

AWARD NUMBER: W81XWH-19-1-0855

TITLE: Therapeutic Targeting of FLCN-Deficient Renal Cancers

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CONTRACTING ORGANIZATION: Massachusetts General Hospital

REPORT DATE: OCTOBER 2020

TYPE OF REPORT: Annual Progress Report

PREPARED FOR: U.S. Army Medical Research and Development Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
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REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

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1. REPORT DATE OCTOBER 2020			2. REPORT TYPE Annual		3. DATES COVERED 30SEPT2019 -29SEPT2020	
4. TITLE AND SUBTITLE Therapeutic Targeting of FLCN-Deficient Renal Cancers					5a. CONTRACT NUMBER	
					5b. GRANT NUMBER W81XWH-19-1-0855	
					5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Othon Iliopoulos, MD E-Mail: ILIOPOUL@HELIX.MGH.HARVARD.EDU					5d. PROJECT NUMBER	
					5e. TASK NUMBER	
					5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Massachusetts General Hospital 55 Fruit Street Boston, MA 021224-2696					8. PERFORMING ORGANIZATION REPORT	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Development Command Fort Detrick, Maryland 21702-5012					10. SPONSOR/MONITOR'S ACRONYM(S)	
					11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited						
13. SUPPLEMENTARY NOTES						
14. ABSTRACT The goals of this proposal are: 1) to identify the cellular phospho-proteome regulated by FLCN, 2) to discover the mechanisms by which FLCN suppresses the non-canonical translational initiation of a specific subset of mRNAs, and 3) to assign FLCN functions to specific protein domains. So far we profiled the cellular proteome changes regulated by FLCN in vitro and we gained major insights into the mechanism by which FLCN regulates protein translation.						
15. SUBJECT TERMS Folliculin, GTPase activity, Rab7A, Phosphoproteome, Receptor Tyrosine Kinase activity, Tumor suppressor gene.						
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON	
a. REPORT	b. ABSTRACT	c. THIS PAGE			19b. TELEPHONE NUMBER <i>(include area code)</i>	
Unclassified	Unclassified	Unclassified	Unclassified	15		

Standard Form 298 (Rev. 8-98)
Prescribed by ANSI Std. Z39-18

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1. INTRODUCTION: *Narrative that briefly (one paragraph) describes the subject, purpose and scope of the research.*

Folliculin (FLCN) is a tumor suppressor gene linked to the development of renal cell carcinoma. We showed before that FLCN acts as a GAP protein for Rab7A and regulates trafficking and therefore activity of EGFR. Our proposal's goal is to show that (1) FLCN regulates not only EGFR, but a panel of cell surface receptor tyrosine kinases in a way similar to EGFR, because the "internalization" of these kinases is a general mechanism of regulation. (2) We showed that FLCN suppresses protein translation and it binds to two translation-promoting factors that are GTPases. We therefore propose to take a system biology approach in order to profile all the kinases that are regulated by FLCN and to evaluate which kinases can be used to target FLCN-driven, rare RCCs. We also propose a series of experiments that will uncover the biochemical details of how FLCN suppresses protein translation. (3) The third goal is to use mutants of FLCN to find out if all these functions depend on different parts of the protein and can be separated from each other. Such a "distribution" of functions (called "domains") will help understand which of these functions are important for tumor suppression.

2. KEYWORDS: *Provide a brief list of keywords (limit to 20 words).*

Folliculin, GTPase activity, Rab7A, Phosphoproteome, Receptor Tyrosine Kinase activity, Tumor suppressor gene.

3. ACCOMPLISHMENTS: *The PI is reminded that the recipient organization is required to obtain prior written approval from the awarding agency grants official whenever there are significant changes in the project or its direction.*

What were the major goals of the project?

List the major goals of the project as stated in the approved SOW. If the application listed milestones/target dates for important activities or phases of the project, identify these dates and show actual completion dates or the percentage of completion.

