

AD-A045 763

ARMED FORCES FOOD SCIENCE ESTABLISHMENT SCOTTSDALE (A--ETC F/6 6/8  
THE EFFECTS OF FEEDING FORMALDEHYDE STABILIZED MEAT TO RATS.(U)  
JUL 77 J R CASLEY-SMITH  
AFFSE-3/77

UNCLASSIFIED

NL

| OF |  
AD  
AO45763



END  
DATE  
FILMED  
11 - 77  
DDC

AD A 045763

UNCLASSIFIED

12

AFFSE REPORT 3/77

AR No. 000786



Department of Defence  
Defence Science and Technology Organisation  
Armed Forces Food Science Establishment  
Scottsdale, Tasmania

AFFSE REPORT 3/77

© COMMONWEALTH OF AUSTRALIA, 1977

# The Effects of Feeding Formaldehyde Treated Meat to Rats

AD No. \_\_\_\_\_  
DDC FILE COPY



DDC  
RECEIVED  
OCT 21 1977  
B

J. R. CASLEY-SMITH

APPROVED  
FOR PUBLIC RELEASE

Commonwealth of Australia  
July, 1977

**APPROVED  
FOR PUBLIC RELEASE**

**THE UNITED STATES NATIONAL  
TECHNICAL INFORMATION SERVICE  
IS AUTHORISED TO  
REPRODUCE AND SELL THIS REPORT**

UNCLASSIFIED

DEPARTMENT OF DEFENCE  
ARMED FORCES FOOD SCIENCE ESTABLISHMENT

DDC  
RECEIVED  
OCT 21 1977  
RECEIVED  
B

14  
AFFSE REPORT - 3/77

6  
THE EFFECTS OF FEEDING FORMALDEHYDE  
STABILISED MEAT TO RATS. (U)

10  
J.R. CASLEY-SMITH

11  
Jul 77

COMMONWEALTH OF AUSTRALIA, 1977

9  
Technical rept.

12  
11p.

S U M M A R Y

Meat was prepared by pre-treating it with formaldehyde and phosphate (pH 11), but without subsequent cooking and drying in order to preserve as much formaldehyde as possible. It was fed to 50 rats for 5 days out of the week, for 12 months. A control group of rats received normal meat. The diets were supplemented with rat nuts (including the remaining 2 days of the week). Histological sections of the brain, liver, stomach, ileum, kidney and skeletal muscle were examined by light microscopy and formaldehyde levels in these tissues were estimated biochemically. No significant differences were found. In addition, there were no significant differences between the growth rates and final weights of either group. Both groups appeared equally active and healthy. The amount of formaldehyde consumed was relatively much greater than that which would be eaten by men consuming formaldehyde-stabilised meat.

It is therefore concluded that there is very little likelihood of meat, treated in such a manner, having any deleterious effect on men. (U)

POSTAL ADDRESS: The Director,  
Armed Forces Food Science Establishment,  
P.O. Box 147,  
Scottsdale, Tasmania, 7254

H10 231

DISTRIBUTION STATEMENT A  
Approved for public release;  
Distribution Unlimited

LB

DOCUMENT CONTROL DATA SHEET

UNCLAS

1. DOCUMENT NUMBERS

a. AR Number: 000786  
b. Document Series and Number: -  
c. Report Number: 3/77

2. SECURITY CLASSIFICATION

a. Complete document:  
Unclas  
b. Title in isolation:  
Unclas  
c. Summary in isolation:  
Unclas

3. TITLE: The Effects of Feeding Formaldehyde Stabilised Meat to Rats

4. PERSONAL AUTHOR:

Casley-Smith, J. R.

5. DOCUMENT DATE:

July, 1977

6. TYPE OF REPORT AND PERIOD COVERED:

Technical Report

7. CORPORATE AUTHOR:

Armed Forces Food Science  
Establishment,  
Scottsdale, Tasmania, Aust.

8. REFERENCE NUMBERS:

a. Task: FSE 76/036  
b. Sponsoring Agency:  
DOD (Army)

9. COST CODE: 241

10. IMPRINT:

AFFSE - July, 1977.

11. COMPUTER PROGRAM:

-

12. RELEASE LIMITATIONS:

Approved for public release.

