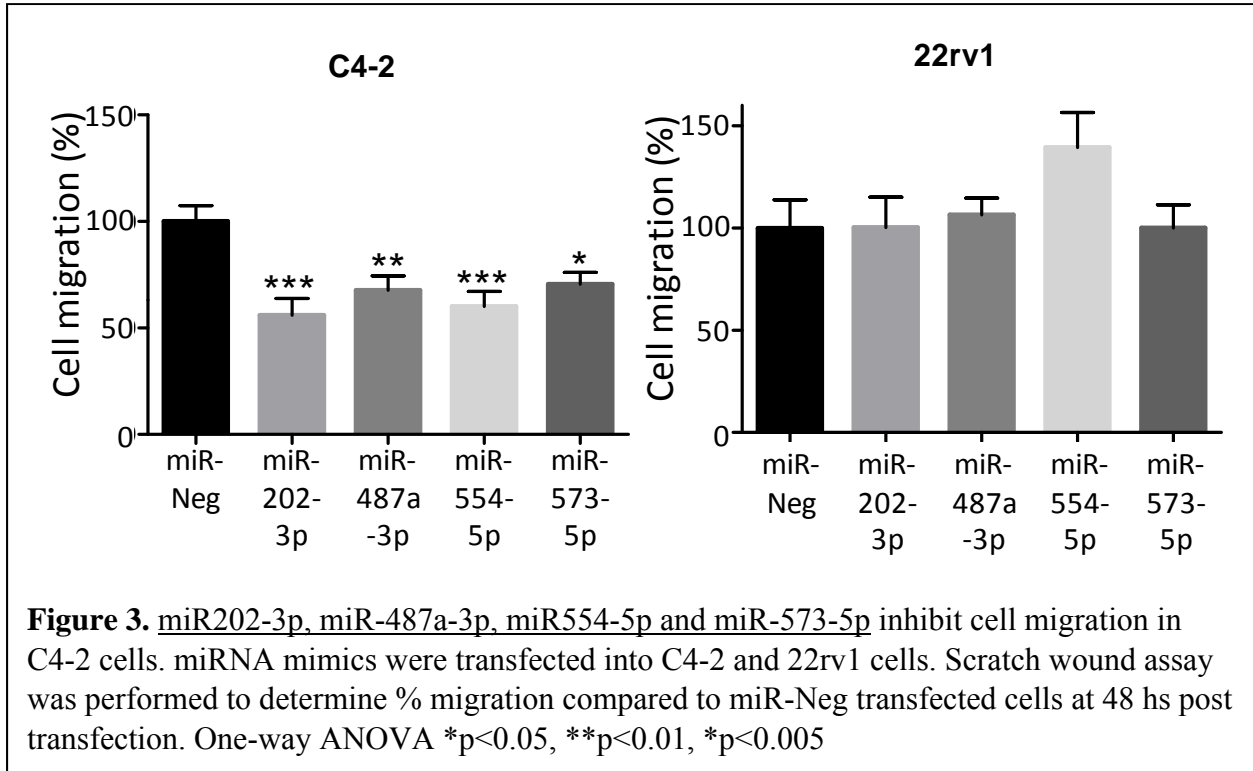
miRNA changed; miR202-3p and miR-510-5p increased in expression and expression of miR-487a-3p and miR-623-5p decreased. We could not get good signals for miR-492-5p to quantitate it.

At the same time, we ordered the miRNA mimics and inhibitors. These were transfected into two castration resistant prostate cancer (CRPC) celllines, C4-2 and 22rv1.

(1) We found that while transfection of miR202-3p, miR-573-5p and miR-623-5p into C4-2 cells inhibited cell growth (MTS assay, (**Figure 2**)), the same was not true for 22rv1 cells. While significance was not reached, miR-623-5p did decrease cell viability in 22rv1 cells.



(2) miR202-3p, miR-487a-3p, miR554-5p and miR-573-5p inhibit cell migration in C4-2 cells (scratch wound assay) but not in 22rv1 cells (**Figure 3**).



(3) However, when we examined the invasive potential of these miRNA transfected cells, we find that four miRNA (miR202-3p, miR-487a-3p, miR554-5p and miR-573-5p) decreased the invasive potential as determined using the Boyden chamber matrix assay (**Figure 4**).