Specific Aim 1: To determine the receptor tyrosine kinases (RTKs) and the cellular phosphoproteome changes regulated by FLCN.

Subtask 1: FLCN-dependent global phosphoproteome changes in cells stimulated with growth factors. Phospho-proteomic and proteomic analysis. Cell lines used: UOK257, FTC-133, infected with wild type FLCN, empty vector control or FLCN mutants.

Subtask 2: FLCN-dependent global phosphoproteome changes in cells stimulated with amino acids. Phospho-proteomic and proteomic analysis. Cell lines used: UOK257, FTC-133, infected with wild type FLCN, empty vector control or FLCN mutants

Specific Aim 2: To fully characterize the biochemical mechanism by which FLCN suppresses initiation of protein translation and identify the subclass of mRNAs regulated by FLCN.

Specific Aim 3: To genetically dissect FLCN functions by using a panel of post-translational modification (PTM) and tumor-associated FLCN mutations.

Subtask 1: Generation of tumor-associated FLCN mutants that lead to expression of stable and detectable FLCN mutants

Subtask 2: Generation of single and compound phospho-mimetic and phospho-inactivating FLCN mutants as well as scanning mutagenesis of FLCN protein.

What was accomplished under these goals?

For this reporting period describe: 1) major activities; 2) specific objectives; 3) significant results or key outcomes, including major findings, developments, or conclusions (both positive and negative); and/or 4) other achievements. Include a discussion of stated goals not met. Description shall include pertinent data and graphs in sufficient detail to explain any significant results achieved. A succinct description of the methodology used shall be provided. As the project progresses to completion, the emphasis in reporting in this section should shift from reporting activities to reporting accomplishments.

SPECIFIC AIM 1: Our preliminary data indicated that FLCN regulates several receptor tyrosine kinases (RTKs) through its function as a Rab7A GAP and has a global effect on the cellular phosphoproteome. *We proposed to use proteomic approaches to map the FLCN-regulated phosphoproteomic changes in cells stimulated by growth factors and/or amino acids.* This knowledge will allow us to validate critical nodes of the phosphoproteomic changes as therapeutic targets.

In SOW we proposed to use the FLCN-null UOK257 and FTC-133 cell lines and create isogenic lines in which wild type FLCN was reintroduced. *We completed this goal* and we used FLCN -/- and +/- cell lines for the experiments described in subtask 1 and subtask 2 of Aim 1. We stimulated the cells lines with amino acids or serum containing growth factors and obtained a phosphoproteome map of the cells. This is achieved through a new phospho-proteomics strategy developed in the Haas laboratory that combines two phospho-peptide fragmentation methods – collision-induced dissociation (CID) and higher-collision-induced dissociation – to increase the sensitivity of multiplexed phospho-proteomics measurements by two-fold (described in our proposal). *We therefore achieved the goal outlined in subtasks 1 and subtask 2 in obtaining a map of phosphoproteins regulated by FLCN.*

The attached .xls files (FLCN proteome and FLCN phosphoproteome) are the raw data of the proteome changes and phosphoproteome changes following serum stimulation of three isogenic cell lines: UOK257 FLCN-/- cells, UOK257 FLCN-replete cells, and UOK257 cells infected with the tumor causing FLCN-C9 mutant. The importance of these experiments consists in the discovery that FLCN regulates several receptor tyrosine kinases (RTKs) and their subsequent signaling pathways implicated in tumorigenesis. Future experiments based on this award will ask how to optimally block these pathways with existing drugs, a goal which is beyond the described scope of the award, but very important for development of medical therapies in FLCN-/- renal cell cancers.