12-0 OVERSEAS: N.O.  P.R.  1  A  B  C  D  E

13. ANNOUNCEMENT LIMITATIONS:

-

14. DESCRIPTORS:

Meat, Rats, Formaldehyde  
Animal Nutrition Physiology,  
Growth.

15. COSATI CODES:

0601-0620

DOCUMENT CONTROL DATA SHEET

(Continued)

UNCLAS

16. SUMMARY:

Meat was prepared by pre-treating it with formaldehyde and phosphate (pH 11), but without subsequent cooking and freeze-drying in order to preserve as much formaldehyde as possible. This was fed to 50 rats for 5 days out of the week, for 12 months. Fifty control rats received normal meat. The diets were supplemented with rat nuts (including the remaining 2 days of the week). Histological sections of the brain, liver, stomach, ileum, kidney and skeletal muscle were examined by light microscopy and formaldehyde levels in these tissues were estimated biochemically. No significant differences were found. In addition, there were no significant differences between the growth rates and final weights of either group. Both groups appeared equally active and healthy. The amount of formaldehyde consumed was relatively much greater than that which would be eaten by men consuming formaldehyde-stabilised meat.

It is therefore concluded that there is very little likelihood of meat, treated in such a manner, having any deleterious effect on men. (U)

UNCLAS

ACCESSION for	
NTIS	White Section <input checked="" type="checkbox"/>
DDC	Black Section <input type="checkbox"/>
UNANNOUNCED	<input type="checkbox"/>
JUSTIFICATION	<input type="checkbox"/>
BY _____	
DISTRIBUTION/AVAILABILITY CODES	
Dist.	AVAIL. CODE or SPECIAL
A	

C O N T E N T S

	<u>Page No.</u>
Introduction    ...    ...    ...    ...    ...    ...	1
Experimental    ...    ...    ...    ...    ...    ...	3
Results    ...    ...    ...    ...    ...    ...	4
Conclusions    ...    ...    ...    ...    ...    ...	5
Acknowledgements .    ...    ...    ...    ...    ...	5
References    ...    ...    ...    ...    ...    ...	6
Distribution List	

THE EFFECTS OF FEEDING FORMALDEHYDE  
STABILISED MEAT TO RATS

by

J.R. CASLEY-SMITH

I N T R O D U C T I O N

The four previous reports (Casley-Smith, 1972; 1975; Casley-Smith and Ehmann, 1973; Casley-Smith et al., 1974) and a review (Casley-Smith, 1973) have covered the general background to this work and the results of experiments up to the end of 1975. The present report covers the final stage of this work which was the direct responsibility of the University of Adelaide, viz. a long-term feeding trial of the product in order to ensure its harmlessness.

The basic problem, outlined by Hutchinson (1970), was to improve the quality of freeze-dried steak (and other large pieces of meat) so that they would be acceptable on reconstitution and could thus be used in ration packs. While normal freeze-drying methods have been found to be very good on certain materials - especially those which come as small particles, they do not work well on large pieces of meat. These on reconstitution, tend to be tough, of lowered satiety value, and tend to remain wet on the outside and dry on the inside, with water being easily expressed from them on slight pressure (Downman, 1971; Hutchinson, 1970; Venkata-Raman, 1971). While this would be of little consequence if the ration packs were only to be used for one or two days, it is considered that they must be suitable for use over periods of 5-7 days.

The food consumption habits and preferences of Australians indicate a desire to have slices or large pieces of meat available in the diet. Freeze-drying enables large pieces of various meats to be incorporated into combat ration packs. Thus research to improve the texture of reconstituted freeze-dried beef has an immediate practical goal. Alcoholic dehydration has been suggested also as an alternative to freeze dehydration (Casley-Smith et al 1974, Casley-Smith, 1975).

The underlying reasons for the difficulties with freeze-drying are unknown. They are likely to remain so until we have a much better understanding of the basic physics and chemistry of water in the tissues and how these are affected by this process and by reconstitution. However, it was considered (Casley-Smith, 1970) that it might be quite possible to find an ad hoc solution to the immediate problems by applying techniques of tissue stabilisation developed for electron microscopy. Here, it is imperative that the fine structure, and frequently the "fine physics and chemistry", of the tissue be preserved, with special emphasis on the cellular

