

AWARD NUMBER: W81XWH-17-1-0201

TITLE: RAGE/Diaph1, Diabetes, and Kidney Disease: Mechanisms and Novel
Therapeutic Strategies

PRINCIPAL INVESTIGATORS: Ann Marie Schmidt, MD

CONTRACTING ORGANIZATION: New York University
New York, NY 10016

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14. ABSTRACT In Year One of the funded grant, we have substantial progress in three critical areas: First, we have established the breeding colonies for all four of the key mouse lines to test the roles of RAGE and DIAPH1 in podocytes and monocytes/macrophages in the pathogenesis of diabetes associated nephropathic changes in the kidney. As detailed in the full progress report, many of these animals (males and females) have already been rendered diabetic and are on time course. Second, we have optimized podocyte isolation procedures as indicated in the grant application. This is a key step which will enable us to probe mechanisms of RAGE and DIAPH1 biology in these cells. Third, Dr. Ramasamy identifies substantial progress in the development and validation of metabolomics and lipidomics assays here at NYU in order to understand detailed mechanisms of the role of these molecules in the diabetic kidney. Taken together, our work in Year 1 ensures that the feasibility of our proposed studies is assured and that through the coming year, our research team will begin to obtain important data to address the questions proposed in our Specific Aims.									
15. SUBJECT TERMS Diabetes; DIAPH1; Floxed Mice; Glomerulosclerosis; Glomerular basement membrane; Inflammation; Macrophage; Nephropathy; Podocyte; RAGE; Small Molecule Antagonist									
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Ann Marie Schmidt, MD – Initiating PI

1. INTRODUCTION:

Our laboratory discovered the receptor for advanced glycation endproducts (RAGE) and we identified that the cytoplasmic domain (tail) of RAGE binds to the formin, DIAPH1, and that this interaction is critical for RAGE ligand-mediated signal transduction and modulation of gene expression linked to cellular perturbation. DIAPH1 mediates actin cytoskeleton functions, cellular migration and activation of the Rho GTPases. DIAPH1 is expressed by immune and vascular cells; we reported that deletion of *Diaph1* in murine macrophages protects against hypoxia-mediated upregulation of proinflammatory (*Egr1* and *Ccl2*) and prothrombotic (*Tf*) and that this protection is analogous to that observed in macrophages devoid of *Ager*. Furthermore, with Dr. Alexander Shekhtman, we have identified the precise mechanism by which the cytoplasmic domain of RAGE binds DIAPH1. Critically, Dr. D'Agati's preliminary data (now published; see below) reveal that DIAPH1, like RAGE, is highly expressed in human diabetic podocytes. **The goal of this grant is to determine the specific mechanisms by which RAGE/DIAPH1 contribute to the pathogenesis of diabetes associated nephropathy and to explore novel RAGE/DIAPH1-directed therapeutic opportunities.**

2. KEYWORDS:

Diabetes
DIAPH1
Floxed Mice
Glomerulosclerosis
Glomerular basement membrane
Inflammation
Macrophage
Nephropathy
Podocyte
RAGE
Small Molecule Antagonist

3. ACCOMPLISHMENTS:

- **What were the major goals of the project?**

There are three specific aims of the funded grant:

AIM 1 will test the hypothesis that RAGE and DIAPH1 mediate diabetic nephropathy (DN) through disengagement of homeostatic actin cytoskeleton dynamics and

upregulation of pro-inflammatory and pro-fibrotic molecules. We will generate mice in which podocyte-specific deletion of *Ager* or *Diaph1* is accomplished via breeding *Ager* or *Diaph1* floxed mice with podocin (*Nphs2*) cre recombinase mice.

AIM 2 will test the hypothesis that RAGE and DIAPH1-expressing macrophages contribute to structural and functional derangements in DN through upregulation of tissue-destructive and profibrotic mediators. We will generate mice in which myeloid cell deletion of *Ager* or *Diaph1* is accomplished by breeding *Ager* or *Diaph1* floxed mice with *Lysm* cre recombinase mice.