To confirm these results in an orthogonal manner and test their conservation through evolution we collaborated with the laboratory of Mo Motamedi, in the MGH Cancer Center. We purified and characterized the Schizosaccharomyces pombe Bhd-Fnp Complex (BFC) and its cellular interactors. We showed that BFC complex physically interacts and regulates the highly conserved peptide transmembrane transporter Ptr2, the phosphoribosylformylglycinamide synthase Ade3, and the V-ATPase complex. These are novel amino acid-dependent regulators of TORC1. BFC mutants exhibited a slower TORC1 repression and proliferate faster than wild-type cells following amino acid starvation. Therefore, we showed that the mammalian function of FLCN on mTORC1 through regulation of transmembrane RTKs is conserved evolutionarily. This work is now accepted in *iScience* (*Calvo et al. The fission yeast FLCN/FNIP complex augments TORC1 repression or activation in response to amino acid availability). The manuscript is attached.*

SPECIFIC AIM 2: We showed that FLCN localizes to the polysomes, associates with factors regulating the initiation of protein translation and inhibits eIF2 γ and EIF5B complex formation with tRNA-Met_i. The FLCN C-terminal domain, which is deleted by tumor-associated FLCN mutations, is necessary for the interaction of FLCN with eIF2 γ and EIF5B. Reintroduction of wild type but not a C-terminus FLCN mutant into FLCN^{-/-} cells results in suppression of serum or amino acid stimulated protein translation, independently of mTORC1 activity. Our data provide insights into a novel mechanism of cell growth restriction by FLCN (*We therefore achieved the goal outlined in Aim 2, subtasks 1, 2 and 3*).

We attach the manuscript “HUMAN FOLLICULIN TUMOR SUPPRESSOR PROTEIN BINDS TO TRANSLATION INITIATION FACTORS eIF2 γ AND EIF5B AND SUPPRESSES PROTEIN SYNTHESIS” by Schneider et al, which describes the experiments corresponding to Aim 2A. This manuscript is currently under review.

Work that remains to be done: In our proposal we describe how we will identify the RNAs regulated by FLCN (Aim 2B). We have isolated the polysome fractions shown in Figure 2 of the Schneider et al manuscript, and we extracted RNA. We will now complete Aim 2B (Aim 2, subtasks 4 and 5) in order to discover these RNAs.

SPECIFIC AIM 3: The goal here is the dissection of FLCN functions by generating a panel of tumor-associated, truncation, phosphor-inactivating and phosphor-mimetic FLCN mutants (Subtasks 1 and 2).

We accomplished the first step of this goal. *We now have generated a panel of phospho-inactivating and phosphomimetic FLCN mutants. We also generated a panel of tumor associated point mutants.*

There raw data about these mutants, their stability and migration are shown in: Manuscript by Schneider et al, Figures 1F and G (truncation mutants and tumor associated) Lavolette et al, Figure 1a,1b, and 3a (Phosphomimetic, phosphor-inactivating and tumor associated mutants).

We plan in the next steps towards completion of this aim, to introduce the FLCN ^{-/-} RCC cells harboring these mutants in nude mice and test their effect on tumor suppression.

What opportunities for training and professional development has the project provided?

If the project was not intended to provide training and professional development opportunities or there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe opportunities for training and professional development provided to anyone who worked on the project or anyone who was involved in the activities supported by the project. “Training” activities are those in which individuals with advanced professional skills and experience assist others in attaining greater proficiency. Training activities may include, for example, courses or one-on-one work with a mentor. “Professional development” activities result in increased

knowledge or skill in one's area of expertise and may include workshops, conferences, seminars, study groups, and individual study. Include participation in conferences, workshops, and seminars not listed under major activities.

Dr. Iliopoulos trained the following people that so far worked on the FLCN project generating the preliminary results of this proposal, the data included in the manuscripts currently submitted for publication and the data included in the current progress report.

Laura Laviolette, PhD. Obtained her PhD from University of Vancouver in Canada, completed her post-doctoral training with Dr. Iliopoulos, worked on FLCN and she is now a senior scientist doing oncology research at Vertex Pharmaceutical.

Meike Schneider, MD completed her post-doctoral training with Dr. Iliopoulos after graduating from the Department of Urology, Medical Center Johannes Gutenberg University, Mainz, Germany and she is now a senior scientist at Bayer Oncology. She is the lead author in the paper describing the effect of FLCN on protein translation.

