

REPORT DOCUMENTATION PAGE			Form Approved OMB NO. 0704-0188		
<p>The public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington VA, 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.</p>					
1. REPORT DATE (DD-MM-YYYY) 16-01-2019		2. REPORT TYPE Final Report		3. DATES COVERED (From - To) 1-Sep-2014 - 31-May-2015	
4. TITLE AND SUBTITLE Final Report: Improving Visual Perception by Brain Centrifugal Modulation			5a. CONTRACT NUMBER W911NF-14-1-0555		
			5b. GRANT NUMBER		
			5c. PROGRAM ELEMENT NUMBER 611102		
6. AUTHORS			5d. PROJECT NUMBER		
			5e. TASK NUMBER		
			5f. WORK UNIT NUMBER		
7. PERFORMING ORGANIZATION NAMES AND ADDRESSES University of Notre Dame ND Research 940 Grace Hall Notre Dame, IN 46556 -5708			8. PERFORMING ORGANIZATION REPORT NUMBER		
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS (ES) U.S. Army Research Office P.O. Box 12211 Research Triangle Park, NC 27709-2211			10. SPONSOR/MONITOR'S ACRONYM(S) ARO		
			11. SPONSOR/MONITOR'S REPORT NUMBER(S) 65632-LS-II.2		
12. DISTRIBUTION AVAILABILITY STATEMENT Approved for public release; distribution is unlimited.					
13. SUPPLEMENTARY NOTES The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision, unless so designated by other documentation.					
14. ABSTRACT					
15. SUBJECT TERMS					
16. SECURITY CLASSIFICATION OF:		17. LIMITATION OF ABSTRACT		15. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT UU	b. ABSTRACT UU	c. THIS PAGE UU	UU		Lei Li
				19b. TELEPHONE NUMBER 574-631-7488	

**RPPR Final Report**  
as of 24-Jan-2019

Agency Code:

Proposal Number: 65632LSII

**Agreement Number: W911NF-14-1-0555**

**INVESTIGATOR(S):**

**Name:** Lei Li  
**Email:** Li.78@nd.edu  
**Phone Number:** 5746317488  
**Principal:** Y

Organization: **University of Notre Dame**

Address: ND Research, Notre Dame, IN 465565708

Country: USA

DUNS Number: 824910376

EIN: 350868188

**Report Date:** 30-Jun-2015

Date Received: 16-Jan-2019

**Final Report** for Period Beginning 01-Sep-2014 and Ending 31-May-2015

**Title:** Improving Visual Perception by Brain Centrifugal Modulation

**Begin Performance Period:** 01-Sep-2014

**End Performance Period:** 31-May-2015

**Report Term:** 0-Other

Submitted By: Lei Li

Email: Li.78@nd.edu

Phone: (574) 631-7488

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

**STEM Degrees:** 4

**STEM Participants:** 3

**Major Goals:** The objectives of the proposed research are to study the mechanisms and physiological roles of sensory integration in visual system functions. In all vertebrate species examined thus far, the retinas receive centrifugal input from the brain. In fish, the centrifugal pathway originates from olfactory terminalis neurons (TNs). The TNs project axons to the retina where they synapse with dopaminergic amacrine cells, and occasionally, retinal ganglion cells. In this research, we will study the mechanisms and roles of olfacto-retinal sensory integration in modulation of retinal sensitivity. The research will be conducted using transgenic zebrafish models. In Aim 1, we will test a hypothesis that in the retina the olfactory signals are separately processed by GnRH- and dopamine-mediated signaling pathways. In Aim 2, we will identify key factors that regulate TN axonal projection to the retina. The proposed research is not about fish vision or olfaction; instead it will study the fundamentals related to multi-sensory perception. The goal of this research is to understand the mechanisms of sensory integration in vertebrates including humans.

**Accomplishments:** During the funding period, we examined the development and pathfinding of terminalis nerves (TNs) in zebrafish. The TNs are located in the olfactory bulb, and they project axons to many brain areas including the neural retina. Using genetic and immunocytochemical techniques, we demonstrated that in the retina, the TN axons synapse with dopaminergic interplexiform cells (DA-IPCs). Furthermore, we examine the role of GnRH (released by TN) and dopamine (released by DA-IPC) signaling in TN axon projection to the retina using transgenic zebrafish. The research was conducted using both developing embryos and adult animals. The results from this research were published in *Developmental Neuroscience*. In summary, we accomplished the goals proposed in the research plan.

