

AD-A045 798

PETER BENT BRIGHAM HOSPITAL BOSTON MASS
INVESTIGATION OF IMMUNOREGULATORY ALPHAGLOBULIN (IRA) IN SHOCK --ETC(U)
OCT 77 J A MANNICK

F/G 6/1
DAMD17-76-C-6076
NL

UNCLASSIFIED

| OF |
AD
AO45798



END
DATE
FILMED

11 -77

DDC

(Unclassified)

12
P. S. AD _____

AD A 045798

Investigation of Immunoregulatory
Alphaglobulin (IRA) in Shock and Trauma

Annual Progress Report

John A. Mannick, M.D.

October 13, 1977

Supported by

U.S. ARMY MEDICAL RESEARCH AND DEVELOPMENT COMMAND
Washington, D.C. 20314

Contract No. DAMD 17-76-C-6076

Peter Bent Brigham Hospital
Boston, Massachusetts 02115

DDC
OCT 20 1977
C

Approved for public release; distribution unlimited

The findings in this report are not to be construed as an
official Department of the Army position unless so designated
by other authorized documents.

(Unclassified)

AD No. _____
DDC FILE COPY

SECURITY CLASSIFICATION OF THIS PAGE (When Data Entered)

REPORT DOCUMENTATION PAGE		READ INSTRUCTIONS BEFORE COMPLETING FORM
1. REPORT NUMBER 6	2. GOVT ACCESSION NO.	3. RECIPIENT'S CATALOG NUMBER 9
4. TITLE (and Subtitle) Investigation of Immunoregulatory Alpha-globulin (IRA) in Shock and Trauma		5. TYPE OF REPORT & PERIOD COVERED Annual Progress Report October 1976 - September 1977
7. AUTHOR(s) John A. Mannick, M. D.		8. CONTRACT OR GRANT NUMBER(s) DAMD 17-76-C-6076
9. PERFORMING ORGANIZATION NAME AND ADDRESS Peter Bent Brigham Hospital Boston, Massachusetts 02115		10. PROGRAM ELEMENT, PROJECT, TASK AREA & WORK UNIT NUMBERS 62772A 3S762772A814 00 014
11. CONTROLLING OFFICE NAME AND ADDRESS US Army Medical Research and Development Cmd. Washington, D. C. 20314		12. REPORT DATE 10/13/77
14. MONITORING AGENCY NAME & ADDRESS (if different from Controlling Office) 12 6p.		13. NUMBER OF PAGES 7 pages
		15. SECURITY CLASS. (of this report) Unclassified
16. DISTRIBUTION STATEMENT (of this Report) Approved for public release; distribution unlimited		15a. DECLASSIFICATION/DOWNGRADING SCHEDULE
17. DISTRIBUTION STATEMENT (of the abstract entered in Block 20, if different from Report)		
18. SUPPLEMENTARY NOTES		
19. KEY WORDS (Continue on reverse side if necessary and identify by block number) Trauma, Burns, Lymphocyte activation, Cellular immunity, Immunosuppression and Sepsis <i>was investigated.</i>		
20. ABSTRACT (Continue on reverse side if necessary and identify by block number) The major accomplishment of the past year of research supported by Contract number DAMD-17-76-C-6076 was the investigation of the nature of the immunosuppressive material present in the serum of patients sustaining major trauma or burns. We have fractionated the serum of such patients and have found the immunosuppressive activity resides in a low molecular weight polypeptide fraction which in turn has been fractionated by sephadex chromatography and high voltage electrophoresis. The active moiety appears <i>showed</i> (con't)		

D D C
APPROVED
OCT 20 1977
RECEIVED

B

20.

to be a basic peptide resolved by high voltage electrophoresis. During the past year we have simultaneously studied the response of lymphocytes recovered from the peripheral blood of patients sustaining major trauma and burns and have noted that the lymphocytes of many such patients are deficient in their ability to respond to mitogen stimulation in vitro and in their ability to form rosettes with sheep red blood cells. An attempt is currently being made to determine the correlation between hyporesponsiveness of lymphocytes from traumatized and burn patients and the suppressive activity of the serum from these individuals as tested against normal human lymphocytes.