Katia Dinkelborg, MD completed her Diploma Thesis with Dr. Iliopoulos, graduated from University of Hannover Medical School in Germany and she is now working as a research fellow in the University of Hannover Department of Hepatology.

Ravi Sundaram, BS, obtained his Bachelor of Sciences from Northeastern University, Boston, MA and worked as a Research Technician for 3 years at the Iliopoulos Lab. He is now enrolled in the University of Sydney Medical School, in Sydney, Australia.

How were the results disseminated to communities of interest?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe how the results were disseminated to communities of interest. Include any outreach activities that were undertaken to reach members of communities who are not usually aware of these project activities, for the purpose of enhancing public understanding and increasing interest in learning and careers in science, technology, and the humanities.

The main avenue of data dissemination is publication of our data. In addition, we generated several collaborations, attracting colleagues to study the basic mechanism of tumor suppression by FLCN (as evidenced by our publication record).

The following manuscripts are submitted for publication and are currently under review. These manuscripts were supported by the DOD award at hand.

1) HUMAN FOLLICULIN TUMOR SUPPRESSOR PROTEIN BINDS TO TRANSLATION INITIATION FACTORS eIF2 α AND EIF5B AND SUPPRESSES PROTEIN SYNTHESIS

(Meike Schneider, Katja Dinkelbor, Syed I.A. Bukhari, Samuel S Truesdell, Vera A. Pisareva, Andrey V. Pisarev, Shobha Vasudevan and Othon Iliopoulos).

2) The fission yeast FLCN/FNIP complex augments TORC1 repression or activation in response to amino acid availability

(Isabel A. Calvo, Shalini Sharma, Joao A. Paulo, Alexander Gulka, Andras Boeszoermyeni, Jingyu Zhang, Jose M. Lombana, Christina M. Palmieri¹, Laura A. Laviolette, Haribabu Arthanari, Steven P. Gygi, Othon Iliopoulos and Mo Motamedi).

Describe briefly what you plan to do during the next reporting period to accomplish the goals and objectives.

Within the next reporting period (12 months) we plan:

- 1) To validate the phosphoproteomic targets we discovered in Aim 1. Our proposal described how we will do this (Aim 1. Subtask 3).
- 2) To identify the mRNAs regulated by FLCN, as described in our proposal (Aim 2, subtasks 4 and 5).
- 3) We will assay the effect of the panel of FLCN mutations in the xenograft tumor suppressor assay Aim 3, subtask 3).

4. **IMPACT:** Describe distinctive contributions, major accomplishments, innovations, successes, or any change in practice or behavior that has come about as a result of the project relative to:

What was the impact on the development of the principal discipline(s) of the project?

If there is nothing significant to report during this reporting period, state "Nothing to Report."

Describe how findings, results, techniques that were developed or extended, or other products from the project made an impact or are likely to make an impact on the base of knowledge, theory, and research in the principal disciplinary field(s) of the project. Summarize using language that an intelligent lay audience can understand (Scientific American style).

We are dissecting the function of FLCN tumor suppressor gene. Our goal is to discover what are the critical biochemical events that are deregulated in cells when FLCN is inactivated. To this end we are linking genetic analysis of FLCN to the biochemical events that are regulated by this tumor suppressor gene, namely changes in the cellular phosphoproteome and in protein translation. During the first year of the project we generated the required reagents and we profiled the phosphoproteome changes. This latter discovery is significant insight into the function of FLCN.

What was the impact on other disciplines?

If there is nothing significant to report during this reporting period, state "Nothing to Report."

Describe how the findings, results, or techniques that were developed or improved, or other products from the project made an impact or are likely to make an impact on other disciplines.

Nothing to Report

What was the impact on technology transfer?

If there is nothing significant to report during this reporting period, state "Nothing to Report."