**Training Opportunities:** During the finding period, we trained three undergraduate students (Delaney Pfister, Dasom Kim, Audrey Drowing) and one graduate student (Jingling Li). All of them were from the Univ of Notre Dame. They have graduated between 2015 and 2017.

**Results Dissemination:** Nothing to Report

**Honors and Awards:** Nothing to Report

**Protocol Activity Status:**

**Technology Transfer:** Nothing to Report

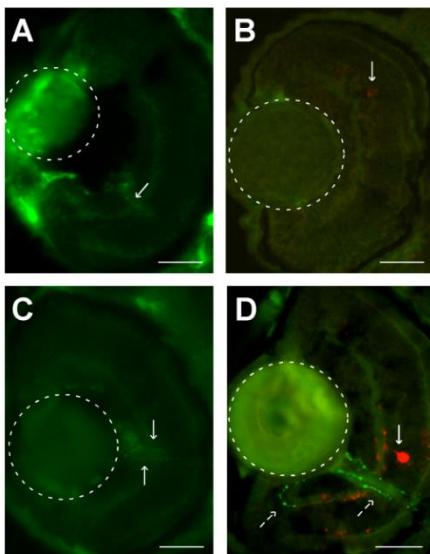


## RESEARCH SUMMARY

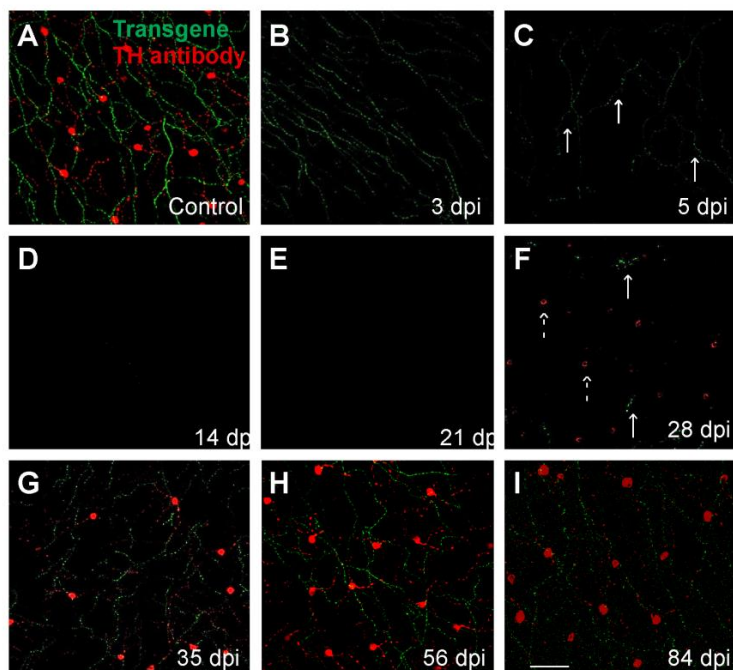
The goal of the proposed research is to investigate the mechanisms and developmental courses of terminalis nerve (TN) input in retinal functions using zebrafish models. In accordance with the research proposal, during the funding period, we performed different sets of experiments. We accomplished the goals proposed in our research plan. Below are summary of the experiments we performed. Some of the research results have been published.

### *Determine the effects of dopaminergic signaling on TN axonal projection*

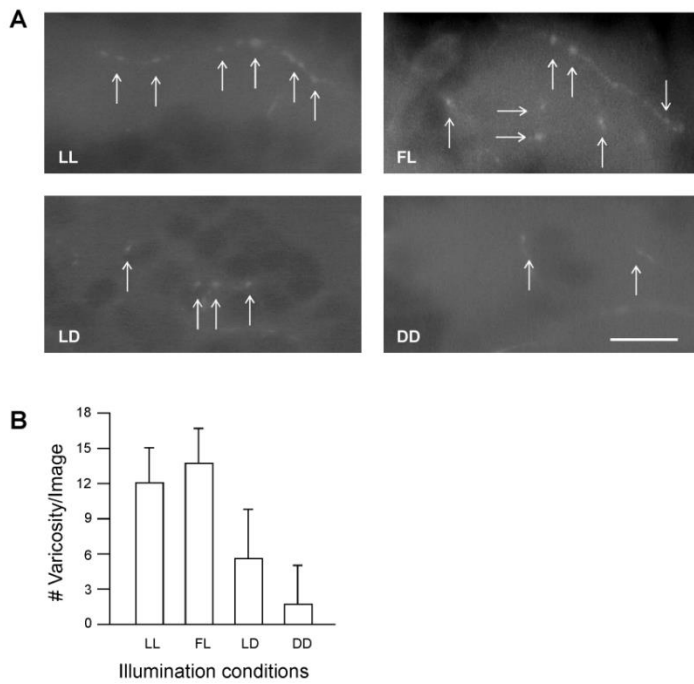
We examined the role of dopamine signaling in axon pathfinding during embryonic development. The research was conducted using transgenic zebrafish (developing embryos and adults), in combination with the use of immunocytochemical methods. We found that dopamine may function as an attractive factor that guides the projection of TN axons to the retina (Fig 1, Fig 2, and Fig. 3).



