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**TITLE: Identifying Different Metabolic Subtypes of Prostate Cancer for Early Therapeutic Assessment**

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<b>14. ABSTRACT</b>  In the second reporting project, we have made substantial progress on Specific Aim I and II and we have started working on the most impactful aim of the project, Specific Aim III on imaging to overcome Enzalutamide resistance by targeting Monocarboxylate Transporter (MCT) pathway. Our preliminary HP-MR data on one of the AR positive Enzalutamide resistant PDX models (274-4) shows that MCT inhibition with the clinical drug, Syrosingopine can overcome resistance to Enzalutamide. Two manuscripts are currently under review and a conference abstract that has resulted in an invite for oral presentation at the 25th International Symposium on Radiopharmaceutical Sciences (iSRS) in May 22-26, 2023. The results from the metabolic imaging and other "omics" data from this project is also laying the foundation of an Artificial Intelligence driven project on classifying the different sub-types of prostate cancer at the time of initial diagnosis.					
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## 1. INTRODUCTION:

Patients with advanced prostate cancer (PCa) receive anti-androgens (Enzalutamide) as the first line of treatment. Unfortunately, many patients develop resistance, and the disease relapses with aggressive histology and metastatic castrate-resistant (mCRPC) phenotype. Treatment options for mCRPC patients are limited and continue to pose a significant oncological challenge. There is growing recognition that alterations in cell metabolism and reprogramming of metabolic pathways are key drivers of PCa aggressiveness, progression and eventual resistant to therapy. The overarching goal of this project is to develop metabolic imaging to target treatment strategies of different metabolic sub-types of PCa. Recent studies from our group have demonstrated that the tumor's metabolic profile that impacts anti-androgens response can be identified early on by using hyperpolarized Magnetic Resonance Imaging (HP-MRI). We also discovered that Monocarboxylate Transporter (MCT) is dysfunctional in mCRPC and may be a viable target for therapeutic intervention. In this research, we will employ HP-MRI and Positron Emission Tomography (PET) to interrogate both glycolysis and carnitine metabolism that is closely linked with fatty acid metabolism in PCa. We will evaluate the treatment response of Enzalutamide in clinically relevant patient derived mouse models of PCa. We will correlate imaging data with metabolomics, immunohistochemistry and transcriptome profile analysis of the *ex vivo* tissue samples to elucidate the metabolic drivers of resistance. In two of the resistant models, we will attempt to restore drug sensitivity by targeting Monocarboxylate Transporter (MCT) pathway and image this transformation from resistance to sensitization by performing HP-MRI and PET. If successful, this research will provide mechanistic insights to advance our knowledge about mCRPC tumors and validate the efficacy of targeting MCT protein for therapeutic benefit.

## 2. KEYWORDS:

prostate cancer, treatment resistance, hyperpolarization,  $^{13}\text{C}$  MR,  $^{18}\text{F}$ -FPIA, PET, MRI, mCRPC, PDX, PCa, MCT, ENO2, metabolic imaging

## 3. ACCOMPLISHMENTS:

**What were the major goals of the project?**

**Specific Aim I:** To perform HP- $^{13}\text{C}$  Pyruvate MRI, [ $^{18}\text{F}$ ]FPIA -PET imaging in parallel in four well characterized patient derived xenograft (PDX) murine models of prostate cancer *in vivo* before and after the antiandrogen therapy with Enzalutamide to image both alteration in glycolytic and fatty acid oxidation pathways after treatment response. (1-15 Months).

Under Specific Aim I, there are four tasks:

**Task 1:** Generation of PDX models: 100% completed

**Task 2:** Synthesize  $^{18}\text{F}$ -Fluoropivalate ([ $^{18}\text{F}$ ]FPIA). 100% completed

**Task 3:**  $^{13}\text{C}$  Pyruvate HP-MRI and [ $^{18}\text{F}$ ]FPIA PET to interrogate both glycolysis and fatty metabolism at the preclinical PET-MRI scanner at the before treatment and three other time points on the 7 day, 14<sup>th</sup> day of treatment and upon resistance. 75% completed

**Task 4:** Harvesting of tumors and storing for NMR spectroscopy, Mass spectrometry, immunohistochemistry and RNA sequencing studies at different time points. **100% completed**

**Specific Aim II:** To elucidate the metabolic drivers of ADT resistance. (9-24 months)

Under Specific Aim II, there are four tasks.

**Task 5:** NMR spectroscopy based metabolomics of flash frozen tissues (four PDX lines; from task 4). **75% completed**

**Task 6:** Mass Spectrometry based metabolomics (four PDX lines; from task 4). **50% completed**

**Task 7:** Transcriptome analysis of Tumors (four PDX lines; from task 4). **25% completed**

**Task 8:** Immunohistochemistry analysis of Tumors (four PDX lines; from task 4). **25% completed**

**Specific Aim III:** To image overcoming the ADT resistance by targeting Monocarboxylate Transporter (MCT) pathway by performing HP-<sup>13</sup>C Pyruvate MRI in two well characterized patient derived xenograft (PDX) murine models of prostate cancer *in vivo*. [<sup>18</sup>F]FPIA -PET imaging, performed in parallel, will inform on correlation of the modulation of glycolysis (via MCT inhibition with the clinical drug, Syrosingopine) with modulation of fatty acid oxidation. (15-36 months)

**Task 9:** Two of the Enzalutamide resistant PDX models. 274-4 and 133 will be employed for imaging experiments. When the SC tumors are about 0.5 cm, the tumor bearing mice will be randomized into two treatment groups (N=20 per PDX sub-type). Mice in Group 1 will receive 10mg/kg body wt Enzalutamide by oral gavage while Group 2 mice will receive Enzalutamide and intraperitoneal injection of Syrosingopine, MCT4 inhibitor (7.5mg/kg body weight). **25% completed**

**Task 10:** Synthesize <sup>18</sup>F-Fluoropivalate ([<sup>18</sup>F]FPIA). **100% completed**

**Task 11:** Simultaneous <sup>13</sup>C Pyruvate HP-MRI and [<sup>18</sup>F]FPIA PET to interrogate both glycolysis and fatty acid oxidation pathways at the preclinical PET-MRI scanner at two time points on the 14<sup>th</sup> and 21<sup>st</sup> day. **25% completed**

**Task 12:** Harvesting of tumors and storing for NMR spectroscopy, Mass spectrometry, immunohistochemistry and RNA sequencing studies at different time points. **20% completed**

**Task 13:** NMR spectroscopy based metabolomics of flash frozen tissues (two PDX lines; from task 12). **20% completed**

**Task 14:** Mass Spectrometry based metabolomics (two PDX lines; from task 12). **Not started**

**Task 15:** Transcriptome analysis of Tumors (two PDX lines; from task 12). **Not started**

**Task 16:** Validation of nominated genes. **Not started**

**Task 17:** Immunohistochemistry analysis of Tumors (two PDX lines; from task 12). **Not started**

**What was accomplished under these goals?**

**A. Major Activities:**

### Aim I, Task 1: Generation of PDX models:

In the proposal, we suggested these following four PDX models for study

274-4 (AR+, Enzalutamide resistant)

133-3 (AR+, Enzalutamide resistant)

183-A (AR+, Enzalutamide sensitive)

170-1 (AR+, Enzalutamide sensitive)

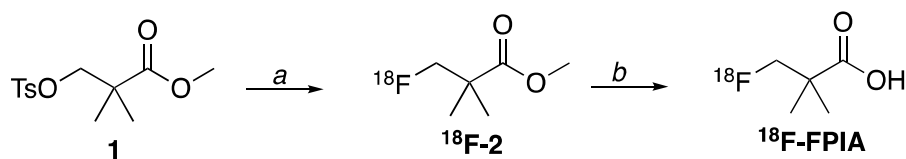
However, we ran into some logistical challenges to generate all PDX models we originally proposed from our PDX core facility as the core ran into several issues in one past one year. Therefore, we have kept performing experiments on the models that are available and fit the overall goals of the project.

### PDX models employed so far in the study

PDX	HP-MR	FPIA-PET	NMR met	MS met	AR	Enzalutamide	phenotype
144	x	x	x	x	negative		
118	x	x	x		negative		
232	x	x	x	x	negative		Adenocarcinoma
337	x	x	x		negative		Neuroendocrine carcinoma
183	x	x	x	x	positive	sensitive	
274	x	x	x	x	positive	resistant	
177	x	x	x		negative		
180	x		x	x	positive	sensitive	
477	x	x	x		positive	resistant	

**Aim I, Task 2:** Radiosynthesis of  $^{18}\text{F}$ -Fluoropivalate ( $^{18}\text{F}$ FPiA) has been streamlined in the last report and the automated radiosynthesis of  $^{18}\text{F}$ -FPiA using the GE TracerLab FX FN™ platform is now routinely performed at MD Anderson Cancer Center. The details were reported in the report last year. I have included a slightly optimized versions in this report as well.

### Radiosynthesis of $^{18}\text{F}$ -Fluoropivalate ( $^{18}\text{F}$ -FPiA).



**Scheme 1** Synthesis of  $^{18}\text{F}$ -FPiA. Reaction conditions: *a*)  $^{18}\text{F}$ -fluoride,  $\text{K}_{222}$ ,  $\text{KHCO}_3$ , DMSO, 105 °C, 10 min; *b*) NaOH, then phosphate buffer.

### Synthesis of precursor methyl 2,2-dimethyl-3-[(4-methylbenzenesulfonyl)oxy]propanoate (**1**)

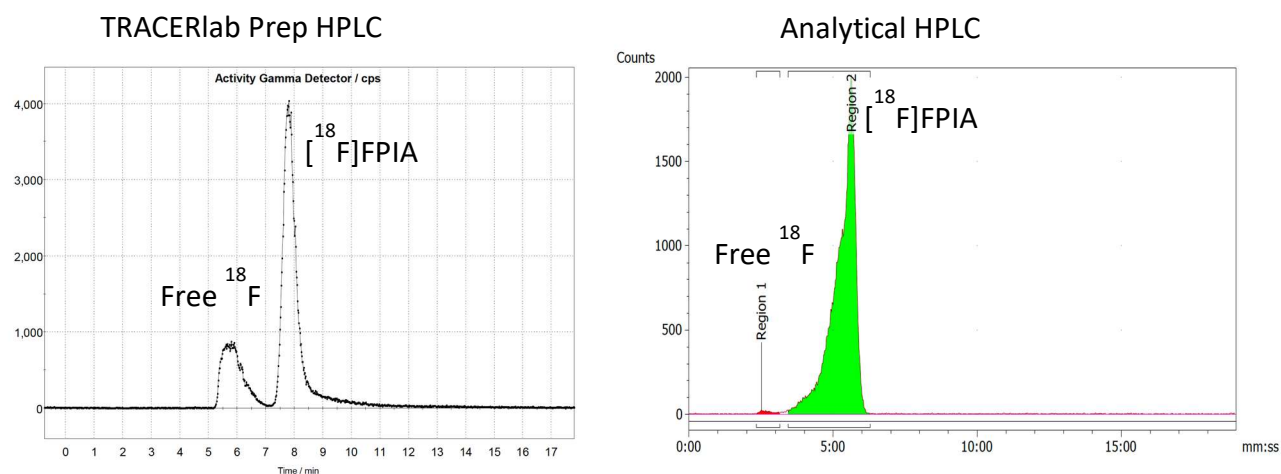
Precursor methyl 2,2-dimethyl-3-[(4-methylbenzenesulfonyl)oxy]propanoate (**1**) was synthesized accordingly with the published procedure (Pisaneschi et al, *Med. Chem. Commun.*, **2013**, *4*, 1350-

1353) and purified with a Biotage® Selekt system using a 25 g column and 7% to 60% ethyl acetate/hexane eluent over 10 min. Precursor **1** was obtained in 70% yield, as a white solid, and spectroscopically characterized to match the published material.

#### Automated radiosynthesis of $^{18}\text{F}$ -FPIA using the GE TracerLab FX FN™ platform.

An aqueous solution of  $^{18}\text{F}$ -fluoride in oxygen-18 enriched water (2 mL) was transferred from the cyclotron target to the warm-cell containing the TracerLab FX FN module (General Electric Healthcare, Münster, Germany). The  $^{18}\text{F}$ -fluoride was trapped on an ion exchange cartridge (pre-conditioned Sep-PAK® Light QMA Cartridge, ABX GmbH, Radeberg, Germany) and eluted into the reactor with a potassium bicarbonate and Kryptofix<sub>222</sub> water/CH<sub>3</sub>CN solution (1 mL; stock solution: 40 mg of KHCO<sub>3</sub>, 240 mg of K<sub>222</sub>, 4 mL of water, 16 mL of CH<sub>3</sub>CN). The solution was dried under vacuum and nitrogen flow at 60 °C for 2 min. Dry CH<sub>3</sub>CN (500 µL) was added and the mixture was azeotropically dried at 120 °C for an additional 3 min. To the reactor containing dry  $^{18}\text{F}$ -fluoride was added precursor **1** (8 mg) in anhydrous DMSO (500 µL) and water (1 µL) and heated at 105 °C for 10 min to produce  $^{18}\text{F}$ -**2**. After cooling to 60 °C, a solution of NaOH (1 mL, 1M) was added to the reactor and heated to 60 °C for 10 min to produce  $^{18}\text{F}$ -FPIA. The reactor was cooled at 40 °C and NaH<sub>2</sub>PO<sub>4</sub> (2.5 mL, 0.5 M) was added. The crude reaction mixture was filtered and transferred to the TracerLab HPLC loading vial. The HPLC injection loop (5 mL) was filled and  $^{18}\text{F}$ -FPIA was purified by semi preparative HPLC (Luna 5 µm C18(2) column, 100 Å, 250 x 10 mm) column with a mobile phase of NaH<sub>2</sub>PO<sub>4</sub> (10 mM) containing 1% EtOH at a flow rate of 4 mL/min. A radioactive peak corresponding to  $^{18}\text{F}$ -FPIA eluted at ca. 6.5 min, which was cut for 30-40 sec via a sterile filter (PALL Acrodisc 0.2 µm) into a sterile vial. Quality control was carried out by analytical radio-HPLC (Agilent 1100, Santa Clara, CA, USA) using an Alltech Econosil C18 column (3.5 µm, 4.6x250 mm) with a 30% to 95% CH<sub>3</sub>CN/water over 19 min gradient. The identity of the radiolabeled compound was confirmed by co-elution with commercially available fluoropivalate (Thermo Fisher Scientific, Waltham, MA).

$^{18}\text{F}$ -FPIA was obtained in 25.4±3.8 (n=9) activity yield (non-decay corrected), >99% radiochemical purity and high UV purity



**Figure 1:** HPLC traces of  $^{18}\text{F}$ -FPIA production in Tracerlab Prep HPLC and analytical HPLC. Analytical HPLC indicated production of  $^{18}\text{F}$ -FPIA > 99% radiochemical purity.

**Aim I, Task 3:** Imaging experiments.

Mice were housed in pathogen-free conditions and cared for in accordance with guidelines set by the AALAC and were supervised by the IACUC at MD Anderson Cancer Center. All procedures are performed using 1.5% isoflurane as the anesthetic. Standard MR sequences will be employed.

**[1-<sup>13</sup>C] Pyruvate HP-MRI:** *In vivo* imaging were performed on a 7T MRI animal scanner situated adjacent to DNP HyperSense polarizer. A solution of 25  $\mu$ L [1-<sup>13</sup>C] pyruvic acid (Isotec, Sigma-Aldrich, St. Louis, MO), 15 mM OX63 trityl radical (GE Healthcare) and 1.5 mM of the gadolinium chelate (ProHance) was polarized at 3.35 T and 1.4 K in the DNP polarizer (HyperSense, Oxford Instruments, UK) for one hour. HP pyruvate was rapidly dissolved in 4 mL superheated alkaline buffer containing 100 mg/L ethylenediaminetetraacetate (EDTA), 40 mM NaOH, 40 mM TRIS buffer, and 30 mM NaCl. The final concentration of injected pyruvate was 80 mM with physiologic pH  $\sim$  7.4. 250  $\mu$ L of HP pyruvate was injected intravenously over a period of 8-10 seconds using a tail vein catheter.