Describe ways in which the project made an impact, or is likely to make an impact, on commercial technology or public use, including:

- *transfer of results to entities in government or industry;*
- *instances where the research has led to the initiation of a start-up company; or*

- *adoption of new practices.*

Nothing to Report

What was the impact on society beyond science and technology?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe how results from the project made an impact, or are likely to make an impact, beyond the bounds of science, engineering, and the academic world on areas such as:

- *improving public knowledge, attitudes, skills, and abilities;*
- *changing behavior, practices, decision making, policies (including regulatory policies), or social actions; or*
- *improving social, economic, civic, or environmental conditions.*

Nothing to Report

- 5. CHANGES/PROBLEMS:** *The PD/PI is reminded that the recipient organization is required to obtain prior written approval from the awarding agency grants official whenever there are significant changes in the project or its direction. If not previously reported in writing, provide the following additional information or state, “Nothing to Report,” if applicable:*

Changes in approach and reasons for change

Describe any changes in approach during the reporting period and reasons for these changes. Remember that significant changes in objectives and scope require prior approval of the agency.

Nothing to Report

Actual or anticipated problems or delays and actions or plans to resolve them

Describe problems or delays encountered during the reporting period and actions or plans to resolve them.

We do not anticipate a delay in going forward.

As it is well known this has been a year of hardship for all. Our laboratory was closed for 4 months due to COVID pandemic. The PI was recruited to provide clinical care of COVID patients. In addition he was infected by COVID19 and had a prolonged course of illness. These issues were *recently* resolved and we therefore hope to move forward without any further obstacles. We are now recruiting a post doctoral fellow to continue work on FLCN project.

Changes that had a significant impact on expenditures

Describe changes during the reporting period that may have had a significant impact on expenditures, for example, delays in hiring staff or favorable developments that enable meeting objectives at less cost than anticipated.

Nothing to Report

Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents

Describe significant deviations, unexpected outcomes, or changes in approved protocols for the use or care of human subjects, vertebrate animals, biohazards, and/or select agents during the reporting period. If required, were these changes approved by the applicable institution committee (or equivalent) and reported to the agency? Also specify the applicable Institutional Review Board/Institutional Animal Care and Use Committee approval dates.

Significant changes in use or care of human subjects

N/A

Significant changes in use or care of vertebrate animals

None expected

Significant changes in use of biohazards and/or select agents

None expected

6. PRODUCTS: *List any products resulting from the project during the reporting period. If there is nothing to report under a particular item, state “Nothing to Report.”*

• **Publications, conference papers, and presentations**

Report only the major publication(s) resulting from the work under this award.

Journal publications. *List peer-reviewed articles or papers appearing in scientific, technical, or professional journals. Identify for each publication: Author(s); title; journal; volume: year; page numbers; status of publication (published; accepted, awaiting publication; submitted, under review; other); acknowledgement of federal support (yes/no).*

Manuscripts submitted as outlined above.

Books or other non-periodical, one-time publications. *Report any book, monograph, dissertation, abstract, or the like published as or in a separate publication, rather than a periodical or series. Include any significant publication in the proceedings of a one-time conference or in the report of a one-time study, commission, or the like. Identify for each one-time publication: author(s); title; editor; title of collection, if applicable; bibliographic information; year; type of publication (e.g., book, thesis or dissertation); status of publication (published; accepted, awaiting publication; submitted, under review; other); acknowledgement of federal support (yes/no).*

Nothing to Report

Other publications, conference papers and presentations. *Identify any other publications, conference papers and/or presentations not reported above. Specify the status of the publication as noted above. List presentations made during the last year (international, national, local societies, military meetings, etc.). Use an asterisk (*) if presentation produced a manuscript.*

Nothing to Report

- **Website(s) or other Internet site(s)**

List the URL for any Internet site(s) that disseminates the results of the research activities. A short description of each site should be provided. It is not necessary to include the publications already specified above in this section.

Nothing to Report

- **Technologies or techniques**

Identify technologies or techniques that resulted from the research activities. Describe the technologies or techniques were shared.