AIM 3 will determine if administration of novel small molecule antagonists of RAGE-DIAPH1 interaction in diabetic mice protects against DN.

○ **What was accomplished under these goals?**

1) *Major Activities*

In the first year of the funded grant, we have focused on the following major activities:

- 1A). As noted in the project narrative, we will generate four different lines of mice to directly test the hypothesis that RAGE and DIAPH1 contribute to the pathogenesis of diabetes-associated nephropathy in the podocytes and/or in myeloid cells/macrophages. To do this, we have bred to experimental stage three of the mouse lines and the fourth is in progress.
- 1B). We are characterizing each of the four mouse lines in order to be certain that the gene of interest is deleted under the conditions to be expected with the cre recombinase driver. In our case, this means that we are testing deletion of *Ager* or *Diaph1* in podocytes – antibodies are optimized and test experiments underway.
- 1C). We are isolating podocytes from the mouse models using described techniques. At this stage, we are now culturing the podocytes after the isolation and have obtained all of the needed reagents in order to secure the characterization of these cells so that in vitro experimentation might be performed.
- 1D). We have determined that the small molecule RAGE antagonist is best administered orally and we are in the process of completing testing of three different doses in order to determine the optimal dose for the addition of the quinapril as described in the narrative. This work is underway.
- 1E). For metabolomics and lipidomics assays, Dr. Ramasamy has set up the validation for all of the measurements to be performed. His laboratory will be testing the tissues/cells from the mice through the time course and he has verified all of his experimental systems for the performance of the outlined studies.
- 1F). All of the colleagues and collaborators are in place, with roles and timing defined, in order to execute the outlined studies as expertly and efficiently as possible.

2) *Specific objectives*

Our objectives in year one were to execute the above six activities in order to be certain that the aims of the study would be completed according to the three Specific Aims outlined in Item #1 above.

3) *Significant results or key outcomes, including major findings, developments, or conclusions (both positive and negative):*

3A). Generation of Mouse Models for Study

At this time, we have made extensive progress in three of the four mouse lines for study; the time course for study once diabetes (or control state) is induced is 6 months – at this time, the first cohort of mice (7 animals) have already completed the time course and tissues are being processed as outlined in the Narrative of the funded grant.

Per line, we have generated the following mice and these mice are on time course (Power calculations as described in the Narrative indicated 12 final mice/condition were needed for statistical significance).

Note that the investigators handling/working with the mice are naïve to the genotype; although they know the diabetes/non diabetes status and the sex of the mice, they are not aware of the genotype until after data are entered post sacrifice.

Aim 1:

Ager flox flox *Npfs2* (+/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes:	Male (N=10)
	Female (N=9)
Non-diabetes:	Male (N=9)
	Female (N=9)

Ager flox flox *Npfs2* (wt/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes:	Male (N=10)
	Female (N=10)
Non-diabetes:	Male (N=10)
	Female (N=9)

Diaph1 flox flox *Npfs2* (+/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes:	Male (N=11)
	Female (N=9)
Non-diabetes:	Male (N=7)
	Female (N=7)

Diaph1 flox flox *Npfs2* (wt/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes: Male (N=11)
 Female (N=10)
 Non-diabetes: Male (N=7)
 Female (N=6)

Note that at this time we are filling in the groups – breeding is well underway for all of the above lines in order to achieve final N=12 / group.

Plan is to sacrifice the mice @ 6 months diabetes or control and perform the studies indicated in the Narrative of the funded grant.

