

AWARD NUMBER: W81XWH-18-1-0124

TITLE: Proinflammatory Epithelial Cells as a Therapeutic Target in Chronic Pancreatitis

PRINCIPAL INVESTIGATOR: L. Charles Murtaugh, PhD

CONTRACTING ORGANIZATION: University of Utah
Salt Lake City, UT 84112

REPORT DATE: July 2019

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

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REPORT DOCUMENTATION PAGEForm Approved
OMB No. 0704-0188

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1. REPORT DATE July 2019		2. REPORT TYPE Annual		3. DATES COVERED 6/1/2018-5/31/2019	
4. TITLE AND SUBTITLE Proinflammatory Epithelial Cells as a Therapeutic Target in Chronic Pancreatitis				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-18-1-0124	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Lewis Charles Murtaugh, PhD				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
E-Mail: murtaugh@genetics.utah.edu					
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Utah 75 South 2000 East Salt Lake City, UT 84112-8930				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Development Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT The focus of our study is the role of de-differentiated exocrine acinar cells in perpetuating inflammation and tissue injury in chronic pancreatitis. We hypothesize that the pro-differentiation transcription factor Ptf1a promotes recovery from acute pancreatitis, in part through secretion of anti-inflammatory signals such as FGF21, and that downregulation of Ptf1a in chronic pancreatitis leads to sustained inflammation. We have performed experiments to further establish the chronic pancreatitis susceptibility model in Ptf1a-knockout mice, and we have begun rescue experiments in which we try to reverse this phenotype by treatment of Ptf1a-knockout mice with FGF21. Progress has been notably slower than expected, however, owing primarily to turnover of personnel. Nonetheless, we have established the essential conditions required to perform our key experiments, and also generated the required compound mutant mouse strains needed for this work, which we anticipate completing in the near future.					
15. SUBJECT TERMS pancreatitis, pancreas, exocrine, acinar, inflammation, differentiation, Ptf1a, FGF21					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT Unclassified	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON USAMRMC
a. REPORT Unclassified	b. ABSTRACT Unclassified	c. THIS PAGE Unclassified			19b. TELEPHONE NUMBER (include area code)

Standard Form 298 (Rev. 8-98)
Prescribed by ANSI Std. Z39.18

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- 1. INTRODUCTION:** *Narrative that briefly (one paragraph) describes the subject, purpose and scope of the research.*

The subject of this proposal is chronic pancreatitis, an inflammatory condition of the pancreas that is debilitating and untreatable. We hypothesize that persistent inflammation in this disease is driven in part by signals from residual epithelial cells that form via de-differentiation of exocrine acinar cells, the majority cell type in the organ. In mouse models, we propose to determine if the pro-differentiation transcription factor Ptf1a and its downstream target gene FGF21 act to inhibit inflammation by stabilizing the differentiated state.

- 2. KEYWORDS:** *Provide a brief list of keywords (limit to 20 words).*

pancreatitis, pancreas, exocrine, acinar, inflammation, differentiation, Ptf1a, FGF21

- 3. ACCOMPLISHMENTS:** *The PI is reminded that the recipient organization is required to obtain prior written approval from the awarding agency grants official whenever there are significant changes in the project or its direction.*

What were the major goals of the project?

List the major goals of the project as stated in the approved SOW. If the application listed milestones/target dates for important activities or phases of the project, identify these dates and show actual completion dates or the percentage of completion.

Aim 1. Determine if PTF1A expression is sufficient to revert ADM cells to a re-differentiated state and resolve inflammation and tissue injury in a chronic pancreatitis model.

Major Task 1. Establish *Ptf1a*-cKO/GOF model, determine effects of re-expressing PTF1A on chronic pancreatitis-like phenotype

Months 1-14 – 25% complete

Aim 2. Determine if exogenous FGF21 administration is sufficient to revert the pro-inflammatory phenotype of ADM cells, and resolve tissue injury, independent of re-differentiation.