Since miR202-3p, miR-573-5p and miR623-5p had effects on cell viability, we proceed with these 3 miRNA.

Our results with miR623-5p is a recent finding in 22rv1, and we still need to perform migration and invasion assays with this construct.

Task 3. To identify function of miRNA: Determining the network of miRNA-regulated genes (Months 1-12)

This part is currently ongoing. We have transfected C4-2 and PC-3 cells with pCMV-Ago, and are screening for protein levels. We anticipate that we should have results by the next annual report.

Task 4. Design functionally active aptamer-pre-miRNA and aptamer-mature-miRNA chimeric molecules (aptPSMA-iRNA) (Months 7-12).

We have designed and made the T7 transcribed miR202-3p-24MERreverse (two separate designs will be tested), miR-573-5p-24MERreverse, miR-623-5p-24MERreverse and aptPSMA-24MER. They have been annealed to give PSMA-Aptamer-iMT (aptPSMA-iRNA, **Figure 5**). Further testing for correct processing, resistance to degradation, and cleavage activity is being done.

Specific Aim 2: To generate Abi-resistant (Abi-R) PCa Patient-derived xenograft (PCa-PDX) mice and identify the differentially expressed miRNA.

Task 2. Order and castrate the 32 patient derived engrafted (32 PCa-PDX mice - Jackson labs) male mice

We have the DoD animal protocol in place and approved. We are planning on ordering the PDX-mice within the next month to perform this part of the experiment. We have talked to Jackson labs and they have the PCa PDX mice ready to ship. We have the protocol in place at the DoD to initialize these studies. Due to distribution of work resulting in personnel issues, we had a slight delay in initiating this part, but are now ready to proceed (Zhou et al., 2016).

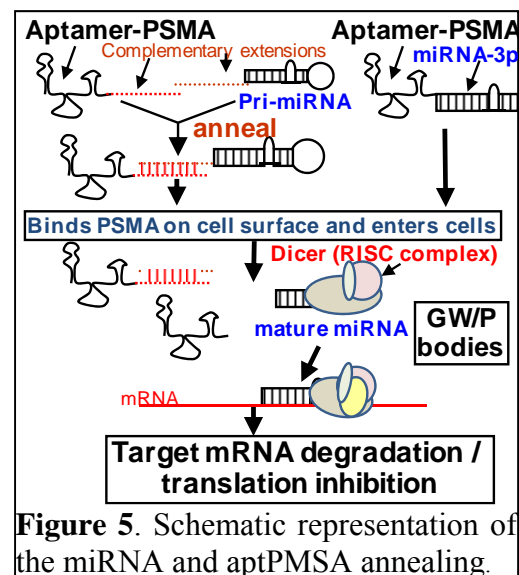


Figure 5. Schematic representation of the miRNA and aptPSMA annealing.

Plan for the next reporting period: We have high hope that we will be able to catch up with all our studies as outlined in the SOW and attain our goals in the near future. During the next reporting

period, we will test out our aptamer-miRNA constructs in C4-2 and 22rv1 cells in presence and absence of Abi for cell viability, motility and invasiveness. We will then use the xenograft tumor model to perform in vivo studies in mice. We will also start with the second aim where we expand the PDX-PCa to required number of mice, and generate castration resistant/abiraterone resistant tumors in PDX-PCa mice.

4. IMPACT

Nothing to Report

5. CHANGES/PROBLEMS:

Nothing to Report

6. PRODUCTS:

Nothing to Report

7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

Name: Pheruza Tarapore

Project Role: PI

Researcher Identifier:

Nearest person month worked: 12

Contribution to project: Worked on designing the cell culture experiments, designed primers and templates for making the aptamer-miRNA constructs, did experiments related to these constructs.

Funding Support: This award and NIH grants

Name: Shuk-mei Ho

Project Role: co-I

Researcher Identifier:

Nearest person month worked: 1

Contribution to project: Helped in overall direction and overview of grant, generating reports, some procedures.

Funding Support: This award and NIH grants

Name: Sarah To

Project Role: Post-doctoral researcher

Researcher Identifier:

Nearest person month worked: 6

Contribution to project: Performed the cell culture based experiments with microRNA mimetics and inhibitors.

Funding Support: National Health and Medical Research Council (GNT1070112 (ST)), Australia.