Aim 2:

Ager flox flox *lysm* (+/+) cre recombinase mice

Mice are being actively bred at this time. As born, they are being genotyped and will be placed on time course (6 months) as follows:

Diabetes: Male (N=0)
 Female (N=0)
 Non-diabetes: Male (N=0)
 Female (N=0)

Ager flox flox *lysm* (wt/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes: Male (N=0)
 Female (N=0)
 Non-diabetes: Male (N=0)
 Female (N=0)

Diaph1 flox flox *lysm* (+/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes: Male (N=3)
 Female (N=1)
 Non-diabetes: Male (N=1)
 Female (N=1)

Diaph1 flox flox *lysm* (wt/wt) cre recombinase mice

Mice have been bred, genotyped and placed on time course (6 months) as follows:

Diabetes: Male (N=5)
 Female (N=2)
 Non-diabetes: Male (N=3)
 Female (N=2)

Note that at this time we are filling in the groups – breeding is well underway for all of the above lines in order to achieve final N=12 / group.

Plan is to sacrifice the mice @ 6 months diabetes or control and perform the studies indicated in the Narrative of the funded grant.

3B). Characterization of mice

3Bi). We first validated antibodies for RAGE and DIAPH1 in mouse kidney and the co-localization antibodies (Synaptopodin for podocyte deletion and CD68 for macrophage deletion) as follows:

Podocyte deletion: We have sacrificed *Ager* flox flox *Nphs2* (+/wt) and (wt/wt) and our findings reveal that based on qualitative assessment of the images that the expression of podocyte RAGE is significantly lower in the CRE+ vs. CRE- mice. Experiments are underway to quantify these findings at this time. In the case of DIAPH1, we have prepared the animals/tissues for the validation and these experiments are pending at this time.

Macrophage deletion: We have sacrificed *Ager* flox flox *lysm* (++) and (wt/wt) mice and discovered that we need to use homozygote *lysm* cre for significant/sufficient deletion of *Ager*. We discovered this by assessment of the bone marrow derived macrophage expression of *Ager*. In the case of DIAPH1 we have sacrificed *Diaph1* flox flox (+/wt) and (wt/wt) mice and our results reveal that hemizyosity is sufficient. That is, we can use *lysm* (+/wt) in order to test the deletion of *Diaph1* in myeloid cells for these studies.

3Bii). We have isolated podocytes from the above mice and just recently successfully cultured them on plastic dishes. At this time, we are performing the fluorescence microscopy and real time PCR that is needed to both show that they are podocytes and also to document their purity. This work is actively underway at this time.

4) Other achievements.

There are no other achievements to report at this time. There are no goals unmet in this period. We have generated all of the needed mice successfully; we have characterized them; and we have assembled all of the needed collaborators to successfully execute the studies in the outlined Narrative of the funded grant.

o **What opportunities for training and professional development has the project provided?**

Dr. NARA SZOSTACZUK under Dr. Schmidt's mentorship has learned a great deal since her arrival to this project. She has learned the complexities of working with floxed/cre mice and how to understand their characterization. She has learned how to induce and monitor diabetes in the animals and she has learned how to keep careful monitoring records of the mice per the protocol and to prepare for their sacrifice and post-mortem studies. Dr. Szostaczuk has learned how to isolate podocytes and bone marrow derived macrophages from mice and how to properly test antibodies for specificity and how to perform real time quantitative PCR with these tissues. Collectively, she has begun to plan the studies and needed materials for the next phase of her work in the funded Narrative of the grant.

o **How were the results disseminated to communities of interest?**

NOTHING TO REPORT

○ **What do you plan to do during the next reporting period to accomplish the goals?**

As we have indicated throughout the narrative above, we plan to:

- 1). Add the mice to completion in the four groups of animals; sacrifice them at 6 months and continue the tissue/urine analysis.
- 2). We plan to isolate podocytes and macrophages and perform the indicated analyses as outlined in the Narrative.
- 3). With Dr. Ramasamy, we plan to perform metabolomics/lipidomics on the indicated cells in order to discern mechanisms of action in these models.
- 4). We plan to administer the small molecule with quinapril (or without) in order to determine if there is synergy.

4. **IMPACT:**

○ **What was the impact on the development of the principal discipline(s) of the project?**

Despite years of research on diabetic kidney disease, the precise cells that mediate the damage in diabetes are not fully clarified vis-à-vis RAGE and DIAPH1. We know that mice globally devoid of *Ager* or *Diaph1* (new publication 2018) are protected from diabetes associated nephropathy but we do not know the cell specific mechanism. This work as outlined in this funded grant holds great promise to uncover new insights into the mechanisms by which diabetes causes nephropathic changes in the kidney.