**T<sub>2</sub>-Weighted Proton MRI:** Conventional anatomic MRI images were acquired using a multi-slice T<sub>2</sub>-weighted RARE (Rapid Acquisition with Relaxation Enhancement) sequence. Images of different view/planes including axial, coronal, and sagittal were acquired to identify the location of the tumors or region of interest on the flank of the mouse. The imaging parameters of the T<sub>2</sub>-weighted scans were: echo time T<sub>E</sub> = 17 ms, repetition time T<sub>R</sub> = 2.5 s, 4 cm field of view, 256  $\mu$ m x 256  $\mu$ m in-plane resolution, ten to fifteen 1 - 1.5 mm slices, and 4 image averages. Three-dimensional (3D) tumor volumes were estimated from these MR images.

***In Vivo* <sup>13</sup>C MR Spectroscopy:** A series of slab-selective <sup>13</sup>C spectra from the slab thickness 6 - 8 mm were acquired right after injection of HP pyruvate using spFLASH sequence. The subcutaneous tumor location and similar size of the tumors employed in this study aided more precise slab placement through most of the cancer tissue and limited the contribution of non-malignant signal and/or keep the minimal contribution consistent throughout the experiments. A total of 90 transients were acquired using a delay time between each transient being 2 s (total time 3 min). Each transient utilized a 15° flip angle excitation Gauss pulse and 2048 data points. A small 8M <sup>13</sup>C-urea phantom doped with Gadolinium-DPTA was placed in each mouse experiment for chemical shift referencing. Experimental data were processed in MATLAB (MathWorks Inc., Natick, MA) and TopSpin (Bruker BioSpin GmbH, Ettingen, Germany) platforms. The phase correction and 10 to 15 Hz line-broadening were introduced to the individual spectrum. The area under the spectral peaks for pyruvate and lactate were integrated over the entire array. The lactate-to-pyruvate metabolic flux ratios (Lac/Pyr) were estimated by taking the ratio over the integration of lactate and pyruvate signals.

**[<sup>18</sup>F]FPIA-PET Imaging:** Mice are anesthetized and then injected intravenously with  $\sim$ 100  $\mu$ Ci [<sup>18</sup>F]FPIA in 150  $\mu$ L maximum total volume of phosphate buffer. Each mouse was allowed to recover for 60 min prior to imaging. The imaging protocol consisted of a 10 min PET followed by a 3 min CT scan CT (400 uA, 45 kV, 120 projections) on a Bruker Albira PET/SPECT/CT scanner (Bruker Biospin Corp., Billerica, MA, USA). The images were reconstructed using the Maximum Likelihood Expectation Maximization (MLEM) method with 12 iterations. Scatter, random decay, and attenuation corrections were applied. Analysis was done with Pmod (PMOD Technologies Ltd., Zürich, Switzerland). The mean % injected dose per millimeter cube (%ID/cc) of the tumor and normal tissue were measured. Data are reported as %ID/cc, or as ratio of the tumor to muscle (T/M).

#### **Aim I, Task 4:** Harvesting and storage of tissue samples

Prostate cancer PDX tumors were collected 24 to 48 hours after PET and HP-MR imaging. Tissue harvested for mass spectrometry analysis was flash frozen in liquid nitrogen and stored at -80°C for later workup. A 15 mg piece of PDX tissue were analyzed using mass spectroscopy to quantify

pyruvate and lactate levels for tumor lactate-to-pyruvate ratio measurements as described by Zacharias N, et al. (Zacharias N, Lee J, et al. *Mol Imaging Biol.* **2019** *21*, 86-94. doi: 10.1007/s11307-018-1199-6). To determine aberrant lipid PDX metabolism as described by Whitney TH, et al. (Whitney TH, Pisaneschi F, et al *J Nucl Med.* **2014**, *55*, 1506. doi: 10.2967/jnumed.114.140343) we plan to use ESI-LC-MS to quantify acetyl-carnitine, butyryl-carnitine, propionyl-carnitine, and palmitoyl-carnitine molecules using selected ion monitoring (SIM) and multiple reaction monitoring (MRM) mass spectrometry (A and B).

**Aim II, Task 5:** NMR spectroscopy-based metabolomics of flash frozen tissues

**Ex Vivo <sup>1</sup>H NMR Spectroscopy:** *Ex Vivo* <sup>1</sup>H NMR spectroscopic measurements were conducted using a protocol that has been established recently in our laboratory (Zacharias N, Lee J, et al. *Mol Imaging Biol.* **2019** *21*, 86-94. doi: 10.1007/s11307-018-1199-6). Briefly, flash-freezing tumor sample was weighed, crushed, and immersed in 3 mL of methanol-to-water mixture of 2:1 ratio in presence of polymer vortex beads. A process of mechanical homogenization was followed to separate the water-soluble metabolites. For NMR experiments the metabolites were dissolved in a solution containing 600  $\mu$ L of D<sub>2</sub>O, 36  $\mu$ L of phosphate buffer, and 4  $\mu$ L of 80 mM DSS (4,4-dimethyl-4-silapentane-1-sulfonic acid). Phosphate buffer was used to stabilize the pH value. The DSS was used as reference standard for normalizing the spectral signal of each metabolite.

NMR Spectra were obtained using a Bruker AVANCE III HD® NMR scanner (Bruker BioSpin Corporation) at a temperature of 298 K that has coupled with cryogenic temperature probe along a Z-axis shielded gradient. The operating frequency was 500 MHz for proton (<sup>1</sup>H) resonance. Water suppression technique was implemented. Spectra were acquired with a 90° pulse width, a 6.0 s scan delay time, 1024 Hz spectral width, and an acquisition time of 1.09 s (16,000 complex points). Spectroscopic data were processed by applying a number of techniques for cross verification and to test the reproducibility. Chenomx NMR Suite 8.1 software (Chenomx Inc., Edmonton, Canada) was applied for detail metabolic profiling. MestReNova software (Mestrelab Research, A Coruña, Spain) were used for quantitative analysis of metabolic peaks and associate area under the curve. The integral value of each peak was normalized by the value of the integral of the DSS reference peak.

**Aim II, Task 6:** Mass Spectrometry-based metabolomics

A 15mg piece of PDX tissue are analyzed using mass spectroscopy to quantify pyruvate and lactate levels for tumor lactate-to-pyruvate ratio measurements as described by Zacharias N, et al. (Zacharias N, Lee J, et al. *Mol Imaging Biol.* **2019** *21*, 86-94. doi: 10.1007/s11307-018-1199-6). To determine aberrant lipid PDX metabolism as described by Whitney TH, et al. (Whitney TH, Pisaneschi F, et al *J Nucl Med.* **2014**, *55*, 1506. doi: 10.2967/jnumed.114.140343) we have used ESI-LC-MS to quantify acetyl-carnitine, butyryl-carnitine, propionyl-carnitine, and palmitoyl-carnitine molecules using selected ion monitoring (SIM) and multiple reaction monitoring (MRM) mass spectrometry (A and B).

**Aim II, Task 7:** Transcriptome analysis of Tumors (four PDX lines; from task 4).

Total RNA was prepared using TRIZOL reagent (Invitrogen, USA) and processed at final step using RNEasy-kit (Qiagen, USA). RNA with high RIN values (>1.8) was processed for library and sequencing at the Genomics Core Facility at MD Anderson Cancer Center and paired-end sequencing was performed on an Illumina HiSeq platform (PE 2x100bp, 2 samples per lane) to provide a good coverage of >100X (after clonal read collapse). All post-sequencing analysis was also done at the core facility.

**Aim II, Task 8:** Immunohistochemistry analysis of Tumors (four PDX lines; from task 4). The tissue cross sections were stained for examining LDH-A, MCT1, MCT4 and Fatty Acid Synthase and Carnitine Acetyltransferase expression levels.

**Aim III, Task 9:** We have taken the Enzalutamide resistant PDX model, 274-4 for this specific aim. When the tumors are about 0.5 cm, the tumor bearing mice are randomized into two treatment groups. Mice in Group 1 received receive 10mg/kg body wt Enzalutamide by oral gavage while Group 2 mice will receive Enzalutamide and intraperitoneal injection of Syrosingopine, MCT4 inhibitor (7.5mg/kg body weight).

**Aim III, Task 10:** Synthesize  $^{18}\text{F}$ -Fluoropivalate ( $[^{18}\text{F}]\text{FPIA}$ ). Described before.

**Aim III, Task 11:** Imaging experiments.

Simultaneous  $^{13}\text{C}$  Pyruvate HP-MRI and  $[^{18}\text{F}]\text{FPIA}$  PET to interrogate both glycolysis and fatty acid oxidation pathways at the preclinical PET-MRI scanner at two time points. Methods described before.

**Aim III, Task 12:** Harvesting of tumors and storing for NMR spectroscopy, Mass spectrometry, immunohistochemistry and RNA sequencing studies at different time points.

**Aim IV, Task 13:** NMR spectroscopy based metabolomics of flash frozen tissues.

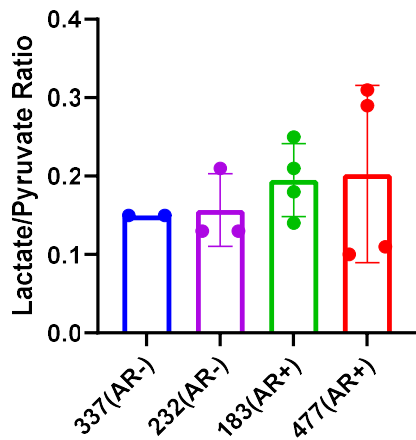
## **B. Specific Objectives:**

For the first reporting period, we are close to achieve the Milestone 1 as outlined in the Statement of Work. HP-MR imaging will inform us about the pyruvate-to-lactate transition in these PDX models as they respond to Enzalutamide and transition from castration sensitive prostate cancer (mCSPC) to mCRPC.  $[^{18}\text{F}]\text{FPIA}$  PET imaging is expected to capture progressive modulation in activity as the transition from mCSPC to mCRPC unfolds.

## **C. Key Outcomes:**

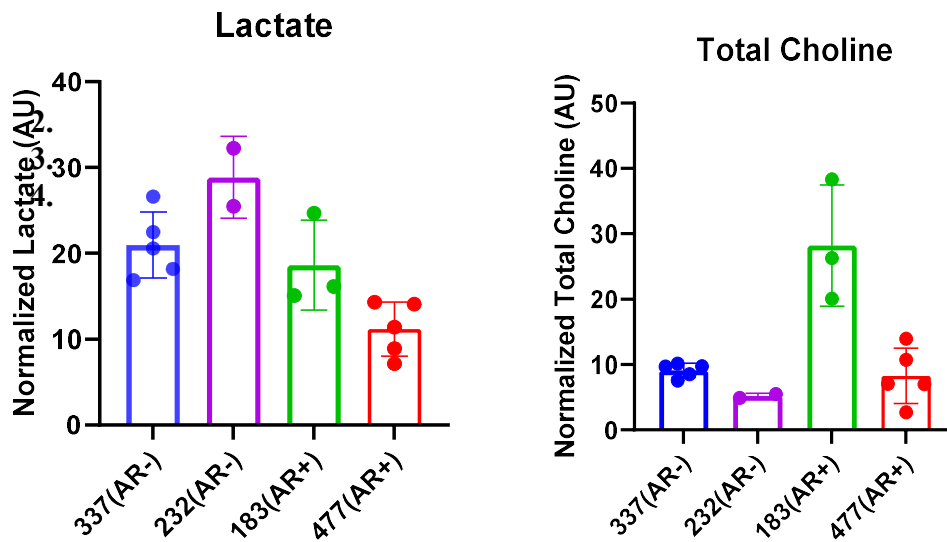
### **1. Potential of metabolic sub-typing of AR positive and negative prostate cancer PDX models by hyperpolarized metabolic imaging and metabolomics.**

This is now developed into a much larger project involving clinical data and Artificial Intelligence (AI) and we are now seeking extramural support from Department of Defense data Science Program, CPRIT and NCI on the data science application of metabolic classification of prostate cancer.