membranes (Sjöstrand, 1967). It was suggested that the lack of stabilisation of these membranes might be the factor chiefly responsible for the difficulties experienced in freeze-drying (Casley-Smith, 1970). Relatively slow freezing rates have to be used, which are highly likely to cause multiple ruptures of many of the cellular membranes because of the formation of large ice crystals. Thus the contents of the cells would no longer be retained in their original compartments after reconstitution. They would be free to migrate from their normal sites so that the whole muscle would resemble a sponge containing meat soup. These suggestions have indeed been confirmed during this project. Electron microscopy has revealed that such ruptures do occur and that they are largely prevented by the appropriate pretreatment (Casley-Smith, 1973; Casley-Smith and Ehmann, 1973). The previous reports also show that such a pretreatment considerably improves the acceptability of the final product.

The pretreatment consists of fixing the tissues with low concentrations of formaldehyde and removing the unreacted reagent by washing in water which also includes phosphate buffers (pH 11) to increase the juiciness and water-holding-capacity (WCP) of the product. When preparing specimens for frozen-section electron microscopy, it is well known that such fixation renders the cells much less susceptible to ice-crystal damage, and probably more natural in appearance on reconstitution after drying following freezing (Sjöstrand, 1967; Bernhard and Leduc, 1967; Bernhard and Viron, 1971; Tokuyasu, 1973). The fixatives bind the proteins of the cells, and their membranes, together with co-valent bonds. It is thus much harder for ice-crystals to rupture cellular membranes and the osmotically-active constituents of the various compartments are much less likely to leave them on reconstitution. Hence water will tend to re-enter these compartments to extents roughly equivalent to the amounts present originally.

Formaldehyde has been used in our work for a number of reasons. It is known to penetrate the tissues very rapidly (Dempster, 1960; Hopwood, 1967, 1969). It is very effective in cross linking the proteins, producing the finest pores in the membranes of any of the conventional fixatives (Sjöstrand, 1967). These pores are quite distinct from those between the cells, upon whose width depends the rapidity of water re-entry during reconstitution - formaldehyde does not alter these intercellular pores (Casley-Smith, 1975). Formaldehyde does have the disadvantage of making the material tougher, but it has been shown that this can be kept to minor levels, which are not significant compared with its advantages (Casley-Smith, 1975; Casley-Smith et al., 1974). There was, however, one potentially most significant disadvantage - it might act as a poison. Certainly it gained an unsavoury reputation when it was used as a preservative by unscrupulous sausage manufacturers early in this century, but this was when it was simply used in high concentrations as a preservative, with no attempt to remove the unreacted reagent. In fact, it has been fed daily to sheep in relatively far greater doses than likely to be received by men eating our product, for periods of years, with no ill effects (Durand, 1971). Also, we have shown that meat

normally contains a small amount of formaldehyde, which actually exceeds the levels which remain after our treatment (Casley-Smith and Ehmann, 1973; Casley-Smith et al., 1974). Washing, cooking and freeze-drying remove the unreacted formaldehyde. The reacted formaldehyde is attached to the proteins, and is digested and used in the body just like any normal food without ill-effects, being converted almost entirely to water and carbon dioxide in a short time (Durand, 1971; Warner, 1972 - personal communication).

However, sheep are not men. Since the gut of a ruminant is indeed so different from that of an omnivore, it was decided to carry out a long-term feeding trial in rats before undertaking extensive testing in men.

#### E X P E R I M E N T A L

One hundred young (~150 g) male rats were randomly divided into two equal groups. They were placed, five to a cage, in air-conditioned, semi-sterile rooms. (The rats were specific-pathogen-free, of the Wistar strain.) Water and rat nuts (W. Charlick Pty. Ltd.) were given ad lib. Five grams of formaldehyde treated, or normal, meat were given to each rat for 5 days of the week. This regime continued for 12 months. Since the meat was cut into small (~1 cm<sup>3</sup>) pieces, there was plenty for all and each animal received approximately the same amount. On the days when the meat was fed, the animals ate this at once, in preference to the rat nuts. Thus these latter supplemented the meat rather than the reverse, but the rats received about 50% of their nourishment from the nuts which provided the requisite vitamins and minerals.