○ **What was the impact on other disciplines?**

NOTHING TO REPORT AT THIS TIME

○ **What was the impact on technology transfer?**

NOTHING TO REPORT AT THIS TIME

○ **What was the impact on society beyond science and technology?**

NOTHING TO REPORT AT THIS TIME

5. **CHANGES/PROBLEMS:**

○ **Changes in approach and reasons for change**

There are no changes in approach.

○ **Actual or anticipated problems or delays and actions or plans to resolve them**

There are no actual or anticipated problems to date that delay in any way our planned work.

○ **Changes that had a significant impact on expenditures**

Staff is hired at this time (once the grant was funded, we began the hire process) and mice are being generated and studied as outlined. Hence although there was a delay, it is not appropriate to hire staff and start mouse studies until notice of funding is achieved. We have thus adhered to this principle and all of the staffing/work is actively underway at this time.

- **Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents**

No human subjects

No select agents

No significant changes in the use of vertebrate animals – any amendments were first submitted at NYU and then submitted to DOD ACURO. These amendments had NO impact on the plans of the funded work but involved staffing and amendments to ensure that the aims were carried out as written in the funded grant.

Approval Date of the IACUC:

Approval Date: 8/17/2018

Effective Date: 8/17/2018

Annual Expiration Date: 6/13/2019

Final Expiration Date: 6/13/2020

6. PRODUCTS:

NOTHING TO REPORT

7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

- **What individuals have worked on the project?**

Name:	Ann Marie Schmidt
Project Role:	PI (Initiating)
Researcher Identifier (e.g. ORCID ID):	SCHMIDTAM (eRA Commons ID)
Nearest person month worked:	1
Contribution to Project:	PI, oversight of project and all administrative work with respect to use of animals and the budgetary requirements
Funding Support:	DOD

Name:	Ravichandran Ramasamy
Project Role:	PI (Partnering)
Researcher Identifier (e.g. ORCID ID):	RAVIRAMASAMY (eRA Commons ID)
Nearest person month worked:	1
Contribution to Project:	Dr. Ramasamy developed the methods for the metabolomics and lipidomics analyses of the mice under study.
Funding Support:	DOD

Name:	Nara Jimena Szostaczuk
Project Role:	Postdoctoral Fellow
Researcher Identifier (e.g. ORCID ID):	szostn01 (eRA commons ID)

Nearest person month worked: 2
 Contribution to Project: Leads studies on work in generating mice for diabetes, time course and molecular analyses
 Funding Support: DOD

Name: Raquel Lopez-Diez
 Project Role: Postdoctoral Fellow
 Researcher Identifier (e.g. ORCID ID): DIEZR01 (eRA Commons ID)
 Nearest person month worked: 8
 Contribution to Project: Contributed to podocyte isolation processes and characterization with immunostaining strategies
 Funding Support: DOD

Name: Karan Singh
 Project Role: Postdoctoral Fellow
 Researcher Identifier (e.g. ORCID ID): SINGHKARAN (eRA Commons ID)
 Nearest person month worked: 3
 Contribution to Project: Contributed to podocyte isolation processes and characterization with real time quantitative PCR
 Funding Support: DOD

Name: Vivette D'Agati
 Project Role: Co-Investigator
 Researcher Identifier (e.g. ORCID ID): VDA1234 (eRA Commons ID)
 Nearest person month worked: 1
 Contribution to Project: Pathological analysis of kidney tissues
 Funding Support: DOD

Name: Richard A. Friedman
 Project Role: Co-Investigator
 Researcher Identifier (e.g. ORCID ID): FRIEDMANR (eRA Commons ID)
 Nearest person month worked: 1
 Contribution to Project: Statistical and bioinformatics analyses
 Funding Support: DOD