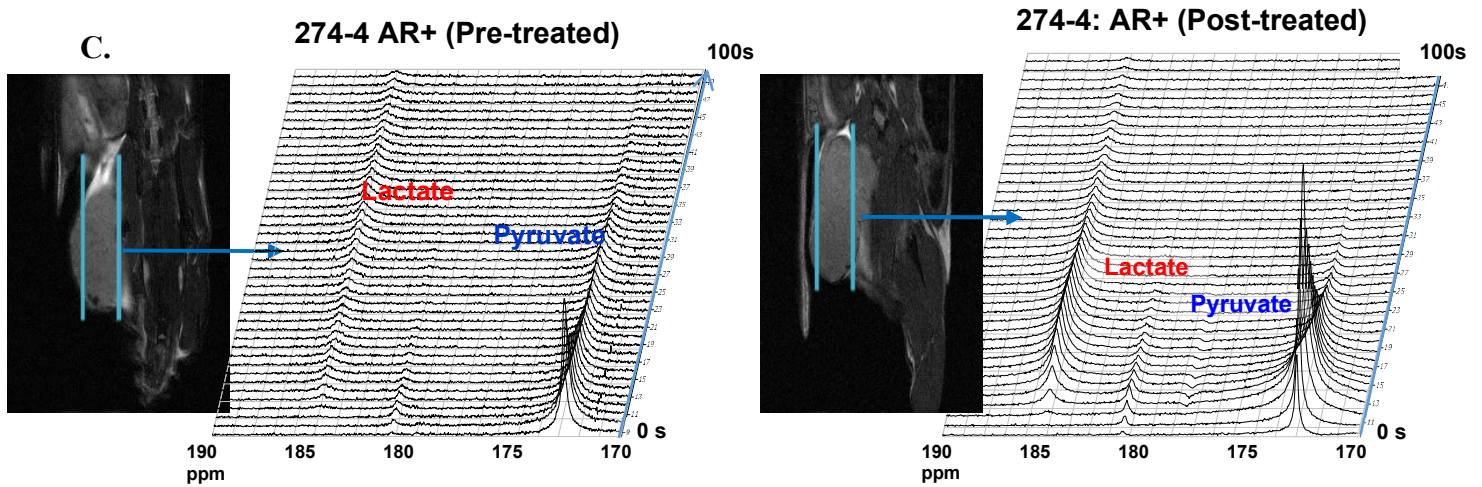
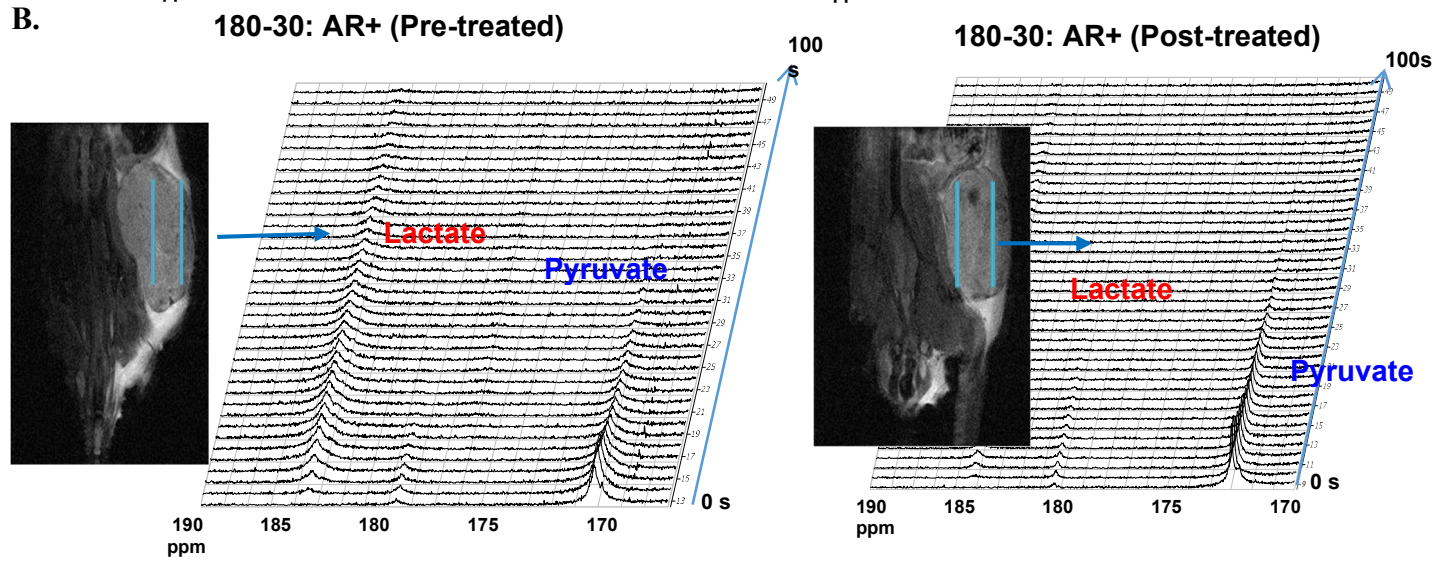
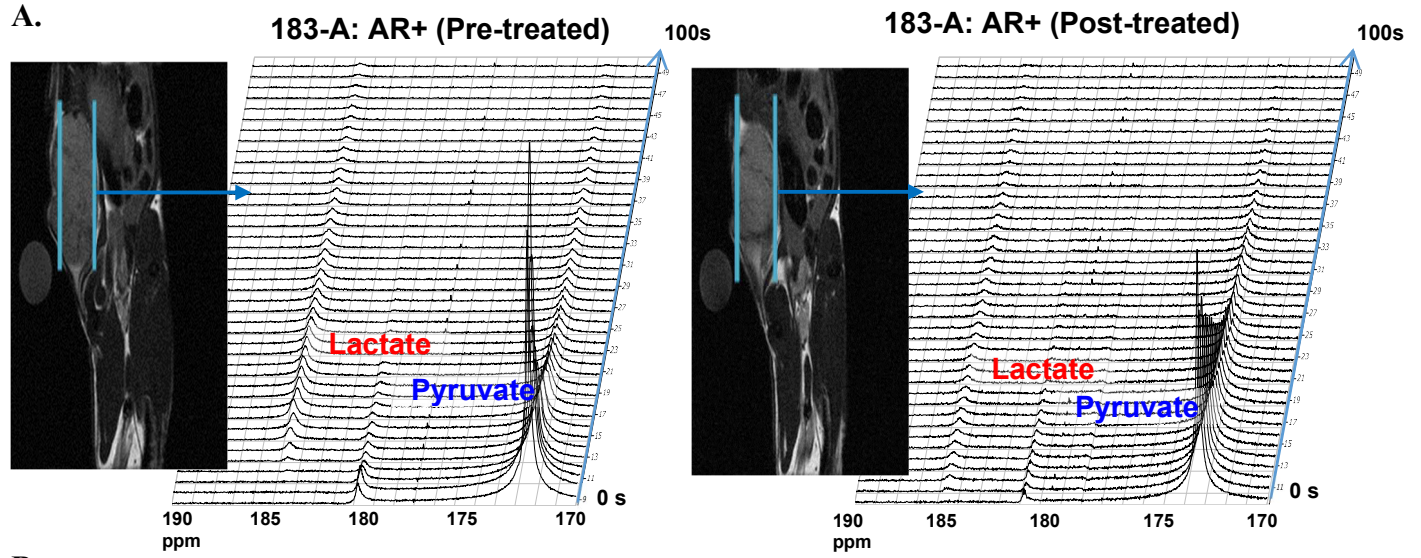
**Figure 2.** Hyperpolarized lactate-to-pyruvate ratio in patient derived xenografts (PDX) of prostate cancer with AR positive and negative status *in vivo*.

**Figure 3.** The normalized lactate and choline metabolites were quantified using nuclear magnetic resonance (NMR) metabolomics of *ex vivo* tumor samples for different prostate cancer PDXs with AR+ and AR- status.

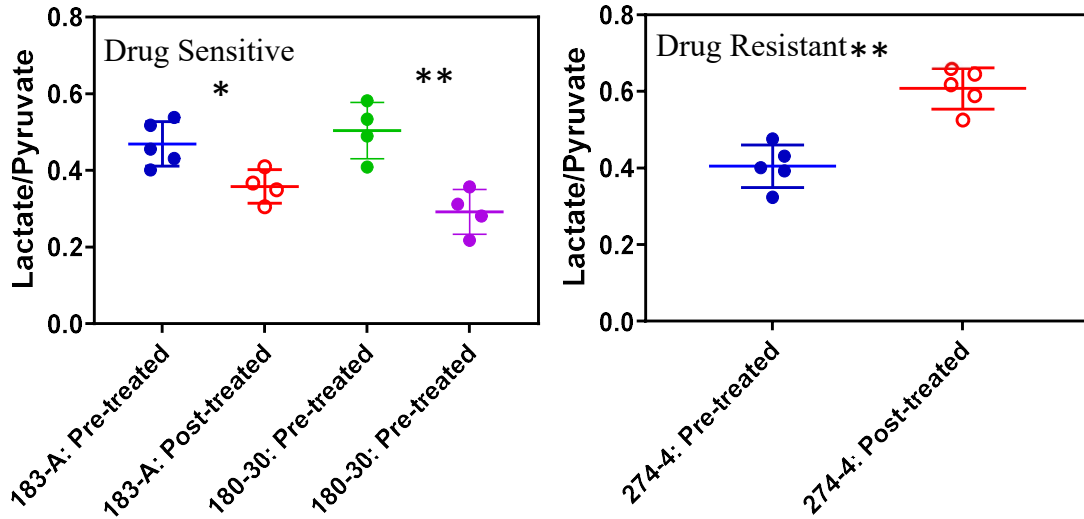


2. Hyperpolarized metabolic imaging of Lactate-to-Pyruvate conversion can be employed to determine Enzalutamide sensitivity or resistance in PDX models of prostate cancer. This was discussed in last year's progress report.

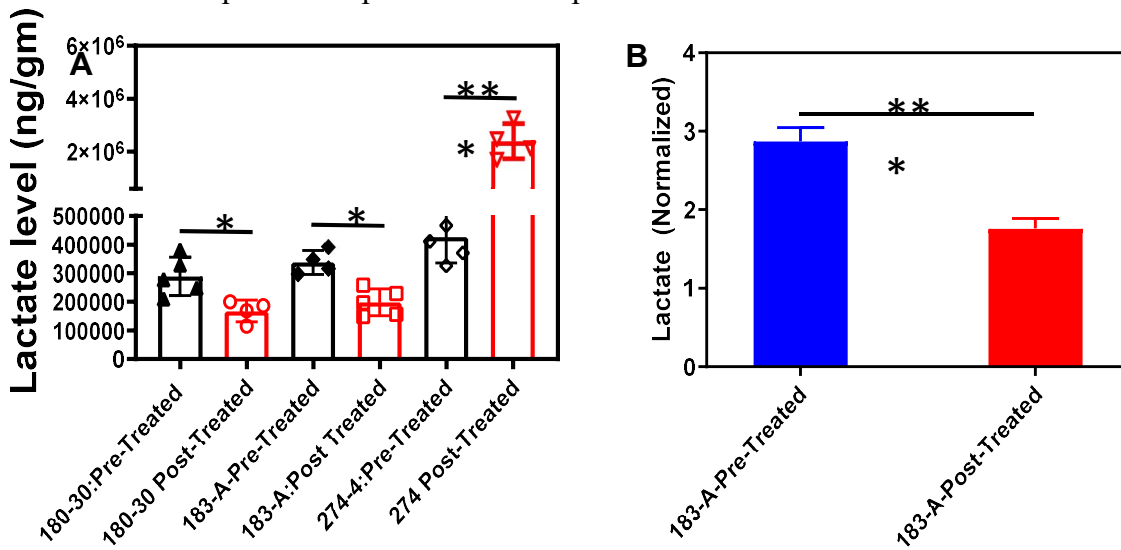
**Figure 4: Representative hyperpolarized metabolic imaging data.** In three Androgen Receptor positive (AR+) PDX models of prostate cancer, of which PDXs 183-A and 180-30 are Enzalutamide sensitive and PDX 274-4 is Enzalutamide resistant, hyperpolarized  $[1-^{13}\text{C}]$  pyruvate metabolic imaging is employed to measure the real-time lactate flux generation before treatment and after 7 days of Enzalutamide treatment (Panels A-C). Panel D demonstrates that lactate-to-pyruvate is significantly reduced in Enzalutamide sensitive tumor while, lactate-to-pyruvate is significantly increased in Enzalutamide resistant tumor. This work has resulted in a manuscript under review at *Cancer Research*.



D.



**Figure 5. Representative metabolomics data.** Both Mass Spectrometry (Panel A) and NMR spectroscopy-based (Panel B) metabolomics of *ex vivo* flash frozen tumor samples support the *in vivo* data from hyperpolarized metabolic imaging (Figure 4). In panel A, the *ex vivo* lactate pool size is higher in post treated samples in Enzalutamide resistant, PDX 274-4, whereas the *ex vivo* lactate pool is lower in post-treated samples in Enzalutamide sensitive PDX 180-30. Similar lower *ex vivo* lactate pool is observed in panel B in post-treated samples in Enzalutamide sensitive PDX 183-A.

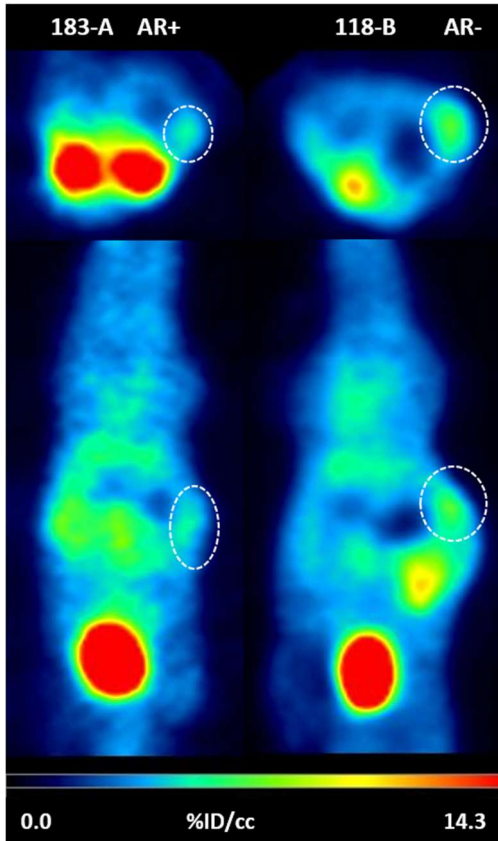


### 3. [<sup>18</sup>F]FPIA-PET imaging shows <sup>18</sup>F-Fluoropivalate (FPIA) uptake in PDX tumors.

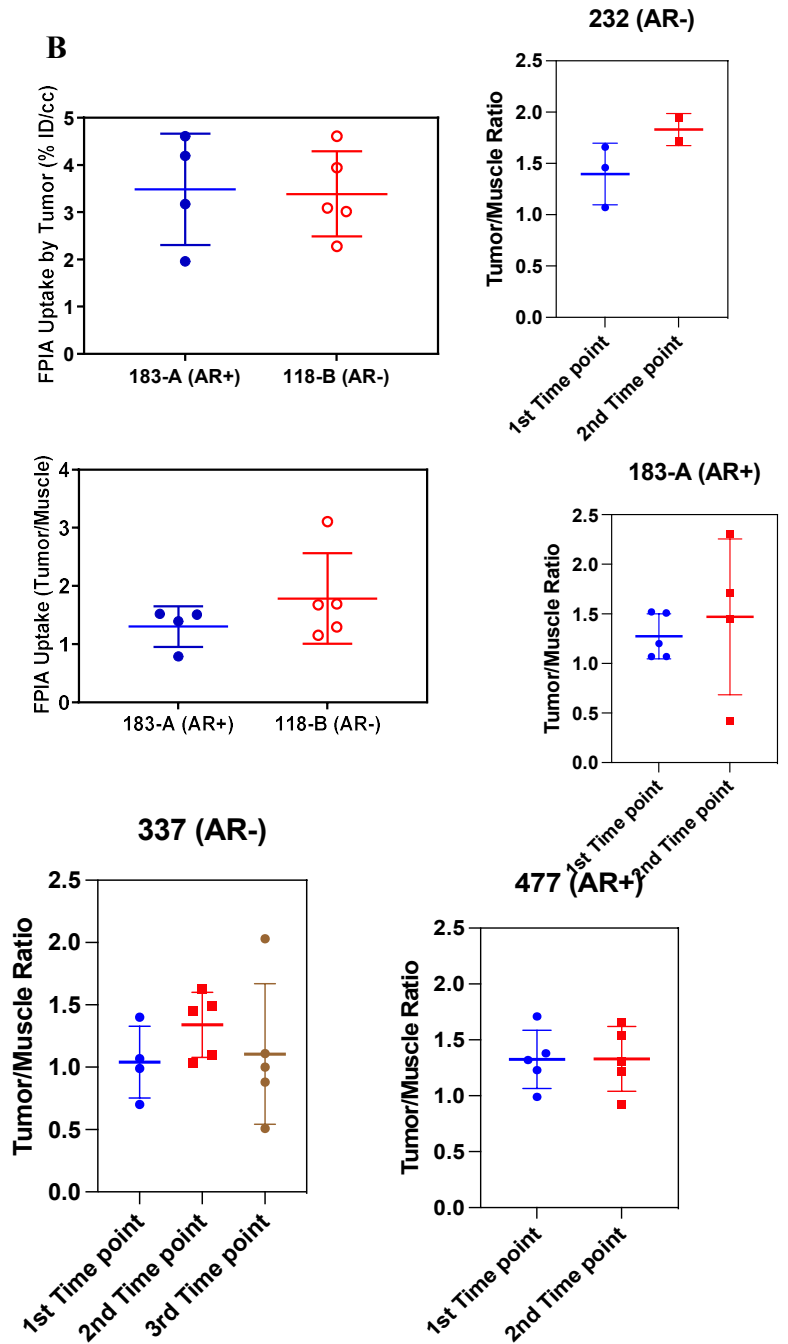
**Figure 6. Representative [<sup>18</sup>F]FPIA-PET imaging data.** We imaged several AR positive and negative PDX models. In all models, we observe FPIA uptake, with an average of about 3.5 % ID/cc. A representative image is shown in Panel A. Representative FPIA uptake plots are shown in Panel B. When data are analyzed as tumor to muscle (T/M), we observe a slightly higher uptake in the AR-PDX, although the difference is not statistically significant (Panel B). Our data shows the potential of

FPIA-PET to visualize prostate cancer, and we hope to employ FPIA-PET to pinpoint differences in metabolic reprogramming following enzalutamide therapy.