The meat was all obtained from a 3 year-old Hereford steer, cut into ~1 cm<sup>3</sup> pieces and either left untreated, or treated with formaldehyde. For this latter, the pieces were placed in 0.3 g/100 ml formaldehyde in a buffer consisting of 0.274 g/100 ml Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> and 0.226 g/100 ml NaH<sub>2</sub>PO<sub>4</sub>, which gave pH 11. Unlike the meat for humans, no sucrose was added. The meat was fixed for 4 hours at 20°C. It was then washed in a second solution for 12 hours. (This was similar to the first, but the formaldehyde was omitted.) In the first case the volume of the solution was 10-20 times that of the meat; in the second it was 50 times. In order to ensure that the final product would contain more formaldehyde than normal, the cooking and freeze-drying were omitted. These processes normally reduce the residual amounts of formaldehyde to about 20% of the amount remaining after the washing; these former levels are about 25% of the normal amounts which occur in untreated meat. Thus the present level was ~2,000µg/g, in place of ~50 µg/g which is usually found after the whole processing. The amount of meat eaten by the rats was approximately equal to 1.25 kg of meat eaten by a 70 kg man each day of the week, but with a formaldehyde content 4 times greater than normal.

Various procedures were used to estimate the effects of the

treated meat on the rats. They were weighed, and the total weights and rates of growth were compared. Samples of the brain, liver, kidneys, stomach, ileum and skeletal muscle were taken and the amounts of formaldehyde in them were determined by the method developed earlier (Casley-Smith, 1972; Casley-Smith and Ehmann, 1973). Samples of these organs were also fixed in Zenker's fixative (which does not contain formaldehyde), processed for light microscopy, and examined. These were randomised, examined without the observer knowing to which group they belonged, and graded for normality.

### R E S U L T S

Both groups appeared equally active and healthy. At the end of the 12 months the mean weight of the animals eating normal meat was 300.1 (5.14) g; that of those eating the treated meat was 294.2 (5.61) g. (The Standard Errors of the Means are given in brackets after the Means.) There are no significant differences between these two groups using the F or the t-tests. The growth gains over the 12 months were 149 (7.3) g for the untreated group and 145 (5.8) g for the treated one: again neither the F nor the t-tests are significant.

The results of the biochemical estimates of formaldehyde are shown below (in  $\mu\text{g/g}$ ):

	<u>Normal Meat</u>	<u>Formaldehyde Treated Meat</u>
Brain	823 (62)	934 (79)
Liver	1310 (82)	1260 (94)
Kidney	1270 (75)	1300 (83)
Stomach	1050 (67)	1110 (75)
Ileum	981 (74)	1300 (69)
Muscle	1000 (50)	950 (76)

With the exception of the ileum, where the t-test is very significant ( $P < 0.001$ ), none of the other t-tests and none of the F-tests are significant. Since the specimens are not washed in any way - except for a very brief rinse to remove the gross contents of the gut - it is highly likely that the increase found in the ileum was caused by residual formaldehyde from the ingested meat, rather than being actually present in the lining of the ileum. This did not appear in the stomach, perhaps because the meat was less finely divided here so that the wash removed all of it.

It is clear that eating the treated meat for a year did not cause significant increases in the amounts of residual formaldehyde in the various organs.

The histological examinations also revealed no significant differences between the two groups. In all cases the organs appeared normal. Since it was found that there was an increase in the numbers of small round cells in the gut of the sheep (Durand, 1971), a special search was made for signs of chronic inflammation. None were found.

#### C O N C L U S I O N S

It is evident that the rats suffered no ill effects from this diet. This is similar to the sheep which ate even larger relative amounts of formaldehyde for twice as long (Durand, 1971). Here, even the mild evidence of chronic inflammation of the gut, which was found in the sheep, was not observed. Thus formaldehyde treatment of meat, followed by cooking and freeze-drying, will give a product which can be continuously consumed by men for very long periods without ill effects.

Taste-panel results of the product have been favourable, it has been shown to be non-toxic and it is a process which is relatively simple and inexpensive.

#### A C K N O W L E D G E M E N T S

I am most grateful to Dr. N. B. Piller, Mrs. A. H. Vincent, B.Sc., Miss M. Quin and Messes K. W. J. Crocker and W. G. Smith for the help they have given with this phase of the project.