- **Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?**

Schmidt, Ann Marie

ACTIVE

P01HL60901	07/15/11-11/30/18	0.12 calendar
NIH	\$897,537	No Cost Extension
RAGE and Mechanisms of Vascular Dysfunction		

(THIS AWARD)

USAMRAA Dept. of the Army 07/01/17-06/30/19 0.96 calendar
 Receptor for AGE (RAGE) Signal Transduction in Amyotrophic Lateral Sclerosis: In Vivo
 Imaging and Novel Therapeutic Approaches
 \$251,092

Major goals of this grant includes testing the hypothesis that microglia RAGE, through ligand-driven upregulation of inflammatory and pro-oxidative stress and suppression of reparative processes in the ALS spinal cord, mediates neuronal death and loss of motor function and probing the hypothesis that PBMM-specific deletion of Ager attenuates neuronal stress, accumulation of A β and amyloid plaques, synaptic dysfunction and cognitive impairment in APP^{swe}/PS1 mice.

Role: PI

(NEW)

USAMRAA Dept. of the Army 09/30/17-08/31/20 0.73 calendar
 \$489,612

RAGE/Diaph1, Diabetes, and Kidney Disease: Mechanisms and Novel Therapeutic Strategies
 Major goals for this grant involves (a) testing the hypothesis that RAGE and DIAPH1 mediate podocyte dysfunction in DN through disengagement of homeostatic actin cytoskeleton dynamics and upregulation of pro-inflammatory and pro-fibrotic molecules (b) testing the hypothesis that RAGE and DIAPH1-expressing macrophages contribute to structural and functional derangements in DN through upregulation of tissue-destructive and profibrotic mediators and (c) determining if administration of novel small molecule antagonists of RAGE-DIAPH1 interaction in diabetic mice protects against DN.

Role: PI (Ramasamy-Partnering PI)

(NEW)

American Heart Association 04/01/17-03/31/21 3.6 calendar
 \$900,663

Braking Inflammation in Obesity & Metabolic Dysfunction: Translational and Therapeutic Opportunities

The major goal of this grant is to investigate the novel hypothesis that impaired adipocyte, macrophage and other inflammatory cell signal transduction thwarts weight loss and its anti-inflammatory and metabolic benefits, at least in part through the activation of the receptor for advanced glycation endproducts, or RAGE pathway, which has been shown to regulate a unique repertoire of inflammatory and metabolic processes.

Role: Center Director, Project 1 Leader

INACTIVE(ENDED)

1R01 HL118565 06/01/13-04/30/18 0.9 calendar
 NIH \$248,449
 RAGE, Macrophages & HDL Biology

This grant examines the molecular mechanisms by which RAGE regulates cholesterol transport.

Role: PI

(ENDED)

Harrington Discovery Institute 01/01/16-12/31/17 0.12 calendar
\$50,000

Targeting RAGE/mDial for the Prevention and Treatment of Diabetic Complications
The goal of the Harrington Discovery Institute project is to develop LOCAL intraocular and transdermal treatments for diabetic retinopathy and diabetic wound healing, respectively.

Role: PI

OVERLAP

None

D'Agati, Vivette

ACTIVE

(NEW)

2 PO1 DK 56492 (Klotman) renewal 05/15/17-04/30/22 1.80 calendar
NIH NIDDK \$91,325 (Core B)

Long-term Consequences of HIV Infection of the Kidney

The aim of this grant is to explore the long-term consequences of HIV infection of the kidney beyond HIVAN.

The role of Core B will be to provide technical and interpretative support in light microscopy, immunohistochemistry, immunofluorescence and electron microscopy in mouse and human kidney tissues for all projects, including study of the mechanisms of ApoL1 induced kidney injury, the kidney as long-term reservoir for HIV, the interactions between gut microbiome and HIVAN in the Tg26 model, and the effects of chronic HIV infection on progression of kidney disease through enhanced pro-inflammatory responses.