**A**

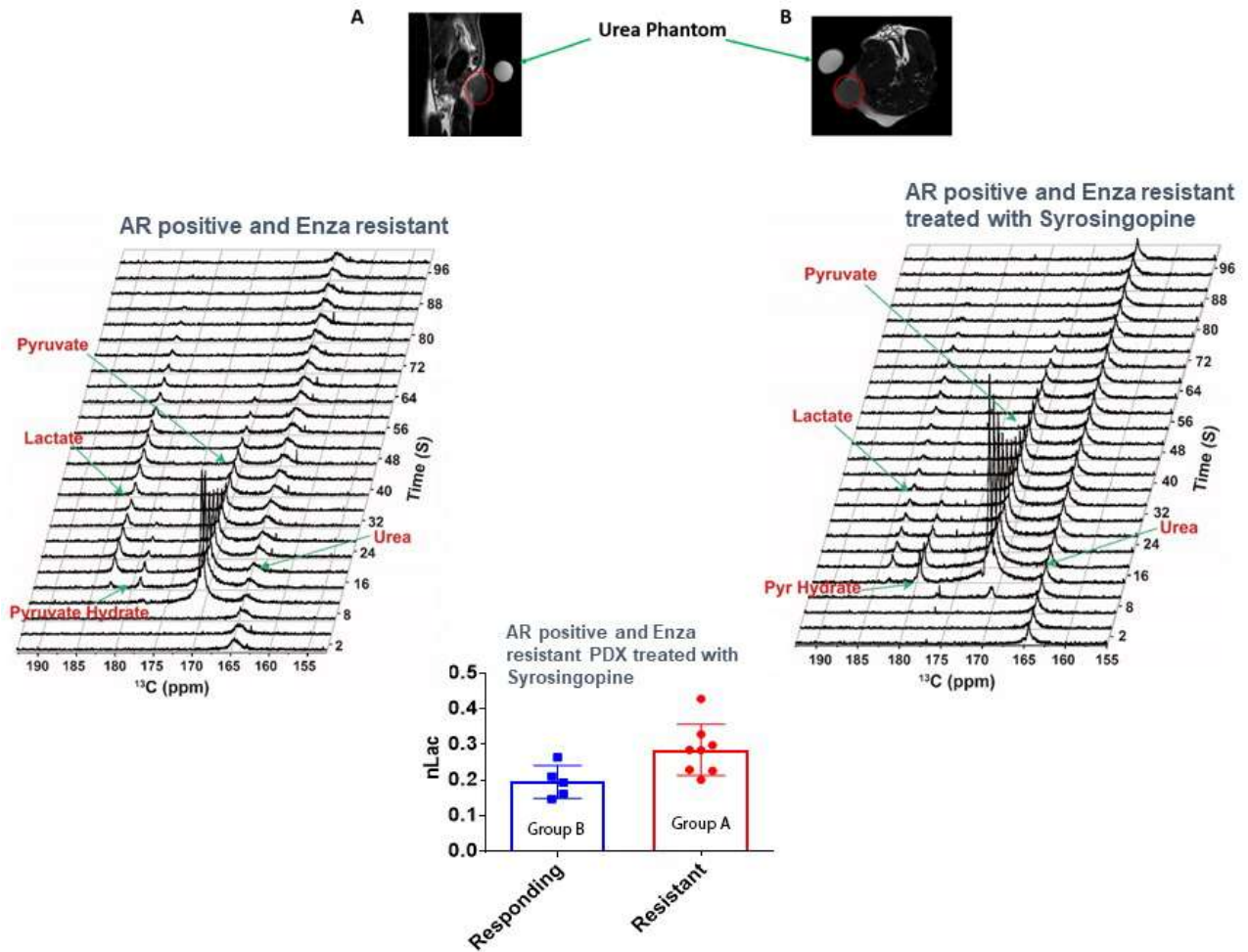


**B**



4. Preliminary HP-MR data in one AR resistant PDX model (274-4) shows that MCT inhibition with the clinical drug, Syrosingopine can overcome resistance to Enzalutamide (Enza). More work is on going to validate this result and imaging work is currently in progress with  $^{13}\text{C}$  Pyruvate HP-MRI and  $^{18}\text{F}$  FPIA PET.

**Figure 7:** Preliminary HP-MR data showing dynamic metabolic flux differences the 274-4 AR+, mice treated with Enzalutamide by oral gavage (Group A) and 274-4 AR+, mice treated with Enzalutamide and intraperitoneal injection of MCT4 inhibitor, Syrosingopine (Group B). nLac (lactate-to-pyruvate ratio) shows the reduced metabolic flux of lactate in the Group B treated with MCT4 inhibitor, Syrosingopine.



What opportunities for training and professional development has the project provided?

Dr. Prasanta Dutta who is supported by this project has given an oral presentation on the work he has accomplished in past two years at the 25th International Symposium on Radiopharmaceutical Sciences (iSRS) in May 22-26, 2023. His abstract was selected through a rigorous competitive process for an oral presentation.

#### **How were the results disseminated to communities of interest?**

There are two manuscripts currently under review based on the work accomplished in this project. One is under peer review at *Cancer Research* and the other one is at the *Journal of Labelled Compounds and Radiopharmaceuticals*. An oral presentation on “*Combining PET and HP-MR to Interrogate Prostate Cancer Metabolism*” at the 25th International Symposium on Radiopharmaceutical Sciences (iSRS) on May 25, 2023. This is biggest professional gathering of molecular imaging scientists dedicated to the understanding of biology and medicine through multimodal *in vivo* imaging of cellular and molecular events involved in normal and pathologic processes and utilization of quantitative molecular imaging in patient care.

*Describe briefly what you plan to do during the next reporting period to accomplish the goals and objectives.*

In the second reporting period of the project, we have made substantial progress on Specific Aim I and II and we have started working on the most impactful aim of the project, Specific Aim III on imaging to overcoming the ADT resistance by targeting Monocarboxylate Transporter (MCT) pathway. Our preliminary HP-MR data on only one of the AR positive Enzalutamide resistant PDX model (274-4) shows that MCT inhibition with the clinical drug, Syrosingopine can overcome resistance to Enzalutamide. The main task of the next reporting period will be to validate this result with multiomics correlative work and imaging is currently ongoing with  $^{13}\text{C}$  Pyruvate HP-MRI and  $^{18}\text{F}$  FPIA PET.

#### **4. IMPACT:**

**What was the impact on the development of the principal discipline(s) of the project?**

Patients with advanced prostate cancer receive anti-androgen drug, Enzalutamide as the first line of treatment. Unfortunately, many patients develop resistance to treatment, and the disease relapses with aggressive histology and metastatic phenotype. we have employed a non-invasive magnetic resonance imaging (MRI) based technique (hyperpolarized metabolic imaging) to image Enzalutamide sensitivity or resistance in prostate cancer. This has been validated by *ex vivo* metabolomics data. Furthermore, we have employed a novel, Positron Emission Tomography (PET) agent,  $^{18}\text{F}$ -FPIA, to image the linked fatty acid oxidation pathway in prostate cancer. We have attempted to restore Enzalutamide sensitivity in a resistant model by targeting the monocarboxylate transporter (MCT) pathway using an MCT inhibitor, Syrosingopine and we have successfully interrogate this transition from Enzalutamide resistance to sensitivity by hyperpolarized metabolic imaging. We believe, we can employ this multi-modal approach to identify the drug resistant patients at an earlier time point. We intend to develop personalized metabolic imaging modality to target treatment strategies of different sub-types of lethal prostate cancer.

**What was the impact on other disciplines?**

Nothing to report as of now

**What was the impact on technology transfer?**

Nothing to report as of now

Nothing to report as of now

## 5. CHANGES/PROBLEMS:

Nothing to report as of now

### Actual or anticipated problems or delays and actions or plans to resolve them

Nothing major to report, In the proposal, we suggested these following four PDX models for study  
274-4 (AR+, Enzalutamide resistant)  
133-3 (AR+, Enzalutamide resistant)  
183-A (AR+, Enzalutamide sensitive)  
170-1 (AR+, Enzalutamide sensitive)

However, we ran into some logistical challenges to generate all PDX models we originally proposed from our PDX core facility as the core ran into several issues in one past one year. Therefore, we have kept performing experiments on the models that are available and fit the goals of the project.

### PDX models employed so far in the study

PDX	HP-MR	FPIA- PET	NMR met	MS met	AR	Enzalutamide
144	x	x	x	x	negative	
118	x	x	x		negative	
232	x	x	x	x	negative	
337	x	x	x		negative	
183	x	x	x	x	positive	sensitive
274	x	x	x	x	positive	resistant
177	x	x	x		negative	
180	x		x	x	positive	sensitive
477	x	x	x		positive	resistant

**Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents**

**Significant changes in use or care of human subjects**

Nothing to report

**Significant changes in use or care of vertebrate animals**

Nothing to report

**Significant changes in use of biohazards and/or select agents**

Nothing to report

**6. PRODUCTS:**

- **Publications, conference papers, and presentations**  
**Journal publications.**

No published manuscript to report.

**Books or other non-periodical, one-time publications.**

Nothing to report

**Other publications, conference papers and presentations.**

Accepted conference abstract for oral presentation at the World Molecular Imaging Congress (WMIC), 2022, Miami.

Dutta P, Enriquez JS, Armijo R, Wang M, Han J, Shepherd P, Frigo DE, Titus MA, Pisaneschi P, Bhattacharya PK. *Combining PET Imaging and HP-MR to Interrogate Prostate Cancer Metabolism.*

- **Website(s) or other Internet site(s)**

Nothing to report

- **Technologies or techniques**

Nothing to report

- **Inventions, patent applications, and/or licenses**

Nothing to report

- **Other Products**

Nothing to report

## **7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS**

**What individuals have worked on the project?**

Name: Pratip Bhattacharya, Ph.D.

Project Role: PD/PI

Researcher Identifier (e.g. ORCID ID): 0000-0002-0625-252X

Nearest person month worked: 1.92

Contribution to Project: Oversight and coordination of all research activities.

Funding Support: Please see attached Other Support Document.

Name: Federica Pisaneschi, Ph.D.

Project Role: Co-I

Researcher Identifier (e.g. ORCID ID): 0000-0002-1989-4417

Nearest person month worked: 1.44

Contribution to Project: Radiochemistry

Funding Support: Please see attached Other Support Document

Name: Prasanta Dutta, Ph.D.

Project Role: Research Scientist

Researcher Identifier (e.g. ORCID ID): 0000-0002-9887-6507

Nearest person month worked: 6.00

Contribution to Project: Assistance for Drs. Bhattacharya & Pisaneschi on HP-MR, PET and metabolomics

Name: Peter Shepherd

Project Role: Coordinator, Research Laboratory

Nearest person month worked: 1.32

Contribution to Project: Generating and maintaining the patient derived (PDX) prostate cancer animal Models and transcriptomics

Name: Abishai Dominic, Ph.D.

Project Role: PostDoc Fellow

Nearest person month worked: 1.00

Contribution to Project: Assistance for Drs. Bhattacharya & Pisaneschi on IHC and cyTOF

**Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?**

As mentioned in the last year's progress report, Dr. Mark Titus, Associate Professor, Department of Genitourinary Medical Oncology, MD Anderson Cancer Center, who was a co-investigator in this proposal has retired from MD Anderson Cancer Center on last Feb 28, 2023. We have replaced his effort with Peter Shepherd. Peter is a research laboratory coordinator at the Department of Genitourinary Medical Oncology, MD Anderson. Peter is responsible for generating and maintaining the patient derived (PDX) prostate cancer animal models in the research plan as well as administering therapy to these animal models. He is also assisting with transcriptomics data acquisition.

**What other organizations were involved as partners?**

Nothing to report

## **8. SPECIAL REPORTING REQUIREMENTS**

**COLLABORATIVE AWARDS:** *F*

**QUAD CHARTS:** *I*

## **9. APPENDICES:**

**Previous/Current/Pending Support for DOD  
Bhattacharya, Pratip**

**PREVIOUS:**

<b>Title:</b>	Circulating Biomarkers and Imaging for Early Detection of Pancreatic Cancer (U01CA214263 – Sen, Killary)
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	National Institutes of Health / National Cancer Institute
<b>Grants Officer:</b>	Ghosh-Janjigian Sharmistha National Cancer Institute 9609 Medical Center Drive Bethesda, MD 20892 ghoshjanjigias@mail.nih.gov
<b>Performance Period:</b>	08/07/2018 – 07/31/2023
<b>Level of Funding:</b>	
<b>Project Goals</b>	To validate integrated blood-based biomarker signature for detection of early stage PDAC and secondly for biomarker panels in concert with novel imaging for the early detection of PDAC at a precursor stage when the disease remains curative.
<b>Specific Aims</b>	<p>Aim 1: Validation of pathway-associated biomarker signatures of early stage resectable tumors and precision imaging for early detection of asymptomatic PDAC utilizing early stage cohorts.</p> <p>Aim 2: Validation of PDAC early precursor lesion associated biomarker signatures and precision imaging developed with precursor lesions harboring genetically engineered mouse models and well characterized large longitudinal cohorts.</p> <p>Aim 3: Blinded validation of early detection biomarker signatures in retrospective pre-diagnostic samples.</p> <p>Aim 4: Prospective screening of integrated biomarkers and imaging using risk scores in pre-diagnostic high risk PDAC cohorts and Pancreatic Cancer High Risk Clinic (PCHRC) patients.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Imaging Biomarkers for Immunotherapy Resistance in Melanoma In Vivo (85152 – Bhattacharya)
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	Melanoma Research Alliance – Pilot Award
<b>Grants Officer:</b>	Rachel Fischer 730 15th Street, NW Washington, DC 20005

	rfischer@curemelanoma.org
<b>Performance Period:</b>	06/01/2021 – 05/31/2023
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop biomarkers to image immunotherapy resistance in vivo and to improve its success rate.
<b>Specific Aims</b>	Aim 1: To monitor changes in tumor pHe and HP lactate-to-pyruvate metabolic flux in B16/B16 melanoma clones that acquired resistance to checkpoint blockade immunotherapy through serial in vivo passage using acidoCEST and HP-MRI.  Aim 2: To pH-sensitize immunotherapy with MCT1 inhibition treatment, as evaluated with acidoCEST and HP-MRI.
<b>Overlap:</b>	None

<b>Title:</b>	Hyperpolarized Silicon Nanoparticle “Spin Batteries” For Imaging Biochemical Reactions in Vivo
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center – Institutional Research Grant
<b>Grants Officer:</b>	Nyma Shah, Director, Research Funding Programs The University of Texas MD Anderson Cancer Center – Research Administration Office 1515 Holcombe Blvd., Unit 1436 Houston, TX 77030
<b>Performance Period:</b>	09/01/2018 – 08/31/2022 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To address the sensitivity and specificity of hyperpolarized metabolic MR technique in multiple animal models of pancreatic premalignancy.
<b>Specific Aims</b>	Aim 1: Validate through-bond transfer of polarization from hyperpolarized silicon-29 nuclei.  Aim 2: Validate through-space transfer of polarization from silicon nucleus to 13C carbonyl using the Nuclear Overhauser Effect (NOE).  Aim 3: Test and validate transfer of 29Si hyperpolarization to 13C as a method of sensing proteolytic enzymes, Granzyme B, and oxidative enzymes, e.g., myeloperoxidase.
<b>Overlap:</b>	None

<b>Title:</b>	Translational Applications in an Animal Model of Pancreatic Cystic Neoplasm and Cancer (1 R01 CA218004-01A1 – Maitra)
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<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	National Institutes of Health
<b>Grants Officer:</b>	Richard V. Mazurchuk Program Director National Cancer Institute BG 9609 MSC 9760 9609 Medical Center Drive Bethesda, MD 20892-9760
<b>Performance Period:</b>	04/04/2018 – 03/31/2022
<b>Level of Funding:</b>	
<b>Project Goals</b>	To enhance the translational applicability of this model by using it as a controlled platform to address key unmet needs in the management of IPMNs in two areas: imaging correlates and circulating biomarkers.
<b>Specific Aims</b>	Aim 1: Utilizing the Kras;Gnas model of cystic neoplasia for identification of imaging correlates of progression to invasive PDAC, with cross-validation of imaging studies in IPMN patients.  Aim 2: Utilizing the Kras;Gnas model of cystic neoplasia for identification of circulating biomarkers of progression to invasive PDAC, with cross-validation in human IPMN-derived biospecimens.
<b>Overlap:</b>	None