**Fig 1.** Cryostat sections of transgenic Tg(GnRH3::GFP) fish that show the TN axons (detected by the expression of the GFP conjugated transgene) and dopaminergic cells (labeled by anti-TH antibodies). A, B: Retinal sections from embryos between 48 and 50 hours of age. TN axons were observed in the optic nerve (A, arrow). At the same developmental stages, a dopaminergic cell was seen in the inner nuclear layer (B, red, pointed by the arrow). C: A cross section from a 52-hour-old embryo. Branches of TN axons were seen in the optic nerve (arrows). D: A cross section from a 62-hour-old embryo. Note that the TN axons (green, pointed by arrows with dashed lines) elongate toward to dopaminergic cell (red, pointed by the arrow with solid line). Dashed circles outline the lens. Scale bar: 50  $\mu$ m.



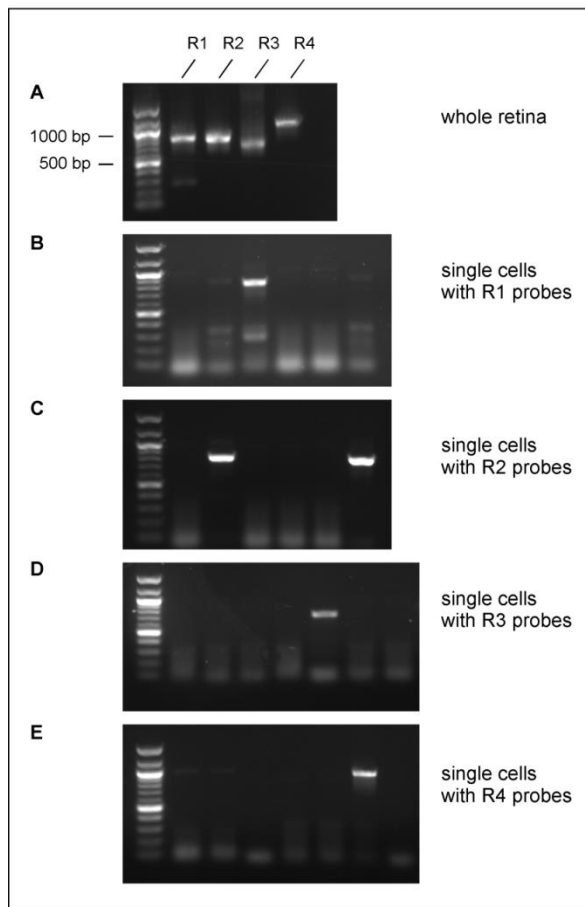
**Fig 2.** Whole-mount retinas of adult Tg(GnRH3::GFP) fish that show the TN axons and dopaminergic cells before (A) and after 6-OHDA treatment (B-I). Images were taken in the nasal-ventral retina, adjacent to the optic nerve. Note the lack of TH immunoreactivity at 3, 5, 14 and 21 dpi (days post injection; panels B, C, D and E). On 28 dpi, regeneration of dopaminergic cells was evident (red, pointed by arrows with dashed lines) and re-distribution of the TN axons (green, pointed by arrows with solid lines) in the retina was detected. At 35, 56 and 84 dpi, abundance in dopaminergic cells and TN axons were found in the retina. Scale bar: 50  $\mu$ m.