Major Task 2. Determine effects of FGF21-Fc on chronic-pancreatitis-like phenotype of *Ptf1a*-cKO mice

Months 1-18 – 25% complete

Aim 3. Identify markers of ADM cells in mouse pancreas, and characterize their expression in human chronic pancreatitis.

Major Task 3. Pilot studies to optimize FACS isolation of specific cell types

Months 4-8 – 75% complete

Major Task 4. RNA-seq analysis of acinar, duct and ADM-specific gene expression

Months 1-13 – 25% complete

Major Task 5. Analyzing protein marker expression in mouse and human chronic pancreatitis

Months 13-18 – 0% complete

What was accomplished under these goals?

For this reporting period describe: 1) major activities; 2) specific objectives; 3) significant results or key outcomes, including major findings, developments, or conclusions (both positive and negative); and/or 4) other achievements. Include a discussion of stated goals not met. Description shall include pertinent data and graphs in sufficient detail to explain any significant results achieved. A succinct description of the methodology used shall be provided. As the project progresses to completion, the emphasis in reporting in this section should shift from reporting activities to reporting accomplishments.

Major activities

Progress on this project has been slow due to personnel turnover: the individual with the greatest expertise in our experimental system, who generated most of the preliminary data, defended their PhD thesis and left the lab in May, 2018, prior to the start of the project; in addition, my lab technician left shortly thereafter for family reasons. I clearly underestimated the challenge of bringing a new graduate student up to speed on the experimental approaches of this project, and we encountered numerous technical challenges related to the reliability of establishing chronic pancreatitis in *Ptf1a* conditional knockout (cKO) mice. These have been overcome, i.e. our model has been re-established in which *Ptf1a* cKO mice, but not controls, develop unresolved injury and inflammation after acute treatment with the drug caerulein (Fig. 1).