<b>Title:</b>	Overcoming ADT Resistance in Metastatic Castration Resistance Prostate Cancer (mCRPC) By Targeting Monocarboxylate Transporter (MCT) Pathway
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	Mike Slive Foundation
<b>Grants Officer:</b>	Emily Capilouto P.O. Box 530748 Birmingham, AL 35253 emily@mikeslivefoundation.org
<b>Performance Period:</b>	01/01/2021 – 12/31/2021
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop personalized metabolic imaging modality to target treatment strategies of different sub-types of PCa and to interrogate different metabolic factors by imaging that are associated with a particular sub-type.
<b>Specific Aims</b>	Aim 1: To perform HP-13C Pyruvate MRI in four well characterized PDX murine models of prostate cancer in vivo before and after the antiandrogen therapy with Enzalutamide to image alteration in glycolytic pathway after treatment response. We will further analyze ADT sensitive and ADT resistant PDX utilizing NMR-spectroscopy combined with mass spectrometry-based metabolomics, immunohistochemistry and

	<p>transcriptome profiling. Using this approach, we expect to delineate pathways of ADT resistance.</p> <p>Aim 2: To image overcoming the ADT resistance by targeting Monocarboxylate Transporter (MCT) pathway (via MCT inhibition with the clinical drug, Syrosingopine) by performing HP-13C Pyruvate MRI in two well characterized PDX murine models of prostate cancer in vivo. We will further analyze the overcoming of ADT resistance utilizing NMR-spectroscopy combined with mass spectrometry-based metabolomics, immunohistochemistry and transcriptome profiling</p>
<b>Overlap:</b>	None

<b>Title:</b>	MD Anderson Cancer Center SPORE in Gastrointestinal Cancer (1 P50 CA221707-01 – Kopetz, Maitra)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	National Institutes of Health / National Cancer Institute
<b>Grants Officer:</b>	Shane Woodward 9609 Medical Center Drive Bethesda, MD 20892 woodwars@mail.nih.gov
<b>Performance Period:</b>	08/20/2019 – 05/31/2024 (Dr. Bhattacharya's effort ended 11/30/2021)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To reduce mortality and morbidity rates from colorectal cancer (CRC) and pancreatic ductal adenocarcinoma (PDAC), and to improve the quality of life of patients afflicted by these diseases.
<b>Specific Aims</b>	<p>Aim 1: To evaluate clinical activity of an optimized combination of immune checkpoint inhibitors with a personalized, neo-antigen peptide vaccine.</p> <p>Aim 2: To determine the contribution of STAT3 signaling to CRC development in high-risk patients with familial syndromes or inflammatory bowel disease.</p> <p>Aim 3: To evaluate oxidative phosphorylation as a therapeutic vulnerability in pancreatic cancer.</p> <p>Aim 4: Empower GI SPORE research and collaborations through research Cores and Programs.</p> <p>Aim 5: Strategic integration with the translational GI research community.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Early Detection of PanIN by Hyperpolarized Metabolic MR Imaging (16-65-BHAT)
<b>Effort:</b>	0.90 CM, 8% effort
<b>Supporting Agency:</b>	Pancreatic Cancer Action Network
<b>Grants Officer:</b>	Maya Bader 1500 Rosecrans Avenue, Suite 200 Manhattan Beach, CA 90266 mbader@pancan.org
<b>Performance Period:</b>	07/01/2016 – 12/31/2018 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop the capability of non-invasively detecting advanced pancreatic intraepithelial neoplasia (PanIN) precursor lesions in pancreas prior to invasive disease using metabolic imaging modality.
<b>Specific Aims</b>	Aim 1: Employ metabolic imaging with hyperpolarized <sup>13</sup> C pyruvate to detect and monitor the progression of PanIN precursor lesions to pancreatic cancer.  Aim 2: Establish the in vivo metabolic flux ratios of alanine-to-lactate (Ala/Lac) and lactate-to-pyruvate (Lac/Pyr) as predictive biomarkers of neoplastic progression of advanced PanIN to aggressive pancreatic cancer.  Aim 3: Employ high resolution Nuclear Magnetic Resonance (NMR) Spectroscopy in ex vivo tumor samples and correlate NMR metabolomics data with real-time in vivo dynamic metabolic flux data in Aim 1 and 2 and immunohistochemistry data for independent validation of the metabolic biomarkers for clinical translation.
<b>Overlap:</b>	None

<b>Title:</b>	TMC-GCC Collaborative Symposium: Metabolism in Cancer
<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	Gulf Coast Consortium - 2016 John S. Dunn Collaborative Research Award Program
<b>Grants Officer:</b>	Suzanne Tomlinson PO Box 1892, MS141 Houston, TX 77251-1892
<b>Performance Period:</b>	04/01/2017 – 03/31/2019
<b>Level of Funding:</b>	
<b>Project Goals</b>	To secure contribution of funds from participating GCC institutions to fund the annual Metabolism in Cancer Symposium for at least two years.
<b>Specific Aims</b>	Aim 1: Learn how metabolism research could benefit cancer patients.

	<p>Aim 2: Bring attention to local investigators working on research projects in metabolism in cancer.</p> <p>Aim 3: Identify possible areas within this broad domain where investigators in the TMC and greater Houston area can collaborate.</p> <p>Aim 4: Assemble investigators into cross-institutional teams and promote their interactions with newly launched TMC initiatives.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Targeted Molecular Imaging of Cell Death in Ovarian Cancer (1R21CA181994-01A1 - Millward, Lu)
<b>Effort:</b>	0.24 CM, 2% effort
<b>Supporting Agency:</b>	NCI Exploratory/Developmental Research Grant Award
<b>Grants Officer:</b>	Anne Menkens 9609 Medical Center Drive Rockville, MD 20850 menkensa@mail.nih.gov
<b>Performance Period:</b>	02/03/2016 – 01/31/2019 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	Develop targeted molecular imaging agents to visualize apoptosis and autophagy in vivo by PET and MRI.
<b>Specific Aims</b>	<p>Aim 1: Molecular imaging of apoptosis and autophagy can provide an immediate pharmacodynamic readout of chemotherapy and radiation therapy efficacy, allowing clinicians to identify the optimal therapeutic regime in days rather than weeks or months.</p> <p>Aim 2: Non-tumor cell death can be assessed in parallel with tumor destruction, providing insight into potential side effects before clinical presentation.</p> <p>Aim 3: Cell death imaging can be integrated into early phase clinical trials (Phase 0/I) of targeted apoptosis- and autophagy-modulating therapies to predict response and resistance, accelerating the stratification of patients most likely to respond to therapy.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Early Detection of Pancreatic Cancer by Targeted Molecular Imaging of Hyperpolarized Silicon Nanoparticles (RP170593 - Pudukalakatti)
<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	Gulf Coast Consortia – Computational Cancer Biology Training Grant

<b>Grants Officer:</b>	Melissa Glueck, Keck Center Director Gulf Coast Consortia – BioScience Research Collaborative (BRC) Suite 160, 6500 Main St. Houston, TX 77030
<b>Performance Period:</b>	04/01/2017 – 03/31/2019
<b>Level of Funding:</b>	
<b>Project Goals</b>	To improve early detection of pancreatic cancer by Magnetic Resonance Imaging (MRI) using a highly sensitive and specific biomarker of this virtually incurable disease.
<b>Specific Aims</b>	Aim 1: Optimize the levels of hyperpolarization achievable by radical-free <sup>29</sup> Si DNP on lactose-functionalized and MUC1-modified hydrogels encapsulated SiNPs as a function of nanoparticle size and <i>in vitro</i> targeting efficacy.  Aim 2: Demonstrate targeted real-time hyperpolarized MRI with lactose functionalized and MUC1-hydrogel encapsulated SiNPs in two distinct animal models of pancreatic cancer.
<b>Overlap:</b>	None

<b>Title:</b>	Virtual Colonoscopy for Early Detection of Colorectal Cancer
<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	Colon Cancer Coalition
<b>Grants Officer:</b>	Anne Carlson, Executive Director Colon Cancer Coalition 5666 Lincoln Drive, Suite 270 Edina, MN 55436  anne@coloncancercoalition.org
<b>Performance Period:</b>	06/28/2018 – 08/31/2019 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To employ hyperpolarized silicon nanoparticles that target highly accessible transmembrane mucin glycoproteins (MUC1) that are overexpressed in colon cancer, as viable MRI based molecular imaging agents for novel, early diagnosis of colorectal abnormalities and cancer.
<b>Specific Aims</b>	Aim 1: Establish and validate real-time targeted molecular imaging of CRC using MUC1 functionalized SiNPs.  Aim 2: Apply real-time targeted molecular imaging technique to well annotated CRC mouse models to validate early detection by MUC1 functionalized HP SiNPs.
<b>Overlap:</b>	None

<b>Title:</b>	Elimination of hypoxia sensitizes resistant solid tumors to Immunotherapy (RP170399 - Curran)
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<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	Cancer Prevention and Research Institute of Texas – Individual Investigator Research Award
<b>Grants Officer:</b>	Wayne Roberts Cancer Prevention and Research Institute of Texas PO Box 12097 Austin, TX 78711
<b>Performance Period:</b>	12/01/2016 – 11/30/2019
<b>Level of Funding:</b>	
<b>Project Goals</b>	Determine whether disruption of hypoxic zones will compromise the integrity of the suppressive myeloid and myofibroblast network thereby sensitizing solid tumors to T cell co-stimulatory immunotherapy.
<b>Specific Aims</b>	Aim 1: Determine how targeted ablation of tumor hypoxia affects T cell infiltration and function, as well as sensitivity to T cell immunotherapy, in murine models of prostate cancer.  Aim 2: Assay the effects of hypoxia ablation and immunotherapy on the immune composition and functional capacity of suppressive myeloid cells in the prostate cancer tumor microenvironment.  Aim 3: Assess the impact of hypoxia ablation on the viability, distribution, and immunosuppressive function of myofibroblasts in prostate cancer.
<b>Overlap:</b>	None

<b>Title:</b>	Development of Platelet-based Metabolic Biomarkers for Ovarian Cancer
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	Rivkin Center for Ovarian Cancer
<b>Grants Officer:</b>	Kiran Dhillon, PhD Director of Scientific Programs Rivkin Center for Ovarian Cancer 801 Broadway, Suite 701 Seattle, WA 98122
<b>Performance Period:</b>	04/01/2019 – 03/31/2020
<b>Level of Funding:</b>	
<b>Project Goals</b>	We hypothesize that tumor educated platelets from blood of ovarian cancer patients would exhibit altered metabolism compared to platelets from healthy volunteers.
<b>Specific Aims</b>	Aim 1: Determine metabolic profiles of human platelets (resting) and human platelets (activated with thrombin or/and collagen) and compare that with the metabolic profiles of human ovarian cancer cell lines with platelets and human ovarian cancer cell lines without platelets.

	Aim 2: Determine metabolic profiles from platelets isolated from healthy donors and ovarian cancer patient donors.
<b>Overlap:</b>	None

<b>Title:</b>	Interdisciplinary Translational Pre/Postdoctoral Program in Cancer Nanotechnology (1T32CA196561-01 - Sokolov)
<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	National Institutes of Health
<b>Grants Officer:</b>	Susan E Lim National Institutes of Health 31 Center Drive MSC 2510 Bethesda, MD 20892-2510
<b>Performance Period:</b>	09/01/2015 – 08/31/2020
<b>Level of Funding:</b>	
<b>Project Goals</b>	The goal of this program is to train scientists who will transfer the great promise of nanotechnology into clinical reality. We have designed a novel training program to educate future leaders in the broad field of nanotechnology with specific interests in cancer-related applications, who are keenly aware of the needs and demands of clinical environment as well as of major challenges of translational research.
<b>Specific Aims</b>	<p>Aim 1: All trainees will work with at least two program faculty mentors (one from Rice and one from MD Anderson) to define and carry out an independent research problem.</p> <p>Aim 2: Incoming trainees will participate in a unique 2-week-long boot camp in “Cancer Management and Nanotechnology” that provides an overview of current opportunities and barriers in the field.</p> <p>Aim 3: Trainees will develop foundational background in the field by taking four courses related to translational cancer or nanotechnology topics.</p> <p>Aim 4: Trainees will gain important lab management skills by participating in a short hands-on course providing an introduction to laboratory and project management.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Metabolic Imaging of Therapeutic Efficacy in Glioblastoma by Hyperpolarized Magnetic Resonance (P50CA127001 – Lang)
<b>Effort:</b>	0.12 SM, 1% effort
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center – Brain SPORE DRP

<b>Grants Officer:</b>	Biny Joseph, Program Director The University of Texas MD Anderson Cancer Center 1515 Holcombe Blvd., Unit 0431 Houston, TX 77030
<b>Performance Period:</b>	09/01/2019 – 08/31/2020
<b>Level of Funding:</b>	
<b>Project Goals</b>	Continue to develop a real-time metabolic imaging platform for early efficacy of therapy in mouse models of brain cancer with the goal of translating this technique in the GBM patients.
<b>Specific Aims</b>	<p>Aim 1: To evaluate the response of individual tumors in three cohorts of patient-derived glioma stem cell intracranial xenografts (GSC) mouse model to 1) radiation therapy, 2) one anti-angiogenic drug (Avastin) and 3) OXPHOS inhibitor IACS-10759 applied <i>in vivo</i>, using HP <sup>13</sup>C pyruvate MR imaging and spectroscopy. Establish the real-time metabolic flux ratio (nLac) as therapeutic response marker.</p> <p>Aim 2: To perform steady-state high-resolution NMR (Nuclear Magnetic Resonance spectroscopy) metabolomics of <i>ex vivo</i> tissues collected from these tumors before and after therapy for metabolic fingerprinting and correlate that <i>in vivo</i> HP MR data from Aim 1.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Molecular Targeted Magnetic Resonance Reporter for Cancer Detection (RP180164 – Bhattacharya, Carson)
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	Cancer Prevention & Research Institute of Texas – High-Impact/High-Risk Research Award
<b>Grants Officer:</b>	Wayne Roberts Cancer Prevention and Research Institute of Texas PO Box 12097 Austin, TX 78711
<b>Performance Period:</b>	08/31/2018 – 02/28/2021 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To create a new construct, combining the molecular affinity of antibodies with the sensitivity of hyperpolarized magnetic resonance, in order to create an early detection technology for colorectal cancer.
<b>Specific Aims</b>	<p>Aim 1: Optimize the affinity to MUC1 and the urease activity our MR Reporters.</p> <p>Aim 2: Analyze the ability of the optimized MUC1 targeted MR Reporter to detect CRC using <i>in vivo</i> imaging in a human MUC1 expressing CRC animal model.</p>

	<p>Aim 3: Combine our MR Reporter with gold nanoparticles and optimize our constructs to determine if a theranostic particle can be generated.</p> <p>Aim 4: Test the in vivo diagnostic ability of our gold nanoparticle MR Reporter in our CRC animal models.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Metabolic Imaging of Patients with Prostate Cancer Using Hyperpolarized Pyruvate (Logothetis)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	Koch Functional Imaging and Prostate Cancer Project
<b>Grants Officer:</b>	Claudia Delgado, Executive Director The University of Texas MD Anderson Cancer Center Grants and Contracts P.O. Box 301407, Unit 1644 Houston, TX 77030
<b>Performance Period:</b>	12/21/2015 – 07/21/2021 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To evaluate sensitivity and specificity of an HP compound for detecting, staging and surveillance of prostate cancer and determining the efficacy of a therapeutic approach.
<b>Specific Aims</b>	<p>Aim 1: Characterize metabolic adaptive response of PDX models in vivo to finasteride using HP pyruvate, fumarate, N-acetyl-aspartylglutamate (NAAG) and Chemical Reaction-Induced Multimolecular Polarization (CRIMP) with histological and in vitro metabolomics correlations. These compounds will interrogate glycolysis, the TCA cycle, PSMA pathway metabolic components respectively that are linked to AR signaling. CRIMP will be employed for real-time pH mapping in vivo.</p> <p>Aim 2: Optimize dynamic MR spectroscopic imaging sequences tailored to the unique spectral and spatiotemporal characteristics of HP substrates on preclinical and clinical imaging systems.</p> <p>Aim 3: Develop and validate GMP production protocol of HP imaging agents.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Mark-III 129Xe Gas Polarizer System (1S10OD027038-01A1 – Bankson)
<b>Effort:</b>	0.00 CM, 0% effort
<b>Supporting Agency:</b>	National Institutes of Health
<b>Grants Officer:</b>	Malgorzata Klosek 9609 Medical Center Drive