Name: Dan Song

Project Role: Technician

Researcher Identifier:

Nearest person month worked: 3

Contribution to project: Assisted in writing the animal protocol for DoD application. Has been getting materials ready for doing the PDX model

Funding Support: This award and NIH grants

8. SPECIAL REPORTING REQUIREMENTS

Nothing to Report

9. APPENDICES:

Bibliography and References Cited

1. Cooperberg MR, Lubeck DP, Meng MV, Mehta SS, Carroll PR. The changing face of low-risk prostate cancer: trends in clinical presentation and primary management. *J Clin Oncol.* 2004; 22(11): 2141-9. PMC2997214.
<http://www.ncbi.nlm.nih.gov/pubmed/15169800>.
2. Etzioni R, Berry KM, Legler JM, Shaw P. Prostate-specific antigen testing in black and white men: an analysis of medicare claims from 1991-1998. *Urology.* 2002; 59(2): 251-5.
<http://www.ncbi.nlm.nih.gov/pubmed/11834397>.
3. American Cancer Society. *Cancer Facts & Figures.* Atlanta: American Cancer Society 2013; <http://www.cancer.org/research/cancerfactsstatistics/allcancerfactsfigures/index>
4. Pendleton J, Pisters LL, Nakamura K, Anai S, Rosser CJ. Neoadjuvant therapy before radical prostatectomy: where have we been? Where are we going? *Urol.Oncol.* 2007; 25(1): 11-8. <http://www.ncbi.nlm.nih.gov/pubmed/17208133>.
5. Hadaschik BA, Sowery RD, Gleave ME. Novel targets and approaches in advanced prostate cancer. *Curr.Opin.Urol.* 2007; 17(3): 182-7.
<http://www.ncbi.nlm.nih.gov/pubmed/17414516>.
6. Hadaschik BA, Gleave ME. Therapeutic options for hormone-refractory prostate cancer in 2007. *Urol.Oncol.* 2007; 25(5): 413-9.
<http://www.ncbi.nlm.nih.gov/pubmed/17826663>.
7. Kosaka T, Miyajima A, Yasumizu Y, Miyazaki Y, Kikuchi E, Oya M. Limited in vitro efficacy of CYP17A1 inhibition on human castration resistant prostate cancer. *Steroids* 2014; <http://www.ncbi.nlm.nih.gov/pubmed/25150014>. S0039-128X(14)00208-6;10.1016/j.steroids.2014.07.017
8. Bruno RD, Vasaitis TS, Gediya LK, Purushottamachar P, Godbole AM, Ates-Alagoz Z, Brodie AM, Njar VC. Synthesis and biological evaluations of putative metabolically stable analogs of VN/124-1 (TOK-001): head to head anti-tumor efficacy evaluation of VN/124-1 (TOK-001) and abiraterone in LAPC-4 human prostate cancer xenograft model. *Steroids* 2011; 76(12): 1268-79. PMC3171567.
<http://www.ncbi.nlm.nih.gov/pubmed/21729712>. S0039-128X(11)00196-6;10.1016/j.steroids.2011.06.002
9. Mostaghel EA, Marck BT, Plymate SR, Vessella RL, Balk S, Matsumoto AM, Nelson PS, Montgomery RB. Resistance to CYP17A1 inhibition with abiraterone in castration-resistant prostate cancer: induction of steroidogenesis and androgen receptor splice variants. *Clin.Cancer Res.* 2011; 17(18): 5913-25. PMC3184252.

<http://www.ncbi.nlm.nih.gov/pubmed/21807635>. 1078-0432.CCR-11-0728;10.1158/1078-0432.CCR-11-0728

10. Mostaghel EA. Abiraterone in the treatment of metastatic castration-resistant prostate cancer. *Cancer Manag.Res.* 2014; 6: 39-51. PMC3912049.
<http://www.ncbi.nlm.nih.gov/pubmed/24501545>. 10.2147/CMAR.S39318;cmar-6-039
11. Zhou Z, Kennell C, Lee JY, Leung YK, Tarapore P. Calcium phosphate-polymer hybrid nanoparticles for enhanced triple negative breast cancer treatment via co-delivery of paclitaxel and miR-221/222 inhibitors. *Nanomedicine.* 2016;
<http://www.ncbi.nlm.nih.gov/pubmed/27520723>. S1549-9634(16)30106-X [pii];10.1016/j.nano.2016.07.016 [doi]