R E F E R E N C E S

- Bernhard, W. and Leduc, E. H. (1967). Ultrathin frozen sections. I. Methods and ultrastructural preservation. *J. Cell Biol.* 34, 757.
- Bernhard, W. and Viron, A. (1971). Improved techniques for the preparation of ultrathin frozen sections. *J. Cell Biol.* 49, 731.
- Casley-Smith, J. R. (1970). In "Status of Water in Biological Materials", Report No. 2/70. Armed Forces Food Science Establishment, Scottsdale, Tasmania, p. 15.
- Casley-Smith, J. R. (1972). Minor Report No. CR/1. Armed Forces Food Science Establishment, Scottsdale, Tasmania.
- Casley-Smith, J. R. (1973). The structure of muscle cells, the distribution of water in them, and how these affect freeze-drying. In "Report of a Convention on Freeze Drying", Report No. 2/73, Armed Forces Food Science Establishment, Scottsdale, Tasmania, pp 6-18.
- Casley-Smith, J. R. (1975). Minor Report No. CR/7. Armed Forces Food Science Establishment, Scottsdale, Tasmania.
- Casley-Smith, J. R. and Ehmann, H. F. W. (1973). Minor Report No. CR/5. Armed Forces Food Science Establishment, Scottsdale, Tasmania.
- Casley-Smith, J. R., Ehmann, H. F. W., Venkata-Raman, S. and Driver, G. E. (1974). Minor Report No. CR/6. Armed Forces Food Science Establishment, Scottsdale, Tasmania.
- Dempster, A. (1960) - cited Hopwood (1967).
- Dowman, I. D. (1971) Report No. 1/71. Armed Forces Food Science Establishment, Scottsdale, Tasmania.
- Durand, M. R. E. (1971). Rural Research in the C.S.I.R.O., June, 1971.
- Hopwood, D. (1967). Some aspects of fixation with glutaraldehyde. *J. Anat.* 101, 83.
- Hopwood, D. (1969). Fixatives and fixation: a review. *J. Histochem. Cytochem.* 1, 323.
- Hutchinson, R. C. (1970). In "Status of Water in Biological Materials", Report No. 2/70. Armed Forces Food Science Establishment, Scottsdale, Tasmania.
- Sjöstrand, F. S. (1967). "Electron Microscopy of Cells and Tissues", Vol. 1. Academic Press, N.Y. and Lond.
- Tokuyasu, K. T. (1973). A technique for ultracryotomy of cell suspensions and tissues. *J. Cell Biol.* 57, 551.
- Venkata-Raman, S. (1971). Minor Report No. MR/80. Armed Forces Food Science Establishment, Scottsdale, Tasmania.

D I S T R I B U T I O N   L I S T

No. of Copies

DEFENCE ESTABLISHMENTS

Department of Defence,  
Russell Offices,  
CANBERRA.    A.C.T.    2600

- (a) Secretary 6
- (b) Chief Defence Scientist 1
- (c) Executive Controller,  
Australian Defence Scientific Service 1
- (d) Controller,  
Service Laboratories and Trials Division 2
- (e) Superintendent,  
Defence Science Administration Division 1
- (f) JIO (DDSTI) 1

Department of Defence (Army Office),  
Russell Offices,  
CANBERRA.    A.C.T.    2600

- (a) Directorate of Army Development 2
- (b) Director of Infantry 2
- (c) Director of Catering 1
- (d) Scientific Adviser - Army 1

Department of Defence (Air Force Office)  
Russell Offices,  
CANBERRA.    A.C.T.    2600

- (a) Air Force Scientific Adviser 1
- (b) Director of Catering and Services  
(DCATSERV-AF) 2

Department of Defence,  
Campbell Park Offices,  
CANBERRA.    A.C.T.    2601

- (a) Defence Information Services Branch 16
- (b) Defence Central Library 1

Senior Librarian,  
Aeronautical Research Laboratories,  
P.O. Box 4331,  
MELBOURNE.    Vic.    3001

Senior Librarian,  
Weapons Research Establishment,  
G.P.O. Box 2151,  
ADELAIDE.    S.A.    5001

Royal Australian Navy Research Laboratory,  
P.O. Box 706,  
DARLINGHURST.    N.S.W.    2010

No. of Copies

DEFENCE ESTABLISHMENTS (Con't)