	Bethesda, MD 20892 klosekm@csr.nih.gov
<b>Performance Period:</b>	02/01/2020 – 01/31/2021
<b>Level of Funding:</b>	
<b>Project Goals</b>	This Shared Instrumentation Grant for a <sup>129</sup> Xe gas polarizer would bring new capabilities for imaging lung function to MD Anderson and the Texas Medical Center.
<b>Specific Aims</b>	N/A
<b>Overlap:</b>	None

**CURRENT:**

<b>Title:</b>	Hyperpolarized MRI and Artificial Intelligence (AI) to Inform Metabolic Evolution of Glioblastoma Throughout Tumor Progression (Bhattacharya)
<b>Effort:</b>	0.00 CM, 0% effort
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center SPORE in Brain
<b>Grants Officer:</b>	Biny Joseph, Program Director The University of Texas MD Anderson Cancer Center 1515 Holcombe Blvd., Unit 0431 Houston, TX 77030
<b>Performance Period:</b>	09/01/2022 – 04/30/2024 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	The overarching goal of this proposal is to develop AI driven HP metabolic MRI-based technique to reduce the time necessary to detect brain tumor evolution in three key areas of tumor progression: initial development of the tumor, its regression following therapy, and the eventual recurrence of the tumor.
<b>Specific Aims</b>	Aim 1: To correlate glycolytic metabolic competency in two cohorts of GSC xenografts with graded aggressiveness profiles from medium, to low by employing MRI, HP metabolic imaging and DL for early detection.  Aim 2: To evaluate the response of individual tumors in the same two cohorts of GSC xenografts to radiation therapy using MRI, HP metabolic imaging and DL for interrogating early efficacy of therapy and early relapse.
<b>Overlap:</b>	None

<b>Title:</b>	Real Time Metabolic Imaging to Interrogate Early Detection and Prevention of Pancreatic Cancer (PA220132 – Bhattacharya)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	Department of Defense PCARP TRPA
<b>Grants Officer:</b>	TBA
<b>Performance Period:</b>	10/01/2023 – 09/30/2026
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop the capability of non-invasively detecting and imaging the regression of premalignant pancreatic lesions and early stage disease following immunoprevention in preclinical and clinical cohorts using hyperpolarized metabolic imaging (HP-MRI).
<b>Specific Aims</b>	Aim 1: To determine the utility of preclinical imaging with [1-13C] pyruvate hyperpolarized MRI to track the efficacy of an immunopreventive intervention.  Aim 2: To demonstrate the feasibility and utility of pancreatic premalignant metabolic imaging with HP [1-13C] pyruvate MRI in a pilot imaging study.
<b>Overlap:</b>	None

<b>Title:</b>	Real Time Metabolic Imaging to Interrogate Early Detection and Prevention of Pancreatic Cancer (Enriquez)
<b>Effort:</b>	0.0 CM, 0% effort (Mentor)
<b>Supporting Agency:</b>	National Institutes of Health
<b>Grants Officer:</b>	Mariam Eljanne Program Representative NATIONAL CANCER INSTITUTE eljannem@mail.nih.gov
<b>Performance Period:</b>	09/01/2023 – 08/31/2025
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop, demonstrate and validate novel imaging approaches (HP MR and acidoCEST MRI) to detect the early stages of pancreatic cancer prior to invasive disease.
<b>Specific Aims</b>	Specific Aim 1: To demonstrate and validate the sensitivity and specificity of dynamic hyperpolarized metabolic imaging (Dissertation Research Project; F99 phase).  Specific Aim 2: To demonstrate detection of premalignant pancreatic lesions with a combination of HP and acidoCEST MR and to determine the efficacy of immunoprevention on premalignant models (Postdoctoral Research Direction; K00 phase).
<b>Overlap:</b>	There is some conceptual overlap with the trainee grant in applying hyperpolarized metabolic imaging for early detection

	of pancreatic cancer with Department of Defense PCARP TRPA and PREVENT. There is no budgetary overlap.
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<b>Title:</b>	Employing AI to Predict Clinical Outcomes in Ovarian Cancer (Shams)
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	San Jose State University/OCRA
<b>Grants Officer:</b>	N/A
<b>Performance Period:</b>	01/01/2023 – 12/31/2025
<b>Level of Funding:</b>	
<b>Project Goals</b>	In this proposal, we will apply artificial intelligence (AI) in this context of morphologic and multi-platform omics differences observed in HGSOc.
<b>Specific Aims</b>	<p>Aim 1: To detect distinct morphologic patterns in HGSOc by AI at the time of laparoscopy and to correlate these patterns with clinical outcomes.</p> <p>Aim 2: To integrate clinical and laboratory variables with multi-platform omics data obtained from tissue samples at the time of laparoscopic assessment of disease burden to detect distinct biologic patterns in HGSOc by AI.</p> <p>Aim 3: To employ the observed morphologic, clinical, and omics patterns identified above in the development of a hybrid biomarker optimized to predict clinical outcomes for patients with advanced HGSOc.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Interdisciplinary Translational Pre/Postdoctoral Program in Cancer Nanotechnology (T32CA196561 – Sokolov)
<b>Effort:</b>	0.00 CM, 0% effort
<b>Supporting Agency:</b>	National Institutes of Health / National Cancer Institute
<b>Grants Officer:</b>	N/A
<b>Performance Period:</b>	04/01/2022 – 03/31/2027
<b>Level of Funding:</b>	2,659,935
<b>Project Goals</b>	The overall goal of this project is to develop a novel training program to prepare Ph.D. scientists who will transfer the many promises of cancer nanotechnology into clinical reality of the future
<b>Specific Aims</b>	N/A
<b>Overlap:</b>	None

<b>Title:</b>	Evaluation of Real-time Metabolic Imaging Biomarkers for Detection of Pancreatic Premalignant Lesions (75N91019D00021 – Bhattacharya / McAllister / Brown)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	National Institutes of Health / National Cancer Institute

	PREVENT
<b>Grants Officer:</b>	Christy L. Hunter 2115 E. Jefferson St. MSC 8500 Suite 4B 432 Bethesda, MD 20892 Christy.hunter@nih.gov 301-624-8788
<b>Performance Period:</b>	09/16/2021 – 12/15/2023
<b>Level of Funding:</b>	
<b>Project Goals</b>	To determine the sensitivity and specificity of a novel metabolism-based imaging technique for early detection of PDAC in the gold standard murine models as well as define correlative biomarkers which could serve to validate findings in humans within pancreatic cancer high risk individuals in the future who undergo yearly screening.
<b>Specific Aims</b>	NA
<b>Overlap:</b>	None

<b>Title:</b>	Developing Imaging Biomarkers for Immunotherapy Resistance In Vivo (RP220270 – Bhattacharya)
<b>Effort:</b>	2.40 CM, 20% effort
<b>Supporting Agency:</b>	Cancer Prevention & Research Institute of Texas (CPRIT) – Individual Investigator Research Awards
<b>Grants Officer:</b>	Cancer Prevention & Research Institute of Texas (CPRIT) 1701 North Congress Avenue, Suite 6-127 Austin, TX 78701 cprit@cpriti.state.tx.us
<b>Performance Period:</b>	03/01/2022 – 02/28/2025
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop biomarkers to image immunotherapy resistance in vivo and to improve the success rate of immunotherapy for patients with cancer.
<b>Specific Aims</b>	<p>Aim 1 To monitor changes in tumor pHe and HP lactate-to-pyruvate metabolic flux in B16/BL6 melanoma clones that acquired resistance to checkpoint blockade immunotherapy through serial in vivo passage using acidoCEST MRI and HP MRS.</p> <p>Aim 2 To identify the best pH-sensitizer with acidoCEST MRI and HP MRS in melanoma model of progressive immunotherapy resistance.</p> <p>Aim 3 To improve the response to immunotherapy in a melanoma model of progressive immunotherapy resistance by</p>

	first applying a pH-sensitizer while monitoring with acidoCEST MRI and HP MRS.
<b>Overlap:</b>	None

<b>Title:</b>	Institutional Startup Research Funding
<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center
<b>Grants Officer:</b>	Nyma Shah, Director, Research Funding Programs The University of Texas MD Anderson Cancer Center – Research Administration Office 1515 Holcombe Blvd., Unit 1436 Houston, TX 77030
<b>Performance Period:</b>	04/01/2012 – 08/31/2024 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop a real-time molecular and metabolic imaging program at the University of Texas, MD Anderson Cancer Center employing different modalities of hyperpolarization.
<b>Specific Aims</b>	No specific aims, as funding is geared toward investigator's overall research program.
<b>Overlap:</b>	None

<b>Title:</b>	WS #28 – Metabolic Imaging of Prostate Cancer Patients for Monitoring Therapeutic Efficacy (Bhattacharya / Bankson / Kundra)
<b>Effort:</b>	0.60 CM, 5% effort
<b>Supporting Agency:</b>	General Electric Healthcare/Center for Biomedical Imaging
<b>Grants Officer:</b>	Leslie Billings, Administrator The Center for Advanced Biomedical Imaging 1881 East Rd., Unit 1905 Houston, TX 77054
<b>Performance Period:</b>	08/26/2016 – 02/29/24 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	To employ hyperpolarized pyruvate for prostate cancer risk stratification in preclinical mouse models of prostate cancer and prepare for clinical trial in prostate cancer patients at MD Anderson Cancer Center.
<b>Specific Aims</b>	Aim 1: Infrastructure development for applying MR hyperpolarization imaging on clinical prostate cancer patients at MDACC.

	<p>Aim 2: Optimize dynamic MR spectroscopic imaging sequences tailored to the unique spectral and spatiotemporal characteristics of HP substrates on a 3T clinical scanner.</p> <p>Aim 3: Determine the sensitivity and specificity of MR hyperpolarization with <sup>13</sup>C(1)-pyruvate to predict clinical response in high grade prostate cancer patients enrolled in an on-going abiraterone therapeutic clinical trial.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Real-time Metabolic Imaging Biomarkers for Detection of Pancreatic Premalignant Lesions (Bhattacharya)
<b>Effort:</b>	0.12 CM, 1% effort
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center /Duncan Family Institute for Cancer Prevention and Risk Assessment Seed-funding Research Program
<b>Grants Officer:</b>	Lori Armstrong 1515 Holcombe Blvd. Houston, TX 77030-4009 laarmstrong@mdanderson.org
<b>Performance Period:</b>	06/01/2021 – 05/31/2024 (NCE)
<b>Level of Funding:</b>	
<b>Project Goals</b>	The overarching goal of this seed proposal is to develop the capability of non-invasively detecting advanced pancreatic intraepithelial neoplasia (PanIN) precursor lesions in pancreas prior to invasive disease in by employing hyperpolarized metabolic imaging.
<b>Specific Aims</b>	<p>Aim 1: To demonstrate the sensitivity of dynamic hyperpolarized metabolic biomarkers to detect and monitor the progression of pancreatic premalignant lesions to pancreatic cancer in inducible GEM models at multiple time points in vivo.</p> <p>Aim 2: The overarching goal of this seed proposal is to develop the capability of non-invasively detecting advanced pancreatic intraepithelial neoplasia (PanIN) precursor lesions in pancreas prior to invasive disease in by employing hyperpolarized metabolic imaging.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Identifying Different Metabolic Sub-types of Prostate Cancer for Early Therapeutic Assessment (W81XWH-21-1-0763 – Bhattacharya)
<b>Effort:</b>	2.40 CM, 20% effort
<b>Supporting Agency:</b>	Department of Defense – Prostate Cancer Research Program – Idea Development Award
<b>Grants Officer:</b>	Joshua McKean 820 Chandler St.

	Fort Detrick, MD 21702 joshua.d.mckean3.civ@mail.mil
<b>Performance Period:</b>	09/01/2021 – 08/31/2024
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop personalized metabolic imaging modality to target treatment strategies of different sub-types of PCa and to interrogate different metabolic factors by imaging that are associated with a particular sub-type.
<b>Specific Aims</b>	<p>Aim 1: To perform HP-13C Pyruvate MRI, [18F]FPIA -PET imaging in four well characterized patient derived xenograft (PDX) murine models of prostate cancer in vivo before and after the antiandrogen therapy with Enzalutamide to image both alteration in glycolytic and fatty acid oxidation pathways after treatment response.</p> <p>Aim 2: To elucidate the metabolic drivers of ADT resistance. We will analyze ADT sensitive and ADT resistant PDX utilizing NMR-spectroscopy combined with mass spectrometry-based metabolomics, immunohistochemistry and transcriptome profiling. Using this approach, we expect to delineate pathways of ADT resistance.</p> <p>Aim 3: To image overcoming the ADT resistance by targeting Monocarboxylate Transporter (MCT) pathway (via MCT inhibition with the clinical drug, Syrosingopine) by performing HP-13C Pyruvate MRI in two well characterized patient derived xenograft (PDX) murine models of prostate cancer in vivo. [18F]FPIA -PET</p>
<b>Overlap:</b>	This grant has some overlap with the CPRIT Individual Investigator Research Award (IIRA) -RP220313 where co-investigator, Dr. Federica Pisaneschi serves as PI. Conceptually both grants are similar. The scope of the CPRIT grant is however much broader than the DoD Idea Grant in terms of 1) larger numbers and types of PDX mouse models employed, 2) a different hyperpolarized metabolic reagent, 15N-L-Carnitine-d6 employed to track the carnitine pathway, and 3) finally, Enolase 2 (ENO2) (via ENO2 inhibition with natural phosphonate antibiotic, SF2312) inhibition has been employed as a strategy to overcome ADT resistance. CPRIT grant essentially builds up on the DoD grant and is essentially on parallel track with little direct overlap.

<b>Title:</b>	Imaging Metabolic Reprogramming in Prostate Cancer (RP220313 – Pisaneschi)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	Cancer Prevention and Research Institute of Texas

<b>Grants Officer:</b>	Patty Moore Cancer Prevention and Research Institute of Texas 1701 North Congress Avenue, Suite 6-127 Austin, TX 78701 pmoore@cprit.texas.gov
<b>Performance Period:</b>	03/01/2022 – 02/28/2025
<b>Level of Funding:</b>	
<b>Project Goals</b>	This proposal will build up on these findings, investigating by PET and HP-MRI how carnitine metabolism is modulated in these PDXs before and after Enzalutamide-treatment. Furthermore, we will attempt to overcome Enzalutamide resistance by targeting MCT and ENO2 proteins with inhibitors, Syrosingopine and SF2312 respectively, and image this transformation from resistance to sensitization by HP-MRI and PET.
<b>Specific Aims</b>	<p>Aim 1: HP MRI and PET imaging. To perform <sup>13</sup>C-Pyruvate, <sup>15</sup>N-L-Carnitine-d6 HP-MRI and [<sup>18</sup>F]FPIA-PET imaging in six well characterized patient derived xenograft (PDX) murine models of prostate cancer in vivo before and after the antiandrogen therapy with Enzalutamide to image both alteration in glycolytic and fatty acid oxidation pathways after treatment response.</p> <p>Aim 2: Ex vivo analyses. To understand the mechanistic basis of the success and/failure of HP-MRI and FPIA-PET in imaging specific PDX sub-types of PCa xenografts through metabolic and transcriptome profiling and analyses of key protein expressions in the metabolic pathways.</p> <p>Aim 3: Overcoming resistance. To image overcoming the Enzalutamide resistance by targeting Monocarboxylate Transporter (MCT) (via MCT inhibition with the clinical drug, Syrosingopine) and Enolase 2 (ENO2) (via ENO2 inhibition with natural phosphonate antibiotic, SF2312) and define whether MCT and/or ENOs are viable targets for potential combination therapy with anti-androgens.</p>
<b>Overlap:</b>	This grant has some overlap with DoD PCRP grant (W81XWH-21-1-0763 – Bhattacharya) where co-investigator, Dr. Federica Pisaneschi serves as PI. Conceptually both grants are similar. The scope of the CPRIT grant is however much broader than the DoD Idea Grant in terms of 1) larger numbers and types of PDX mouse models employed, 2) a different hyperpolarized metabolic reagent, <sup>15</sup> N-L-Carnitine-d6 employed to track the carnitine pathway, and 3) finally, Enolase 2 (ENO2) (via ENO2 inhibition with natural phosphonate antibiotic, SF2312) inhibition has been employed as a strategy to overcome ADT resistance. CPRIT grant essentially builds up on the DoD grant and is essentially on parallel track with little direct overlap. Dr.