Role: Core B Leader

1R01MD009223-01 (Multi-PI: Gharavi & Bomback) 07/01/14-06/30/19 0.60 calendar
NIH/NIMHD \$250,000

Ancestry, Genetic Risk and Health Disparities in Immune-Mediated Nephritis

The major goal of this project is to investigate the role of shared and distinct genetic factors among Europeans, Asians, Hispanics, and African-Americans in the onset, course, and ultimate outcomes of IgA nephropathy, membranous nephropathy and lupus nephritis.

Role: Co-I

1R24DK103032-01 (Schmidt) 08/01/14/-07/31/19 0.70 calendar

NIH \$541,112

Targeting RAGE-mDia1 in Diabetic Complications: Mechanisms & Therapeutics

This application focuses on the role of the receptor for advanced glycation endproducts (RAGE) and its cytoplasmic domain binding partner, mammalian form of diaphanous1, mDia1, which is essential for RAGE signaling as a fundamental therapeutic target for diabetic complications.

Role: Co-I

(NEW)

Dept. of the Army – USAMRAA (Gharavi) 08/15/16 – 08/14/19 0.84 calendar
Grant number: PR151419 \$250,000

Multispecies, integrative GWAS for focal segmental glomerulosclerosis

Goals: The major goal of this project is to devise new targeted therapies that will impact and benefit treatment for chronic kidney disease at large in the general population as well as in the active duty personnel, veterans and their families. Aims: - Specific aim 1: A Genome-wide association study for common single nucleotide polymorphisms and rare copy number variations in 7,559 FSGS and over 50,000 controls - Specific aim 2: A GWAS for FSGS in a mouse leveraging the power of the newly developed DO strains - Specific aim 3. Cross annotation between human and mouse GWAS and identification of downstream dysregulated pathways and networks.

Role: Co-I

1R01DK106436-01A1 (Winchester) 01/25/16-12/31/19 0.6 calendar
NIH/NIDDK \$211,308

Significance of Intrarenal T Cells in SLE Nephritis

The overall goal of this study is to define the role of T cells in the development and progression of chronic lupus nephritis. It addresses the clinical problem of why nearly half of lupus nephritis cases do not adequately respond to therapy and much progress to chronic glomerulonephritis. SLE kidneys have a variable but often extensive infiltrate of predominantly clonally expanded CD4 and/or CD8 T cells with features that suggest they could drive the inflammatory process. We hypothesize that while acute glomerulitis is driven by immune complexes, chronic SLE nephritis is driven by the development of CD4 and especially CD8 T cell clonal recognition of self-peptides, resulting in glomerular and tubular cell injury. In the first aim we will delineate the extent and character of the renal CD4 or CD8 T cell infiltration in new onset nephritis and correlate these findings with short- and long-term pathological and clinical outcomes that predict poor response to therapy and progressive renal disease. In the second part of this aim we will also determine the T cell characteristics of cases with worsening renal involvement requiring repeat biopsy, comparing current and prior biopsies for T cell features that predict progression. In the second aim we will define the properties of the infiltrating T cells and their potential mechanisms to mediate renal injury, discriminating between the phenotype of clonally expanded CD4 or CD8 T cells that potentially drive renal injury and that of the polyclonal T cells secondarily recruited by inflammation.

Role: Co-I

(NEW)

R01 DK109544-01A1 (Lee) 04/01/17-03/31/22 0.60 calendar
 NIH \$225,000

Paneth cells and acute kidney injury

This proposal seeks to elucidate the mechanisms as well as therapies for remote organ dysfunction after renal ischemia and reperfusion injury.

Role: Co-I

(NEW)

1R01 DK115694-01 (Lee) 09/13/17-07/31/22 0.60 calendar
 NIH \$250,000

Peptidylarginine deiminase-4 and acute kidney injury

This project seeks to elucidate the mechanisms as well as therapies for inflammation and injury after renal ischemia and reperfusion.