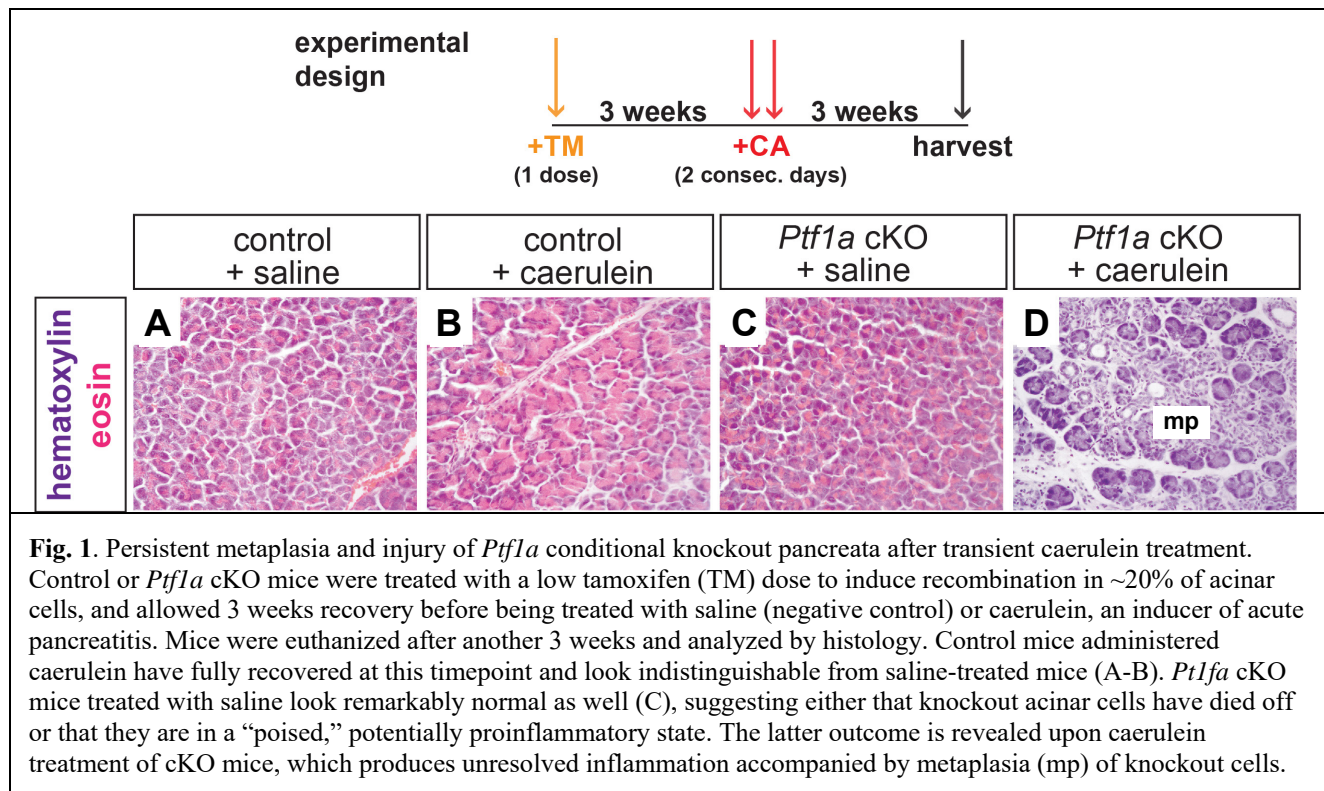


Fig. 1. Persistent metaplasia and injury of *Ptf1a* conditional knockout pancreata after transient caerulein treatment. Control or *Ptf1a* cKO mice were treated with a low tamoxifen (TM) dose to induce recombination in ~20% of acinar cells, and allowed 3 weeks recovery before being treated with saline (negative control) or caerulein, an inducer of acute pancreatitis. Mice were euthanized after another 3 weeks and analyzed by histology. Control mice administered caerulein have fully recovered at this timepoint and look indistinguishable from saline-treated mice (A-B). *Ptf1a* cKO mice treated with saline look remarkably normal as well (C), suggesting either that knockout acinar cells have died off or that they are in a “poised,” potentially proinflammatory state. The latter outcome is revealed upon caerulein treatment of cKO mice, which produces unresolved inflammation accompanied by metaplasia (mp) of knockout cells.

The outcome depicted in Fig. 1D represents the starting point for the interventions described in our Specific Objectives, below. After some challenges in expanding our mouse colony, we have established a robust breeding scheme to generate the mice needed for these objectives, and we have optimized multiple aspects of our tamoxifen and caerulein treatment methodologies to ensure

consistency. We have also confirmed the biological activity of FGF21 in control mice; as described by others, we find that treating control mice with FGF21 protein (provided by Amgen) causes reduced weight gain with age as well as improving glycemic control. Finally, we have established a reasonably effective dissociation procedure, in control mice, for FACS isolation of EYFP-labeled acinar cells; this will be applied in the *Ptf1a* cKO/caerulein model to identify changes in gene expression.

Our major activities in this period have therefore focused mainly on establishing proper experimental paradigms, including training of research personnel; we have underspent our award, as a result, in anticipation of completing this work in the next 6-12 months.

What opportunities for training and professional development has the project provided?

If the project was not intended to provide training and professional development opportunities or there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe opportunities for training and professional development provided to anyone who worked on the project or anyone who was involved in the activities supported by the project. “Training” activities are those in which individuals with advanced professional skills and experience assist others in attaining greater proficiency. Training activities may include, for example, courses or one-on-one work with a mentor. “Professional development” activities result in increased knowledge or skill in one’s area of expertise and may include workshops, conferences, seminars, study groups, and individual study. Include participation in conferences, workshops, and seminars not listed under major activities.