	Pisaneschi has declared the overlap to CPRIT before accepting the award.
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<b>Title:</b>	Targeted Hyperpolarized Molecular Beacons for Colorectal Cancer Detection (R21EB031217 – Bhattacharya)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	National Institutes of Health / National Institute of Biomedical Imaging and Bioengineering
<b>Grants Officer:</b>	Joseph Peterson 9609 Medical Center Drive Bethesda, MD 20892 petersonjt@mail.nih.gov
<b>Performance Period:</b>	09/30/2021 – 07/31/2024
<b>Level of Funding:</b>	
<b>Project Goals</b>	To combine the versatile molecular affinity of antibodies with the sensitivity of hyperpolarized magnetic resonance (HP MR) to develop “hyperpolarized (HP) molecular beacons” for deep tissue imaging for CRC detection.
<b>Specific Aims</b>	Aim 1: To develop binding and activity assays for MUC1 targeted HP molecular beacons and to optimize construct (0-12 months).  Aim 2: To perform non-invasive in vivo MR imaging with optimized MUC1 targeted HP molecular beacon (8-24 months).  Aim 3: To generate and optimize binding of gold nanoparticle coupled MUC1 HP molecular beacon construct (18-36 months).
<b>Overlap:</b>	There is scientific overlap of this grant with the CPRTP postdoctoral fellowship given below but there is no budgetary overlap for these projects.

<b>Title:</b>	Hyperpolarized molecular beacons for early detection of colorectal cancer (Dominic)
<b>Effort:</b>	0.0 CM, 0% effort (Mentor)
<b>Supporting Agency:</b>	CPRTP Janice Davis Gordon Memorial in Colorectal Cancer Prevention
<b>Grants Officer:</b>	N/A
<b>Performance Period:</b>	09/01/2023 – 08/31/2025
<b>Level of Funding:</b>	
<b>Project Goals</b>	To significantly increase the ability of detecting cancer by utilizing several amplification steps and combining the versatile binding affinity and specificity of antibodies with the sensitivity of hyperpolarized Magnetic Resonance (HP MR).

<b>Specific Aims</b>	<p>Specific Aim 1: To develop binding and activity assays for MUC1-targeted HP molecular beacons and optimize constructs.</p> <p>Specific Aim 2: To perform non-invasive in vivo MR imaging with optimized MUC1-targeted HP molecular beacons.</p> <p>Specific Aim 3: To expand the repertoire of HP molecular beacons to other cell surface markers in CRC that can be combined to target key biomarkers of CRC.</p>
<b>Overlap:</b>	There is scientific overlap of this trainee grant with the R21EB031217 grant from NIBIB but there is no budgetary overlap for these projects. This fellowship only pays portion of the postdoctoral fellow's salary.

## PENDING

<b>Title:</b>	Targeting of metabolic crosstalk between leukemia and macrophages as a therapeutic strategy for disease eradication and restoration of innate immune system (Baran)
<b>Effort:</b>	0.36 CM, 3% effort
<b>Supporting Agency:</b>	Cancer Prevention Research Institute of Texas (CPRIT)
<b>Grants Officer:</b>	TBD
<b>Performance Period:</b>	03/01/2024 – 02/28/2027
<b>Level of Funding:</b>	
<b>Project Goals</b>	To incorporate the concept of OXPHOS/Lactate blockade into anti-leukemia therapy.
<b>Specific Aims</b>	<p>Specific Aim 1: Determine the anti-leukemia efficacy of MCT1/MCT4 blockade on OXPHOS activity and the impact of dual inhibition of MCT1/MCT4 and OXPHOS on AML cells, macrophages, and AML/macrophage co-culture, and characterize the molecular mechanisms of action.</p> <p>Specific Aim 2: To determine the anti-leukemia efficacy of OXPHOS/MCT1/MCT4 blockade in AML models with or without macrophages infusion as a consolidation therapy in vivo.</p> <p>Specific Aim 3: To characterize transcriptomic and metabolic signatures of AML cells and macrophages vulnerable to OXPHOS/MCT1/MCT4 inhibition, and spatial signature of AML-macrophage cross talk upon MCT1/MCT4/OXPHOS inhibition in vivo.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Cancer Prevention Education: Student Research Experiences (Chang)
<b>Effort:</b>	0.0 CM, 0% effort (Mentor)
<b>Supporting Agency:</b>	National Institutes of Health
<b>Grants Officer:</b>	TBA
<b>Performance Period:</b>	12/01/2023 – 11/30/2028
<b>Level of Funding:</b>	
<b>Project Goals</b>	To complement and/or enhance the training of a workforce to meet the nation's biomedical, behavioral and clinical research needs;
<b>Specific Aims</b>	<p>Specific Aim 1: Recruit, select, and train high-performing students for 25 10-week positions annually, using innovative and effective strategies to attract a demographically diverse student population.</p> <p>Specific Aim 2: Cultivate and support the pool of participating faculty mentors.</p> <p>Specific Aim 3: Provide a superlative mentored research training experience in a variety of cancer prevention disciplines</p> <p>Specific Aim 4: Provide highly relevant didactic, career and professional skills development, and research ethics curricula in cancer prevention research and cancer prevention careers.</p> <p>Specific Aim 5: Contribute to broadening participation of groups underrepresented in research careers through development of students' identity as scientists, particularly in cancer research.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Artificial Intelligence for early identification of androgen-indifferent, lethal prostate cancer and therapy selection (Merenyi, Bhattacharya, Pilie)
<b>Effort:</b>	1.20 CM, 10% effort
<b>Supporting Agency:</b>	Rice University/Cancer Prevention Research Institute of Texas (CPRIT)
<b>Grants Officer:</b>	TBA
<b>Performance Period:</b>	03/01/2023 – 02/28/2027
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop strategies to define the biology of lethal prostate cancer, to develop effective treatments, and address mechanisms of resistance for men with mCRPC.
<b>Specific Aims</b>	Specific Aim 1: Evaluate prediction accuracy of neural map-based AI models using ex vivo and laboratory measurements complemented by in vivo HP-MRI/CSI before and after treatment at multiple time points.

	Specific Aim 2: Evaluate prediction accuracy of AI models using laboratory measurements and in vivo HP-MRI/CSI before and after treatment at multiple time points.
<b>Overlap:</b>	None

<b>Title:</b>	Development of Silicon Nanoparticles as Molecular Beacons for Targeted Molecular Imaging (Bhattacharya)
<b>Effort:</b>	1.20 CM, 10 % effort
<b>Supporting Agency:</b>	National Institutes of Health
<b>Grants Officer:</b>	TBA
<b>Performance Period:</b>	12/01/2023 – 11/30/2025
<b>Level of Funding:</b>	
<b>Project Goals</b>	To develop a suite of molecular beacons of functionalized hyperpolarized SiNPs for real-time MRI detection of all/most polyp subtypes along with a new platform technology for real-time, non-invasive molecular imaging of early-stage CRC using MRI employing targeted imaging, formerly only in the realm of positron emission tomography (PET) and single-photon emission computed tomography (SPECT).
<b>Specific Aims</b>	<p>Specific Aim 1: Establish targeted molecular imaging by MUC1 functionalized HP SiNPs in a well annotated CRC humanized mouse model.</p> <p>Specific Aim 2: Create a library of functionalized SiNPs that recognize and identify most/all polyp subtypes, including flat lesions, without binding to normal tissue. A) Expand the repertoire of receptor targeting moieties to other cell surface markers in CRC that can be targeted with functionalized SiNPs using bioengineered polyp avatars.</p> <p>Specific Aim 3: Measure the ability of the multifunctional SiNP formulation library to recognize the spectrum of pathologically different subtypes of freshly collected human polyp tissue.</p>
<b>Overlap:</b>	None

### **CERTIFICATION**

I, PD/PI, Partnering PI, or other senior/key personnel, certify that the statements herein are current, accurate, and complete to the best of my knowledge; I agree to update such disclosure at the request of the agency prior to the award of support and at any subsequent time the agency determines appropriate during the term of the award; and I have been made aware of the requirements under Section 223(a)(1) of the William M. (Mac) Thornberry National Defense Authorization Act for Fiscal Year 2021. I am aware that any false, fictitious, or fraudulent statements or claims may subject me to criminal, civil, or administrative penalties (U.S. Code, Title 218, Section 1001).

DocuSigned by:

\*Signature:

*Pratip Bhattacharya*

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9/20/2023 | 8:54 AM CDT

Date:

## Previous/Current/Pending Support for DOD

Pisaneschi, Federica

### PREVIOUS:

<b>Title:</b>	Evaluating the Predictive Power of Novel PET Imaging Agents at the intersection of NASH and ICT response		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2022	0.12 Calendar	
	2. 2023	0.12 Calendar	
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center – QIAC Partnership in Research Grant		
<b>Grants Officer:</b>	Lori Armstrong The University of Texas MD Anderson Cancer Center 1515 Holcombe Blvd. Houston, TX 77030-4009		
<b>Performance Period:</b>	05/01/2021 – 04/30/2023		
<b>Level of Funding:</b>			
<b>Project Goals</b>	PET imaging of inflammation and apoptosis, supported by a panel of serum biomarkers, can be used to construct an integrated robust diagnostic model that predicts ICT induced liver damage in the context of NASH and melanoma		
<b>Specific Aims</b>	Aim 1: Utilize <sup>18</sup> F-TBD in combination with serum biomarkers to discriminate progression to HCC from NASH, NAFLD, and normal murine livers. Aim 2: Utilize [ <sup>18</sup> F]-FN PET in combination with serum biomarkers to discriminate progression to HCC from NASH, NAFLD, and normal murine livers.		
<b>Overlap:</b>	None		

<b>Title:</b>	PET Imaging of Apoptosis and Inflammation for the Prevention of Hepatocellular Carcinoma (HCC)		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2021	0.60 Calendar	
	2. 2022	0.60 Calendar	
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center – CABI Pilot Project Program		
<b>Grants Officer:</b>	Lori Armstrong The University of Texas MD Anderson Cancer Center 1515 Holcombe Blvd. Houston, TX 77030-4009		
<b>Performance Period:</b>	07/01/2020 – 03/31/2022		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To change the clinical practice of HCC diagnosis, and prevention through non-invasive imaging technologies to identify NASH and monitor its treatment.		

<b>Specific Aims</b>	Aim 1: Utilize [ <sup>18</sup> F]-TBD PET to distinguish NASH from NAFLD and measure its value as a predictive imaging biomarker for dietary and pharmacological intervention with Emricasan. Utilize [ <sup>18</sup> F]-TBD PET to distinguish NASH from NAFLD and measure its value as a predictive imaging biomarker for dietary and pharmacological intervention with Emricasan. Aim 2: Utilize [ <sup>18</sup> F]4FN PET to distinguish NASH from NAFLD and measure its value as a predictive imaging biomarker for dietary and pharmacological intervention with Atorvastatin
<b>Overlap:</b>	None

<b>Title:</b>	SUPR Peptides for Immune Checkpoint Blockade Imaging		
<b>Effort:</b>	<b>Year (YYYY)</b>	<b>Person Months (##.##)</b>	
	2. 2018	1.20 Calendar	
	3. 2019	1.20 Calendar	
	4. 2020	1.20 Calendar	
	5. 2021	1.20 Calendar	
	6. 2022	1.20 Calendar	
<b>Supporting Agency:</b>	CABI/G.E. In-Kind Research Award		
<b>Grants Officer:</b>	Praveena Gangakhedkar Academic Research Program Manager Praveena.gangakhedkar@ge.com (281) 536-0503		
<b>Performance Period:</b>	07/01/2016 – 06/30/2021		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To develop a SUPR peptide-based PET tracer for PD-L1 imaging and prediction of response to PD-1/PD-L1 therapies.		
<b>Specific Aims</b>	Aim 1: Generate and Characterize SUPR Peptides Against PD-L1. Aim 2: In vivo Optical Imaging of PD-L1 with Cy5-labeled SUPR Peptides. Aim 3: PET/CT Imaging of PD-L1 using <sup>18</sup> F-SUPR Peptides		
<b>Overlap:</b>	None		

<b>Title:</b>	Measuring Tumor Acidosis with PET/MRI Contrast Agents		
<b>Effort:</b>	<b>Year (YYYY)</b>	<b>Person Months (##.##)</b>	
	1. 2019	0.6 Calendar	
	2. 2020	0.6 Calendar	
<b>Supporting Agency:</b>	NIH		
<b>Grants Officer:</b>	Tatjana Atanasijevic 9900 Rockville Pike Bethesda, MD 20892 301-451-6873		
<b>Performance Period:</b>	03/01/2019 – 12/31/2020		
<b>Level of Funding:</b>			

<b>Project Goals</b>	To quantitatively measure extracellular pH (pHe) in the tumor microenvironment to assess tumor acidosis.
<b>Specific Aims</b>	Aim 1: To develop and apply PET/MRI contrast agents that measure in vivo tumor pHe.
<b>Overlap:</b>	None

<b>Title:</b>	Glutamine Imaging for the Detection of Renal Cell Carcinoma Lung Metastases		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2020	0.84 Calendar	
	2. 2021	0.84 Calendar	
<b>Supporting Agency:</b>	The University of Texas MD Anderson Cancer Center – CABI Pilot Project Program		
<b>Grants Officer:</b>	Lori Armstrong The University of Texas MD Anderson Cancer Center 1515 Holcombe Blvd. Houston, TX 77030-4009		
<b>Performance Period:</b>	04/01/2019 – 01/31/2021		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To determine if radioactive glutamine can discern ccRCC lung metastases versus inflammation in pre-clinical mouse models.		
<b>Specific Aims</b>	Aim 1: In vitro studies: Compare the sensitivity and specificity of uptake of [ <sup>11</sup> C]Gln and [ <sup>18</sup> F]FGln versus [ <sup>18</sup> F]FDG in parental and lung metastases ccRCC cell lines, normal kidney cells (HEK293) and normal lung cells (BEAS-2B).		
<b>Overlap:</b>	None		

<b>Title:</b>	In Vivo Imaging of Over-expressed Micro-RNAs for Lung Cancer Diagnosis and Prognosis		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2019	0.12 Calendar	
	2. 2020	0.12 Calendar	
	3. 2021	0.12 Calendar	
<b>Supporting Agency:</b>	IRCCS-AUSL Reggio Emilia (Italy)		
<b>Grants Officer:</b>	Dott.ssa Sara Li Dooni Infrastruttura Ricerca-Statistica, Direzione Scientifica Sara.lidonni@ausl.re.it		
<b>Performance Period:</b>	01/01/2019 – 12/31/2021		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To synthesize and radiolabel PNA based probes against miRNA-146a, -21, and -155; to validate the selectivity of anti-miRNA-probes in vitro by using tumor cell lines over-expressing the miRNAs target; to exploit the most promising anti-miRNA-probes for molecular imaging of cancer by means of PET in murine models; and to develop relationship between the two institutions in the field of research, mobility		

	of personnel, and activities for students.
<b>Specific Aims</b>	<p>Aim 1: To validate the selectivity of anti-miRNA-probes in vitro by using tumor cell lines over-expressing the miRNAs target.</p> <p>Aim 2: To exploit the most promising anti-miRNA-probes for molecular imaging of cancer by means of PET in murine models;</p> <p>Aim 3: To develop relationship between the two institutions in the field of research, mobility of personnel, and activities for students.</p>
<b>Overlap:</b>	None