Tropical Trials Establishment, Box 931, INNISFAIL. Qld. 4860	1
Directorate of Supply (Technical Services), Department of Defence (Army Office), G.P.O. Box 1932R, MELBOURNE. Vic. 3001	2
Supply Division, HQ Logistic Command, St. James Plaza, G.P.O. Box 1932R, MELBOURNE. Vic. 3001	1
Directorate of Army Health Services, G.P.O. Box 1932R, MELBOURNE. Vic. 3001	2
The Director of Naval Victualling, Department of Defence (Navy Office), Victoria Barracks, St. Kilda Road, MELBOURNE. Vic. 3004	2
Headquarters, Field Force Command, Victoria Barracks, PADDINGTON. N.S.W. 2021	3
Headquarters, Logistic Command, St. James Plaza, G.P.O. Box 1932R, MELBOURNE. Vic. 3001	3
Headquarters, Training Command, Victoria Barracks, PADDINGTON. N.S.W. 2021	1
Headquarters Operational Command, (Staff Officer Catering), RAAF, PENRITH. N.S.W. 2750.	1
Headquarters, Support Command (CE04/E41), RAAF, Defence Centre, 366 St. Kilda Road, MELBOURNE. Vic. 3000	1
Catering Supervisor, Log Branch, HQ 1 MD, Victoria Barracks, BRISBANE. Qld. 4000	1

	<u>No. of Copies</u>
<u>DEFENCE ESTABLISHMENTS (Con't)</u>	
Headquarters, 2 Military District, Victoria Barracks, SYDNEY. N.S.W. 2000	1
Headquarters, 3 Military District, Victoria Barracks, MELBOURNE. Vic. 3000	1
Headquarters, 4 Military District, Keswick Barracks, ADELAIDE. S.A. 5035	1
51 Sup Bn, Irwin Barracks, KARRAKATTA. W.A. 6010	1
Headquarters, 6 Military District, Anglesea Barracks, HOBART. Tas. 7002	1
Headquarters, 7 Military District, Larrakeyah Barracks, DARWIN. N.T. 5790	1
Commandant, Royal Military College, DUNTROON. A.C.T. 2600	1
Commandant, Australian Staff College, FORT QUEENSCLIFFE. Vic. 3255	2
Commanding Officer/Chief Instructor, RAAOC Centre, Milpo, BANDIANA. Vic. 3662	2
Commandant, RAAF Staff College, RAAF Base, Fairbairn, CANBERRA. A.C.T. 2600	2
Director, Institute of Aviation Medicine, POINT COOK. RAAF. Vic. 3029	2
<u>CIVILIAN ESTABLISHMENTS</u>	
The Librarian, CSIRO, Tasmanian Regional Laboratory, Stowell Avenue, HOBART. Tas. 7000	1

No. of Copies

CIVILIAN ESTABLISHMENTS (Con't)

The Librarian, CSIRO Division of Food Research P.O. Box 52, NORTH RYDE. N.S.W. 2113	1
The Librarian, CSIRO Dairy Research Laboratory, P.O. Box 20, HIGHETT. Vic. 3190	1
Government Analyst, Department of Science, CANBERRA. A.C.T. 2600	1
Dr. R. I. Garrod, Department of Education and Science, P.O. Box 826, CANBERRA. A.C.T. 2601	1
The Central Library, Department of Health, P.O. Box 100, WODEN. A.C.T. 2606	2
Dr. R. C. Hutchinson, Derwentlaken Road, GREGSON. Tas. 7402	1
Head, Library Department, Central Library, Royal Melbourne Institute of Technology, Floor 1, Casey Wing, 368 Swanston Street, MELBOURNE. Vic. 3000	1
Tasmanian Collection State Library of Tasmania, 91 Murray Street, HOBART. Tas. 7000	1
Serials Section, State Library of Tasmania, 91 Murray Street, HOBART. Tas. 7000	1
Reference Section, Hellyer Regional Library, Alexander Street, BURNIE. Tas. 7320	1
Director, Food Preservation Laboratory, Department of Primary Industries, HAMILTON. Qld. 4000	1
Head, Food School, East Sydney Technical College, Forbes Street, DARLINGHURST. N.S.W. 2010	1

No. of Copies

CIVILIAN ESTABLISHMENTS (Con't)

Professor R. A. Edwards, 1  
Head,  
School of Food Technology,  
University of New South Wales,  
P.O. Box 1,  
KENSINGTON. N S.W. 2033

The Medical Library, 1  
Flinders University of South Australia,  
BEDFORD PARK. .S.A. 5042

OVERSEAS ESTABLISHMENTS

BRITAIN

Australian Army Representative, 2  
Australia House,  
Strand,  
London, WC2 B4LA, England.