Role: Co-I

(NEW)

1UG3DK114926-01 (Kiryluk, Barasch, Bomback) 07/01/17 - 06/30/22 0.90 calendar
 NIH/NIDDK \$300,000

Kidney Precision Medicine Program (KPMP): Columbia AKI Recruitment Site

The national Kidney Precision Medicine Project (KPMP) aims to reduce the significant global health burden of acute kidney injury (AKI) by elucidating mechanisms and effective therapies through precision medicine technologies.

Role: Co-I

(THIS AWARD)

Dept. of the Army – USAMRAA (Schmidt) 07/01/17 - 06/30/20 0.60 calendar
 \$489,612

RAGE/Diaph1, Diabetes, and Kidney Disease: Mechanisms and Novel Therapeutic Strategies

This project will test the hypothesis that RAGE and DIAPH1 mediate podocyte dysfunction in DN through disengagement of homeostatic actin cytoskeleton dynamics and upregulation of pro-inflammatory and pro-fibrotic molecules and will determine if administration of novel small molecule antagonists of RAGE-DIAPH1 interaction in diabetic mice protects against DN.

Role: Co-I

INACTIVE

1 UM1 DK100876-01 (Gharavi) 09/16/13-05/31/18 1.2 calendar
 NIH/NIDDK \$353,143

Advancing Clinical Research in Primary Glomerular Diseases

The major goals of this project are to develop a longitudinal observational cohort of patients with biopsy-documented forms of major glomerular diseases, including minimal change disease, focal segmental glomerulosclerosis, membranous glomerulopathy and IgA nephropathy.

Role: Co-I

OVERLAP

None

Friedman, Richard A.

ACTIVE

5 P30 CA13696-44 (Emerson)	07/01/14 - 06/30/19	1.2 calendar
NCI	\$2,248,065	
Cancer Center Support Grant		

This grant supports the leadership of Columbia University's lab, clinical & population-based cancer research programs & the shared resources serving the University's Cancer Center members.

Dr. Friedman's role as a member of the Biomedical Informatics Shared Resource is to provide the bioinformatics and statistical component of cancer research projects.

Role: Staff Member of the Biomedical Informatics Shared Resource.

1R35CA210088-01 (Wang-PI)	12/09/16-11/30/23	1.2 calendar
NIH/NCI	\$540,000	

The Role of Stem Cells and the Microenvironment in Gastrointestinal Cancer

This project seeks to investigate the role of nerves and other stromal cells in the development of digestive cancers, including stomach, esophageal, colon and pancreas. The project builds on previous work that suggests that these elements can regulate stem cells and that inhibiting stromal cells in the microenvironment, it may be possible to inhibit the development of tumors. Dr. Friedman's role is to design and analyze RNASeq and other genomic experiments and perform other statistical analyses.

Role: Co-I

(NEW)

1U54CA163004-06 (Wang, PI)	05/12/17-4/30/22	0.60 calendar
NIH/NCI	\$978,861	

The Role of the Microenvironment in Barrett's Esophagus

To investigate how the tumor microenvironment leads to adenocarcinoma. Dr. Friedman's role is to design and analyze RNASeq experiments.

Role: Co-I

1R01CA208711-01 Sepulveda (PI) 09/01/2016-08/31/21 1.0 calendar
NIH/NCI \$228,750

Genomics and Mechanisms of Esophageal Carcinogenesis

The goal of this project is to investigate the role CDKN1A/P16 mutations the genesis of esophageal cancer; to use expression, polymorphism, and methylation data to predict progression to cancer, and to test various drugs for their ability to prevent this progression from occurring. Dr. Friedman's role is to perform statistical analyses.

Role: Co-I

1R01CA178445-04 (Su, PI) 07/01/15-06/30/20 0.6 calendar
NIH/NCI \$294,426

The Role of wild-type Kras in the context of tumor progression and metastasis

To elucidate the mechanism of the role of Kras in human pancreatic ductal adenocarcinoma by means of a mouse model. Dr. Friedman's role is to design and analyze RNASeq and PCR experiments to measure gene expression.