Most of this project is being undertaken by a graduate student in the Murtaugh lab, Diane Hernandez, who has been working closely with the PI over the past year to develop her expertise in mouse and pancreas biology. (Her previous experimental background was in *Drosophila*.) She has presented her work in progress to the department, at annual retreats and student research seminars. Ms. Hernandez has also received training in mentorship, having been joined in her project by two undergraduate lab aides whose work she supervises. Finally, in the past year Ms. Hernandez was awarded a competitive spot on a campus training grant (Training Program in Genetics, NIH 5T32GM007464), which is now supporting her stipend. This has contributed as well to our underspent budget, as we had previously budgeted for her support under this award.

How were the results disseminated to communities of interest?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe how the results were disseminated to communities of interest. Include any outreach activities that were undertaken to reach members of communities who are not usually aware of these project activities, for the purpose of enhancing public understanding and increasing interest in learning and careers in science, technology, and the humanities.

The PI has presented findings related to chronic pancreatitis in *Ptf1a* cKO mice, and discussed the potential for rescue by *Ptf1a* re-expression or FGF21 treatment, at meetings including the Gordon Research Conference on Pancreatic Diseases (2019) and seminars including Baylor College of Medicine (2019). No other public outreach per se has been undertaken yet.

What do you plan to do during the next reporting period to accomplish the goals?

If this is the final report, state “Nothing to Report.”

Describe briefly what you plan to do during the next reporting period to accomplish the goals and objectives.

Aim 1. Re-express *Ptf1a* after establishing chronic pancreatitis in the *Ptf1a* cKO/caerulein model (Fig. 1D), using a DOX-dependent *tetO-Ptf1a* transgene, and determine if this is sufficient to resolve inflammation and injury. We have recently published on the efficacy of this transgene:

Krah et al. [2019] *Developmental Cell* 50: 744-754; doi.org/10.1016/j.devcel.2019.07.012t

Aim 2. Administer FGF21 after establishing chronic pancreatitis in the *Ptf1a* cKO/caerulein model, determine if this is sufficient to resolve inflammation and injury.

Aim 3. FACS isolate dedifferentiated acinar cells from the *Ptf1a* cKO/caerulein model, perform RNA-seq to identify changes in gene expression relative to controls. Using antibodies against select upregulated gene products in mouse, analyze whether similar upregulation occurs in human chronic pancreatitis.

4. **IMPACT:** *Describe distinctive contributions, major accomplishments, innovations, successes, or any change in practice or behavior that has come about as a result of the project relative to:*

What was the impact on the development of the principal discipline(s) of the project?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe how findings, results, techniques that were developed or extended, or other products from the project made an impact or are likely to make an impact on the base of knowledge, theory, and research in the principal disciplinary field(s) of the project. Summarize using language that an intelligent lay audience can understand (Scientific American style).

At this point we have not achieved major accomplishments. Our preliminary data has been well-received in the field, and we anticipate that this work will make a splash comparable to our recent *Developmental Cell* paper on *Ptf1a* and pancreatic cancer.

What was the impact on other disciplines?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe how the findings, results, or techniques that were developed or improved, or other products from the project made an impact or are likely to make an impact on other disciplines.

Nothing to report.

What was the impact on technology transfer?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe ways in which the project made an impact, or is likely to make an impact, on commercial technology or public use, including:

- *transfer of results to entities in government or industry;*
- *instances where the research has led to the initiation of a start-up company; or*
- *adoption of new practices.*

Nothing to report.

What was the impact on society beyond science and technology?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe how results from the project made an impact, or are likely to make an impact, beyond the bounds of science, engineering, and the academic world on areas such as:

- *improving public knowledge, attitudes, skills, and abilities;*
- *changing behavior, practices, decision making, policies (including regulatory policies), or social actions; or*
- *improving social, economic, civic, or environmental conditions.*

Nothing to report.

- 5. CHANGES/PROBLEMS:** *The PD/PI is reminded that the recipient organization is required to obtain prior written approval from the awarding agency grants official whenever there are significant changes in the project or its direction. If not previously reported in writing, provide the following additional information or state, “Nothing to Report,” if applicable:*

Changes in approach and reasons for change

Describe any changes in approach during the reporting period and reasons for these changes. Remember that significant changes in objectives and scope require prior approval of the agency.