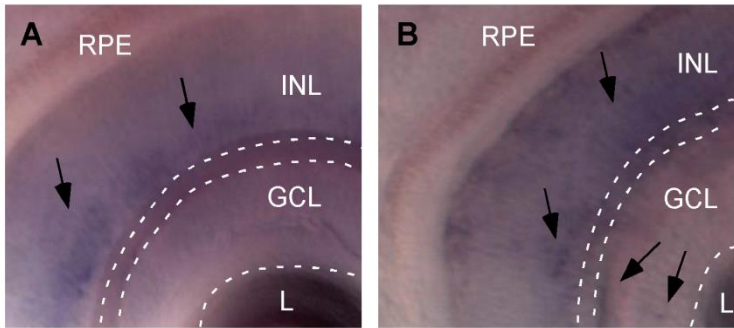
**Fig 3.** Effects of different lighting conditions (which result in different concentrations of dopamine in the retina) on TN axonal projection. A: Fluorescent images of 7-day-old retinas grew up under different lighting conditions. The TN axons were labeled by the GnRH::GFP transgene. Images were taken in the ventral retina adjacent to the optic nerve (captured using a regular camera, not confocal). LL, constant light; FL, randomly expose to 15-min light pulses during the day and night; LD, normal light-dark cycles; DD, constant dark. Scale bar: 100  $\mu$ m. B: Quantitative analysis of TN axonal varicosities in the retina under different lighting conditions. Data represent the means  $\pm$  SE. (n=8 for each condition).

#### Determine the expression of GnRH receptors in zebrafish retinas

We examined the expression of GnRH receptors in zebrafish retinas using whole-retina and single-cell PCR as well as in situ hybridization methods. We found that the GnRH receptors are expressed in the retina, where they mediate the development and function of TN input from the olfactory bulb (Fig 4 and Fig 5).



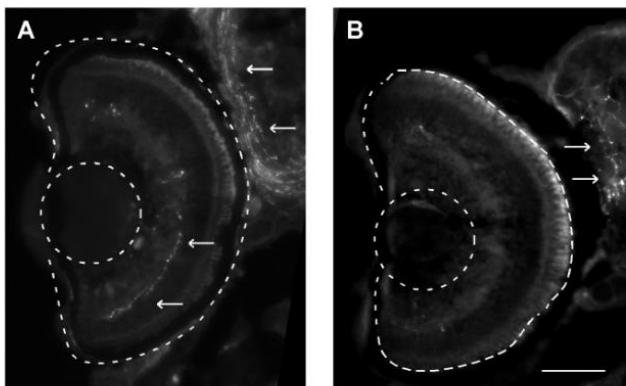
**Fig 4.** RT-PCR of the expression of GnRH receptors in zebrafish retinas. (A) Whole-retina RT-PCR. The PCR products of GnRH receptors (R1-R4) are 831, 807, 676, and 1,004 bp, respectively. We detected the expression of all the 4 types of GnRH receptors from whole-retina preparations. (B-E) Single-cell RT-PCR with R1, R2, R3, and R4 receptor probes, respectively. Cells were isolated in dissociated retinal tissues. Based on their morphologies, they are most likely amacrine cells and ganglion cells. However, we haven't identified the cell types using cell-specific gene markers. Note that individual GnRH receptor types were detected in individual retinal cell types.



**Fig 5.** In situ hybridization of type-1 (A) and type-2 (B) GnRH receptors in zebrafish retinas (from 5 dpf embryos). It showed that type-1 receptors are expressed in the INL, whereas type-2 receptors are expressed in the INL and GCL (arrows). Dashed lines outline the lens and interplexiform layer. Abbreviations: RPE, retinal pigment epithelium; INL: inner nuclear layer; GCL, ganglion cell layer; L, lens.

*Determine the effect of GnRH signaling in TN axonal pathfinding*

We also examined the effects of GnRH signaling transduction on TN projection in developing zebrafish retinas. Because the retinal cells express GnRH receptors, we tested a hypothesis that GnRH, which is released from the TN, is required for TN axonal projection to the retina. The experiments were performed using transgenic zebrafish, and the function of GnRH receptors was blocked by intraocular injection of pharmacological compounds MPDM. We found that in the lack of GnRH signaling, TN axons are no longer projected to the retina (Fig 6).



**Fig 6.** Cryostat sections that show the TN axons in control (A) and MPDM-injected (B) animals. In control animals, TN axons (labeled by the expression of the transgene GnRH3::GFP; pointed by arrows) were seen in the retina and the brain. However, in MPDM-injected fish, no GFP-labeled TN axons were detected in the retina, but TN axons were seen in the brain (arrows). Dashed lines outline the retinas and lenses. Scale bar: 50  $\mu$ m.