<b>Title:</b>	Evaluating the Predictive Power of Novel PET Imaging Agents in the Transition from NAFLD through NASH to HCC	
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)
	1.	0.6 Calendar
	2.	0.6 Calendar
	3.	0.6 Calendar
	4.	0.6 Calendar
	5.	0.6 Calendar
<b>Supporting Agency:</b>	National Institutes of Health / The University of Texas MD Anderson Cancer Center – HCC SPORE DRP	
<b>Grants Officer:</b>	Lori Armstrong The University of Texas MD Anderson Cancer Center 1515 Holcombe Blvd. Houston, TX 77030-4009	
<b>Performance Period:</b>	09/01/2020 – 08/31/2021	
<b>Level of Funding:</b>		
<b>Project Goals</b>	To test the hypothesis that PET imaging of inflammation and apoptosis, supported by a panel of serum biomarkers, can be used to construct an integrated, robust diagnostic model that discriminates HCC progression from the following confounders: normal liver, NAFLD, and NASH livers with an AUROC of $\geq 0.85$ .	
<b>Specific Aims</b>	To test the hypothesis that PET imaging of inflammation and apoptosis, supported by a panel of serum biomarkers, can be used to construct an integrated, robust diagnostic model that discriminates HCC progression from the following confounders: normal liver, NAFLD, and NASH livers with an AUROC of $\geq 0.85$	
<b>Overlap:</b>	None	

<b>Title:</b>	PET Imaging of Neuroinflammation with 4- <sup>[18F]</sup> fluoronaphthol	
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)
	1. 2022	0.60 Calendar

	2. 2023	0.60 Calendar	
<b>Supporting Agency:</b>	QIAC Partnership in Research Grant		
<b>Grants Officer:</b>	Lori Armstrong 1515 Holcombe Blvd. Houston, TX 77030-4009 laarmstrong@mdanderson.org		
<b>Performance Period:</b>	05/01/2021 – 04/30/2023		
<b>Level of Funding:</b>			
<b>Project Goals</b>	Due to the high production of ROS and upregulation of NOX activity in microglia as a consequence of brain injuries, we hypothesize that [ <sup>18</sup> F]4FN-PET will be able to detect neuroinflammation in mouse models of radiation-induced inflammation and neurodegenerative diseases.		
<b>Specific Aims</b>	Aim 1: [ <sup>18</sup> F]4FN-PET in LPS models of neuroinflammation – a proof of concept experiment. Aim 2: [ <sup>18</sup> F]4FN-PET in a clinically relevant model of neuroinflammation due to Cranial Radiotherapy Aim 3: Intervention. Test the use of [ <sup>18</sup> F]4FN-PET as a marker for drug response using NBD-peptide treatment of CRT- induced brain inflammation.		
<b>Overlap:</b>	None		

<b>Title:</b>	[ <sup>18</sup> F]4FN PET Imaging of Innate Immunity Activation During Immunotherapy-Induced Adverse Events		
<b>Effort:</b>	<b>Year (YYYY)</b>	<b>Person Months (##.##)</b>	
	1. 2022	3.00 Calendar	
	2. 2023	3.00 Calendar	
<b>Supporting Agency:</b>	National Institutes of Health / National Cancer Institute		
<b>Grants Officer:</b>	Lin, Charles linp@mail.nih.gov		
<b>Performance Period:</b>	09/01/2022 – 08/31/2027		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To test the that imaging high redox potential RONS from activation of key innate immune cells will provide selective diagnostic utility by integrating multiple divergent biochemical inputs that converge to produce the majority of irAEs.		
<b>Specific Aims</b>	Aim 1: Characterize an Established Advanced Pre-clinical Model of irAE on a Background of Auto-immunity by [ <sup>18</sup> F]4FN PET Imaging. Aim 2: Determine the effect of Diet-Induced Metabolic Syndrome on a Novel Pre-clinical Model of irAE by [ <sup>18</sup> F]4FN PET Imaging. Aim 3: Quantify immune effector cell-associated neurotoxicity syndrome (ICANS) by [ <sup>18</sup> F]4FN PET Imaging compared to serum markers in a model of CAR T-cell therapy.		
<b>Overlap:</b>	None, Relinquished my portion to PI when I moved to UTHealth-Houston		

<b>Title:</b>	Acquired Resistance to Therapy and Iron (ARTI) Center		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2022	1.2 Calendar	
	2. 2023	1.2 Calendar	
<b>Supporting Agency:</b>	National Institutes of Health / National Cancer Institute		
<b>Grants Officer:</b>	Michael Espey 240-276-6230 SP@NIH.GOV		
<b>Performance Period:</b>	09/20/2022 – 08/31/2027		
<b>Level of Funding:</b>			
<b>Project Goals</b>	The overall goal of the MIC is to implement and manage the precise imaging support infrastructure (personnel, reagents, animals, software, and customized hardware resources) needed to support each project in the ARTI Center.		
<b>Specific Aims</b>	<p>Aim 1: Collaborate with the ARTI Center and individual investigators in developing imaging protocols and analyses with appropriate biological and biochemical controls, robust test-retest analysis, and imaging statistical support when appropriate.</p> <p>Aim 2: Provide timely access to unique plasmid-based reporters, reporter animals, PET reagents, and fluorescent biosensors, while leveraging internal core and MD Anderson core expertise to modify reagents when appropriate to meet ARTI Center needs.</p> <p>Aim 3: Provide timely service, training, access to customized macro- and microscopic imaging systems, premium image analytics, and custom imaging analysis.</p>		
<b>Overlap:</b>	None, Relinquished my portion to PI when I moved to UTHealth-Houston		

<b>Title:</b>	Membrane Permeant Peptides for Imaging Cell Function		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2019	1.2 Calendar	
	2. 2020	1.2 Calendar	
	3. 2021	1.2 Calendar	
	4. 2022	1.2 Calendar	
	5. 2023	1.2 Calendar	
<b>Supporting Agency:</b>	National Institutes of Health / National Eye Institute		
<b>Grants Officer:</b>	Hemin R. Chin 31 Center Drive MSC 2510 Bethesda, MD 20892-2510		
<b>Performance Period:</b>	09/30/2018 – 08/31/2023		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To conduct quantitative pre-clinical testing in advanced glaucoma models to monitor disease progression and quantify dietary (niacin) and gene therapy (Nmnat1) interventions, as well as toxicology analysis, and metabolite profiling. We will advance a lead peptide toward the clinic through a statistically-robust non-human primate model of		

	glaucoma.
<b>Specific Aims</b>	<p>Aim 1: Test the hypothesis that our caspase-3-activated cell-penetrating peptides can quantify apoptosis in pre-clinical rodent models of RGC degeneration and treatment interventions.</p> <p>Aim 2: Rigorously test the hypothesis that our lead caspase-3- activated cell-penetrating peptide can quantify apoptosis in an advanced non-human primate model of glaucoma.</p> <p>Aim 3: Perform advanced pre-clinical characterization of a lead activatable cell-penetrating peptide in preparation for eIND filing</p>
<b>Overlap:</b>	None, Relinquished my portion to PI when I moved to UTHHealth-Houston

**CURRENT:**

<b>Title:</b>	Identifying Different Metabolic Sub-types of Prostate Cancer for Early Therapeutic Assessment		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2022	2.4 Calendar	
	2. 2023	1.2* Calendar	
	3. 2024	1.2* Calendar	
	*Pending new sub contract Notice		
<b>Supporting Agency:</b>	DoD		
<b>Grants Officer:</b>	Joshua McKean 820 Chandler St. Fort Detrick, MD 21702 joshua.d.mckean3.civ@mail.mil		
<b>Performance Period:</b>	09/2021-08/2024		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To develop personalized metabolic imaging modality to target treatment strategies of different sub-types of PCa and to interrogate different metabolic factors by imaging that are associated with a particular sub-type.		
<b>Specific Aims</b>	<p>Aim 1: To perform HP-<sup>13</sup>C Pyruvate MRI, [<sup>18</sup>F]FPIA -PET imaging in four well characterized patient derived xenograft (PDX) murine models of prostate cancer in vivo before and after the antiandrogen therapy with Enzalutamide to image both alteration in glycolytic and fatty acid oxidation pathways after treatment response.</p> <p>Aim 2: To elucidate the metabolic drivers of ADT resistance. We will analyze ADT sensitive and ADT resistant PDX utilizing NMR-spectroscopy combined with mass spectrometry-based metabolomics, immunohistochemistry and transcriptome profiling. Using this approach, we expect to delineate pathways of ADT resistance.</p> <p>Aim 3: To image overcoming the ADT resistance by targeting Monocarboxylate Transporter (MCT) pathway (via MCT inhibition with the clinical drug, Syrosingopine) by</p>		

	performing HP- <sup>13</sup> C Pyruvate MRI in two well characterized patient derived xenograft (PDX) murine models of prostate cancer in vivo. [ <sup>18</sup> F]FPIA -PET
<b>Overlap:</b>	None

<b>Title:</b>	Imaging Metabolic Reprogramming in Prostate Cancer		
<b>Effort:</b>	<b>Year (YYYY)</b>	<b>Person Months (##.##)</b>	
	1. 2023	2.4 Calendar	
	2. 2024	2.4 Calendar	
	3. 2025	2.4 Calendar	
<b>Supporting Agency:</b>	Cancer Prevention and Research Institute of Texas		
<b>Grants Officer:</b>	Patricia Moore, Program Manager for Research, CPRIT (www.cpriti.state.tx.us)		
<b>Performance Period:</b>	03/2022-02/2025		
<b>Level of Funding:</b>			
<b>Project Goals</b>	This proposal will build up on these findings, investigating by PET and HP-MRI how carnitine metabolism is modulated in these PDXs before and after Enzalutamide-treatment. Furthermore, we will attempt to overcome Enzalutamide resistance by targeting MCT and ENO2 proteins with inhibitors, Syrosingopine and SF2312 respectively, and image this transformation from resistance to sensitization by HP-MRI and PET.		
<b>Specific Aims</b>	<p>Aim 1: HP MRI and PET imaging. To perform <sup>13</sup>C-Pyruvate, <sup>15</sup>N-L-Carnitine-d6 HP-MRI and [<sup>18</sup>F]FPIA-PET imaging in six well characterized patient derived xenograft (PDX) murine models of prostate cancer in vivo before and after the antiandrogen therapy with Enzalutamide to image both alteration in glycolytic and fatty acid oxidation pathways after treatment response.</p> <p>Aim 2: Ex vivo analyses. To understand the mechanistic basis of the success and/failure of HP-MRI and FPIA-PET in imaging specific PDX sub-types of PCa xenografts through metabolic and transcriptome profiling and analyses of key protein expressions in the metabolic pathways.</p> <p>Aim 3: Overcoming resistance. To image overcoming the Enzalutamide resistance by targeting Monocarboxylate Transporter (MCT) (via MCT inhibition with the clinical drug, Syrosingopine) and Enolase 2 (ENO2) (via ENO2 inhibition with natural phosphonate antibiotic, SF2312) and define whether MCT and/or ENOs are viable targets for potential combination therapy with anti-androgens.</p>		
<b>Overlap:</b>	None		

**PENDING**

<b>Title:</b>	Imaging of cranial radiotherapy neuroinflammatory sequelae by Positron Emission Tomography		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2024	2.4 Calendar	
	2. 2025	2.4 Calendar	
	3. 2026	2.4 Calendar	
	4. 2027	2.4 Calendar	
<b>Supporting Agency:</b>	Cancer Prevention and Research Institute of Texas		
<b>Grants Officer:</b>	Patricia Moore, Program Manager for Research, CPRIT (www.cprit.state.tx.us)		
<b>Performance Period:</b>	03/01/2023 – 0/28/2027		
<b>Level of Funding:</b>			
<b>Project Goals</b>	To detect cranial radiotherapy induced neuroinflammation early and assess cognitive impairment by Positron Emission Tomography.		
<b>Specific Aims</b>	<p>Aim 1: To evaluate [<sup>18</sup>F]4FN-PET in clinically relevant, whole brain and focal preclinical models of neuroinflammation due to CRT and to compare [<sup>18</sup>F]4FN with <sup>18</sup>F-PBR06. Neuroinflammation as detected by PET will be correlated to cognitive impairment.</p> <p>Aim 2: To develop <sup>18</sup>F-COG, a new radiopharmaceutical that targets TREM2, and to test it for TREM2-mediated uptake in cellular and animal models. <sup>18</sup>F-COG will then be tested in CRT-induced neuroinflammation models.</p> <p>Aim 3: To use PET to monitor the amelioration of neuroinflammation via treatment with TAD-NBD-peptides, and consequent reduction of cognitive impairment</p>		
<b>Overlap:</b>	None		

<b>Title:</b>	Targeting the androgen receptor with radiation: an alternative approach to treat prostate cancer		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2025	0.6 Calendar	
	2. 2026	0.6 Calendar	
<b>Supporting Agency:</b>	DoD		
<b>Grants Officer:</b>	Not Available		
<b>Performance Period:</b>	05/01/2024 – 04/30/2026		
<b>Level of Funding:</b>			
<b>Project Goals</b>	This study will create radiolabeled small molecule ligands that have the chemical feature of bearing a stable <sup>77</sup> Br-label that can be translocated by AR into the nucleus and to the DNA, where Auger emissions will damage DNA and subsequently induce cell death		
<b>Specific Aims</b>	<p>Aim1. To synthesize and test [<sup>77</sup>Br]BrLGD-3303.</p> <p>Aim2. To synthesize and test [<sup>77</sup>Br]BrJQ1</p>		
<b>Overlap:</b>	None		


<b>Title:</b>	Radiolabeling compound libraries for high-throughput screening of novel radiopharmaceuticals		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2024	0.24 Calendar	
<b>Supporting Agency:</b>	DropletPharm Inc / NIH		
<b>Grants Officer:</b>	Not Available		
<b>Performance Period:</b>	04/01/2024 – 03/31/2025		
<b>Level of Funding:</b>			
<b>Project Goals</b>	The proposal is focused on further developing a high-throughput radiochemistry that my lab developed and the company is planning to commercialize.		
<b>Specific Aims</b>	No aims assigned to UTHealth		
<b>Overlap:</b>	None		

<b>Title:</b>	Targeted Cell Death Imaging in Heart Transplantation		
<b>Effort:</b>	Year (YYYY)	Person Months (##.##)	
	1. 2025	1.2 Calendar	
	2. 2026	1.2 Calendar	
	3. 2027	1.2 Calendar	
<b>Supporting Agency:</b>	Rubicon Biotechnology LLC / NIH		
<b>Grants Officer:</b>	Not Available		
<b>Performance Period:</b>	04/01/2024 – 03/31/2027		
<b>Level of Funding:</b>			
<b>Project Goals</b>	Perform radiolabeling of the fragment with <sup>18</sup> Faluminium fluoride and will provide radiochemical characterization of the new probe (i.e. radiochemical purity, chemical purity, molar/specific activity, shelf stability). Once the radiochemistry is optimized, the probe will be radiosynthesized for animal studies.		
<b>Specific Aims</b>	Perform radiolabeling of the fragment with <sup>18</sup> Faluminium fluoride and will provide radiochemical characterization of the new probe (i.e. radiochemical purity, chemical purity, molar/specific activity, shelf stability). Once the radiochemistry is optimized, the probe will be radiosynthesized for animal studies.		
<b>Overlap:</b>	None		

I, PD/PI or other senior/key personnel confirm that I:

- Certify that the current and pending support provided on the application is current, accurate and complete;
- Agree to update such disclosure at the request of the agency prior to the award of support and at any subsequent time the agency determines appropriate during the term of the award; and
- Have been made aware of the requirements under Section 223(a)(1) of this Act.10 DOD General Application Instructions 18

False, fictitious, or fraudulent statements or claims may result in criminal, civil, or administrative penalties (U.S. Code, Title 18, Section 1001)

Signature: fpisaneschi  Digitally signed by fpisaneschi  
Date: 2023.09.21 08:50:05 -05'00'

Date: \_\_\_\_\_