Ministry of Defence (Navy), 2  
Directorate of Supplies & Transport  
(General and Victualling),  
Empress State Building,  
Lillie Road,  
London, SW6, England

Adviser in Nutrition, 1  
AMD5, (Army Dept), Ministry of Defence,  
Landsdowne House, Berkeley Square,  
London, W1, England.

Deputy Chief Scientist (Army), 2  
Sag (a) 3D,  
Ministry of Defence,  
Main Building, Whitehall Gardens,  
London SW 1A 2HB, England.

The Director, 1  
Army Personnel Research Establishment,  
C/- Royal Aircraft Establishment,  
Farnborough, Hants, England.

The Information Officer, 1  
British Food Manufacturing Industries  
Research Association,  
Randalls Road, Leatherhead,  
Surrey, KT227RY, England.

The Librarian, 1  
Food Research Institute,  
Colney Lane,  
Norwich NOR 70F, England.

Superintendent, 6  
Foods and Nutrition Division,  
Laboratory of the Government Chemist,  
Cornwall House, Stamford Street,  
London, SE1 9NQ, England.

OVERSEAS ESTABLISHMENTS (Con't)

BRITAIN (Con't)

The Librarian, 1  
Meat Research Institute,  
Langford. Bristol. England.

Dr. D. J. McWeeny, 1  
Ministry of Agriculture, Fisheries and Food,  
Food Science Unit,  
Food Research Institute,  
Colney Lane, Norwich,  
Norfolk, NOR 70F, England.

Mr. J. F. Hearne, 3  
Food Standards Science and Safety Division,  
Ministry of Agriculture, Fisheries and Food,  
Great Westminster House, Horseferry Road,  
London, SW1P2AE, England.

The Librarian, 1  
National College of Food Technology,  
University of Reading,  
St. George's Avenue, Weybridge,  
Surrey, England.

The Director, 1  
Tropical Products Institute,  
Gray's Inn Road,  
London WC1, England.

CANADA

National Defence Headquarters,  
Ottawa, Ontario, KLA OK2  
(a) Major N. A. Galbraith, 2  
(DCGE 3-4)  
(b) D Food S 1  
(c) DGOS Technical Library 1

SRI LANKA

Director, 1  
Food Research and Nutrition Council,  
C/- Food Commissioner,  
Union Place, Colombo, Sri Lanka

FEDERAL REPUBLIC OF GERMANY

Armed Services Food Chemist, 1  
89 Supply Depot, RAOC (BFPO 40)  
Viersen,  
Federal Republic of Germany

GHANA

Defence Adviser, 1  
Office of the High Commission for Ghana,  
13 Belgrave Square,  
London, SW1, England.

	<u>No. of Copies</u>
<u>OVERSEAS ESTABLISHMENTS (Con't)</u>	
<u>INDIA</u>	
Director General, Research and Development Organisation, Ministry of Defence, New Delhi, 11	2
The Director, Defence Food Research Laboratory, Jyothi Nagar, P.B. No. 45, Mysore - 1, India	2
<u>MALAYSIA</u>	
Director, Defence Research Centre, Ministry of Defence, Rifle Range Road, Kuala Lumpur, Malaysia	3
Director of Supplies and Transport, Ministry of Defence, C/- G.P.O. Kuala Lumpur	1
<u>NEW ZEALAND</u>	
Chief Scientist, Defence Science Organisation, DSIR, P.O. Box 8010, Wellington, New Zealand	1
<u>PAPUA NEW GUINEA</u>	
Headquarters, Australian Defence Cooperation Group, P.O. Box 2270, Kongdobu, Papua New Guinea	3
<u>PHILIPPINES</u>	
Science Research Supervisor, Food Research Division, Food and Nutrition Research Institute, Manila, Philippines	1
<u>UNITED STATES OF AMERICA</u>	
Chief, Food and Nutrition Section, NASA - Manned Spacecraft Center, Houston, Texas	1
Director, U.S. Army Natick Laboratories (Food Laboratory) Natick, Massachusetts, U.S.A.	2
Director, U.S. Army Medical Research and Nutrition Laboratory, Fitzsimons General Hospital, Denver, Colorado, U.S.A. 80240	2