Nothing to report.

Actual or anticipated problems or delays and actions or plans to resolve them

Describe problems or delays encountered during the reporting period and actions or plans to resolve them.

We encountered serious delays in this work, as noted above, owing primarily to turnover in personnel. While keeping spending of the award to a minimum, we have focused our efforts on establishing highly-reproducible conditions for the *Ptfla* cKO/caerulein model of chronic pancreatitis. This has been successful, and work is going forward on the key experiments.

Changes that had a significant impact on expenditures

Describe changes during the reporting period that may have had a significant impact on expenditures, for example, delays in hiring staff or favorable developments that enable meeting objectives at less cost than anticipated.

The technician who was expected to work on this project left the lab, for family reasons, and has not yet been replaced; in addition, graduate student Ms. Diane Hernandez is now supported by an NIH training grant, which means that her salary has not been charged to the grant. Overall spending has been very constrained, as we have been focusing on experimental optimization rather than the large scale of mouse breeding and analysis anticipated in our proposal.

Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents

Describe significant deviations, unexpected outcomes, or changes in approved protocols for the use or care of human subjects, vertebrate animals, biohazards, and/or select agents during the reporting period. If required, were these changes approved by the applicable institution committee (or equivalent) and reported to the agency? Also specify the applicable Institutional Review Board/Institutional Animal Care and Use Committee approval dates.

Significant changes in use or care of human subjects

Not applicable (no human subjects).

Significant changes in use or care of vertebrate animals

No changes. Our current animal protocol, 19-10003, was approved by the University of Utah IACUC on October 22, 2019

Significant changes in use of biohazards and/or select agents

Not applicable (no biohazards or select agents).

6. PRODUCTS: *List any products resulting from the project during the reporting period. If there is nothing to report under a particular item, state “Nothing to Report.”*

- **Publications, conference papers, and presentations**

Report only the major publication(s) resulting from the work under this award.

Journal publications. *List peer-reviewed articles or papers appearing in scientific, technical, or professional journals. Identify for each publication: Author(s); title; journal; volume; year; page numbers; status of publication (published; accepted, awaiting publication; submitted, under review; other); acknowledgement of federal support (yes/no).*

Nothing to report.

Books or other non-periodical, one-time publications. *Report any book, monograph, dissertation, abstract, or the like published as or in a separate publication, rather than a periodical or series. Include any significant publication in the proceedings of a one-time conference or in the report of a one-time study, commission, or the like. Identify for each one-time publication: author(s); title; editor; title of collection, if applicable; bibliographic information; year; type of publication (e.g., book, thesis or dissertation); status of publication (published; accepted, awaiting publication; submitted, under review; other); acknowledgement of federal support (yes/no).*

Nothing to report.

Other publications, conference papers and presentations. *Identify any other publications, conference papers and/or presentations not reported above. Specify the status of the publication as noted above. List presentations made during the last year (international, national, local societies, military meetings, etc.). Use an asterisk (*) if presentation produced a manuscript.*

“Ptf1a and acinar cell programming, reprogramming and re-reprogramming.” Gordon Research Conference on Pancreatic Diseases, June 20, 2019. Invited presentation by Dr. Murtaugh.

- **Website(s) or other Internet site(s)**

List the URL for any Internet site(s) that disseminates the results of the research activities. A short description of each site should be provided. It is not necessary to include the publications already specified above in this section.

Nothing to report.

- **Technologies or techniques**

Identify technologies or techniques that resulted from the research activities. Describe the technologies or techniques were shared.

Nothing to report.

- **Inventions, patent applications, and/or licenses**

Identify inventions, patent applications with date, and/or licenses that have resulted from the research. Submission of this information as part of an interim research performance progress report is not a substitute for any other invention reporting required under the terms and conditions of an award.

Nothing to report.