Dr. Wilhelm Haas laboratory devised a multiplexed mass spectrometry-based phospho-proteomics to quantitatively map phosphorylation level changes across tumor samples at a depth of 20,000 to 30,000 distinct phosphorylation events. This is enabled through a new phospho-proteomics strategy developed in the Haas laboratory that combines two phospho-peptide fragmentation methods – collision-induced dissociation (CID) and higher-collision-induced dissociation – to increase the sensitivity of multiplexed phospho-proteomics measurements by two-fold. We used this technique in Aim 1.

- **Inventions, patent applications, and/or licenses**

Identify inventions, patent applications with date, and/or licenses that have resulted from the research. Submission of this information as part of an interim research performance

progress report is not a substitute for any other invention reporting required under the terms and conditions of an award.

Nothing to Report

• **Other Products**

Identify any other reportable outcomes that were developed under this project. Reportable outcomes are defined as a research result that is or relates to a product, scientific advance, or research tool that makes a meaningful contribution toward the understanding, prevention, diagnosis, prognosis, treatment and /or rehabilitation of a disease, injury or condition, or to improve the quality of life. Examples include:

- *data or databases;*
- *physical collections;*
- *audio or video products;*
- *software;*
- *models;*
- *educational aids or curricula;*
- *instruments or equipment;*
- *research material (e.g., Germplasm; cell lines, DNA probes, animal models);*
- *clinical interventions;*
- *new business creation; and*
- *other.*

Data generated during this award period are used for teaching of Harvard Medical School's BBS Program students in the lecture "Genetics of Kidney Cancer".

7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

What individuals have worked on the project?

Othon Iliopoulos
PD/PI
1.8 CM

Tupa Basuroy
Post Doc
12 CM

Wilhelm Haas
Co-Investigator
1.2 CM

Shobha Vasudevan
Co-Investigator
0.6 CM

Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

If the active support has changed for the PD/PI(s) or senior/key personnel, then describe what the change has been. Changes may occur, for example, if a previously active grant has closed and/or if a previously pending grant is now active. Annotate this information so it is clear what has changed from the previous submission. Submission of other support information is not necessary for pending changes or for changes in the level of effort for active support reported previously. The awarding agency may require prior written approval if a change in active other support significantly impacts the effort on the project that is the subject of the project report.

No Change

What other organizations were involved as partners?

If there is nothing significant to report during this reporting period, state “Nothing to Report.”

Describe partner organizations – academic institutions, other nonprofits, industrial or commercial firms, state or local governments, schools or school systems, or other organizations (foreign or domestic) – that were involved with the project. Partner organizations may have provided financial or in-kind support, supplied facilities or equipment, collaborated in the research, exchanged personnel, or otherwise contributed.

Provide the following information for each partnership:

Organization Name:

Location of Organization: (if foreign location list country)

Partner’s contribution to the project (identify one or more)

- *Financial support;*
- *In-kind support (e.g., partner makes software, computers, equipment, etc., available to project staff);*
- *Facilities (e.g., project staff use the partner’s facilities for project activities);*
- *Collaboration (e.g., partner’s staff work with project staff on the project);*
- *Personnel exchanges (e.g., project staff and/or partner’s staff use each other’s facilities, work at each other’s site); and*
- *Other.*

Nothing to report

8. SPECIAL REPORTING REQUIREMENTS

COLLABORATIVE AWARDS: *For collaborative awards, independent reports are required from BOTH the Initiating Principal Investigator (PI) and the Collaborating/Partnering PI. A duplicative report is acceptable; however, tasks shall be clearly marked with the responsible PI and research site. A report shall be submitted to <https://ers.amedd.army.mil> for each unique award.*

QUAD CHARTS: *If applicable, the Quad Chart (available on <https://www.usamraa.army.mil>) should be updated and submitted with attachments.*

- 9. APPENDICES:** *Attach all appendices that contain information that supplements, clarifies or supports the text. Examples include original copies of journal articles, reprints of manuscripts and abstracts, a curriculum vitae, patent applications, study questionnaires, and surveys, etc.*