Role: Co-I

1R01DK109675-01 (Schmidt) 04/01/16-03/31/21 0.9 calendar
NIH/NIDDK \$313,163

RAGE/MDIA1, Macrophage Trafficking and Inflammation in High Fat Feeding

To characterize the mechanism by which macrophage Receptor for Advanced Glycation Endproducts regulates obesity, adiposity and metabolic dysfunction in high fat feeding, both inherently and via cross-talk with the adipocyte.

Role: Co-I

1R01HL132516-01-A1 (Ramasamy, Schmidt-MPI) 12/09/16-11/30/20 1.13 calendar
NIH/NHLBI \$375,975

RAGE/mDia1, Macrophage Trafficking and Inflammation in Regression of Diabetic Atherosclerosis

The goal of this project is to characterize how RAGE and mDia1 signaling macrophages affect atherosclerotic regression. Dr. Friedman's role is to perform genomic and statistical analyses.

Role: Co-investigator

(NEW)

1P01HL131481-01A1 (E. Fisher PI) 05/01/17-04/30/19 1.8 calendar
NIH/NHLBI \$1,434,208

Macrophage Dysfunction in Obesity Diabetes and Atherosclerosis

The goal of this project is to characterize how macrophages affect atherosclerosis in patients who are diabetic, and/or obese. Dr. Friedman's role is to perform genomic and statistical analyses.

Role: Co-I

(THIS AWARD)

17-A0-00-007334-01 (Schmidt, Ramasamy, MPI) 09/30/17-09/29/20 0.6 calendar
DOD/CDRMP \$489,612

RAGE/Diaph1, Diabetes, and Kidney Disease: Mechanisms and Novel Therapeutic Strategies
To characterize the mechanism by which macrophage Receptor for Advanced Glycation Endproducts helps cause diabetic kidney disease and develop therapies for the treatment of this disease. Dr. Friedman's role is to analyze RNASeq and other experiments.

Role: Co-I

INACTIVE

W81XWH-15-1-0296 Broustas (PI) 08/31/2015-08/30/2018 0.24 calendar
DOD \$108,000

Targeting MEK5 Enhances Radiosensitivity in Human Prostate Cancer

The goal of this project is to test whether inhibition of MEK5 signaling enhances the response of human prostate cancer cell lines to radiation therapy. Dr. Friedman's role is to perform statistical analyses

Role: Co-investigator

1R01HL118565-04 (Schmidt/Friedman, PI) 06/01/13-4/30/18 1.2 calendar
NIH/NHLBI \$262,295

RAGE, Macrophages, and HDL Biology

To link RAGE (Receptor for Advanced Glycation Endproducts) function to that of High Density Lipoproteins. Dr. Friedman's role is to design and analyze RNASeq experiments to discover molecular mechanisms underlying obesity.

Role: Co-I

5R03CA186218-02 (Abrams, PI) 07/01/15-06/30/17 0.3 calendar
NIH/NIDCR

Randomized placebo-controlled trial of a gastrin receptor (Abrams: PI)

Time Commitment: 0.3 calendar months

The goal of this project is to test the effect of a netazepide (YF476), a gastrin receptor antagonist, on biomarkers associated with progression to esophageal adenocarcinoma. Dr. Friedman's role is to design and analyze RNASeq and PCR experiments to measure gene expression.

Specific Aims:

Role: Co-I

OVERLAP

None

- **What other organizations were involved as partners?**

Organization Name: Columbia University

Location of Organization: New York, NY

Collaborating Investigators: Drs. Vivette D'Agati and Richard A. Friedman

Collaboration: Dr. D'Agati performed the pathological analysis of kidney tissues. Dr. Friedman performed statistical and bioinformatics analyses.

8. SPECIAL REPORTING REQUIREMENTS

- **COLLABORATIVE AWARDS:**

Dr. Ravichandran Ramasamy, PhD (Partnering PI) progress report follows Dr. Schmidt's section.

9. APPENDICES:

Nothing to report