- **Other Products**

Identify any other reportable outcomes that were developed under this project. Reportable outcomes are defined as a research result that is or relates to a product, scientific advance, or research tool that makes a meaningful contribution toward the understanding, prevention, diagnosis, prognosis, treatment and /or rehabilitation of a disease, injury or condition, or to improve the quality of life. Examples include:

- *data or databases;*
- *physical collections;*
- *audio or video products;*
- *software;*
- *models;*
- *educational aids or curricula;*
- *instruments or equipment;*
- *research material (e.g., Germplasm; cell lines, DNA probes, animal models);*
- *clinical interventions;*
- *new business creation; and*
- *other.*

Nothing to report.

7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

What individuals have worked on the project?

Provide the following information for: (1) PDs/PIs; and (2) each person who has worked at least one person month per year on the project during the reporting period, regardless of the source of compensation (a person month equals approximately 160 hours of effort). If information is unchanged from a previous submission, provide the name only and indicate “no change”.

Name: Lewis Charles Murtaugh
Project Role: PI
Researcher Identifier (e.g. ORCID ID): LCMURTAUGH (NIH eCommons)
Nearest person month worked: 3
Contribution to Project: Designed and led project including assistance with training and trouble-shooting.
Funding Support: National Institutes of Health (2R01DK061220-11A1, 5R01CA194941-04)

Name: Diane Hernandez
Project Role: graduate student
Researcher Identifier (e.g. ORCID ID): DHERNANDEZ37 (NIH eCommons)
Nearest person month worked: 6
Contribution to Project: Mouse breeding and experimentation.
Funding Support: Training Program in Genetics, NIH 5T32GM007464

Name: Julie Ann Straley
Project Role: technician
Researcher Identifier (e.g. ORCID ID): n/a
Nearest person month worked: 3
Contribution to Project: Mouse breeding.
Funding Support: National Institutes of Health (2R01DK061220-11A1, 5R01CA194941-04)

Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

If the active support has changed for the PD/PI(s) or senior/key personnel, then describe what the change has been. Changes may occur, for example, if a previously active grant has closed and/or if a previously pending grant is now active. Annotate this information so it is clear what has changed from the previous submission. Submission of other support information is not necessary for pending changes or for changes in the level of effort for active support reported previously. The awarding agency may require prior written approval if a change in active other support significantly impacts the effort on the project that is the subject of the project report.

Nothing to Report.

What other organizations were involved as partners?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe partner organizations – academic institutions, other nonprofits, industrial or commercial firms, state or local governments, schools or school systems, or other organizations (foreign or domestic) – that were involved with the project. Partner organizations may have provided financial or in-kind support, supplied facilities or equipment, collaborated in the research, exchanged personnel, or otherwise contributed.

Provide the following information for each partnership:

Organization Name:

Location of Organization: (if foreign location list country)

Partner’s contribution to the project (identify one or more)

- *Financial support;*
- *In-kind support (e.g., partner makes software, computers, equipment, etc., available to project staff);*
- *Facilities (e.g., project staff use the partner’s facilities for project activities);*
- *Collaboration (e.g., partner’s staff work with project staff on the project);*
- *Personnel exchanges (e.g., project staff and/or partner’s staff use each other’s facilities, work at each other’s site); and*
- *Other.*

Nothing to Report.

8. SPECIAL REPORTING REQUIREMENTS

COLLABORATIVE AWARDS: *For collaborative awards, independent reports are required from BOTH the Initiating Principal Investigator (PI) and the Collaborating/Partnering PI. A duplicative report is acceptable; however, tasks shall be clearly marked with the responsible PI and research site. A report shall be submitted to <https://ers.amedd.army.mil> for each unique award.*

QUAD CHARTS: *If applicable, the Quad Chart (available on <https://www.usamraa.army.mil>) should be updated and submitted with attachments.*

9. **APPENDICES:** *Attach all appendices that contain information that supplements, clarifies or supports the text. Examples include original copies of journal articles, reprints of manuscripts and abstracts, a curriculum vitae, patent applications, study questionnaires, and surveys, etc.*