REPORT DOCUMENTATION PAGE (mirrored bleed-through from the reverse side of the page)

RECEIVED
OCT 26 1971
D E C
RESEARCH
C

ACCESSION for	White Section <input checked="" type="checkbox"/>
	Buff Section <input type="checkbox"/>
NTIS	
DDC	
ANNOUNCED	
ABSTRACTICATION	
BY	
DISTRIBUTION/AVAILABILITY CODES	
	SPECIAL
A	

INSTRUCTIONS FOR PREPARATION OF REPORT DOCUMENTATION PAGE

RESPONSIBILITY. The controlling DoD office will be responsible for completion of the Report Documentation Page, DD Form 1473, in all technical reports prepared by or for DoD organizations.

CLASSIFICATION. Since this Report Documentation Page, DD Form 1473, is used in preparing announcements, bibliographies, and data banks, it should be unclassified if possible. If a classification is required, identify the classified items on the page by the appropriate symbol.

COMPLETION GUIDE

General. Make Blocks 1, 4, 5, 6, 7, 11, 13, 15, and 16 agree with the corresponding information on the report cover. Leave Blocks 2 and 3 blank.

Block 1. Report Number. Enter the unique alphanumeric report number shown on the cover.

Block 2. Government Accession No. Leave Blank. This space is for use by the Defense Documentation Center.

Block 3. Recipient's Catalog Number. Leave blank. This space is for the use of the report recipient to assist in future retrieval of the document.

Block 4. Title and Subtitle. Enter the title in all capital letters exactly as it appears on the publication. Titles should be unclassified whenever possible. Write out the English equivalent for Greek letters and mathematical symbols in the title (see "Abstracting Scientific and Technical Reports of Defense-sponsored RDT/E," AD-667 000). If the report has a subtitle, this subtitle should follow the main title, be separated by a comma or semicolon if appropriate, and be initially capitalized. If a publication has a title in a foreign language, translate the title into English and follow the English translation with the title in the original language. Make every effort to simplify the title before publication.

Block 5. Type of Report and Period Covered. Indicate here whether report is interim, final, etc., and, if applicable, inclusive dates of period covered, such as the life of a contract covered in a final contractor report.

Block 6. Performing Organization Report Number. Only numbers other than the official report number shown in Block 1, such as series numbers for in-house reports or a contractor/grantee number assigned by him, will be placed in this space. If no such numbers are used, leave this space blank.

Block 7. Author(s). Include corresponding information from the report cover. Give the name(s) of the author(s) in conventional order (for example, John R. Doe or, if author prefers, J. Robert Doe). In addition, list the affiliation of an author if it differs from that of the performing organization.

Block 8. Contract or Grant Number(s). For a contractor or grantee report, enter the complete contract or grant number(s) under which the work reported was accomplished. Leave blank in in-house reports.

Block 9. Performing Organization Name and Address. For in-house reports enter the name and address, including office symbol, of the performing activity. For contractor or grantee reports enter the name and address of the contractor or grantee who prepared the report and identify the appropriate corporate division, school, laboratory, etc., of the author. List city, state, and ZIP Code.

Block 10. Program Element, Project, Task Area, and Work Unit Numbers. Enter here the number code from the applicable Department of Defense form, such as the DD Form 1498, "Research and Technology Work Unit Summary" or the DD Form 1634, "Research and Development Planning Summary," which identifies the program element, project, task area, and work unit or equivalent under which the work was authorized.

Block 11. Controlling Office Name and Address. Enter the full, official name and address, including office symbol, of the controlling office. (Equates to funding/sponsoring agency. For definition see DoD Directive 5200.20, "Distribution Statements on Technical Documents.")

Block 12. Report Date. Enter here the day, month, and year or month and year as shown on the cover.

Block 13. Number of Pages. Enter the total number of pages.

Block 14. Monitoring Agency Name and Address (if different from Controlling Office). For use when the controlling or funding office does not directly administer a project, contract, or grant, but delegates the administrative responsibility to another organization.

Blocks 15 & 15a. Security Classification of the Report: Declassification/Downgrading Schedule of the Report. Enter in 15 the highest classification of the report. If appropriate, enter in 15a the declassification/downgrading schedule of the report, using the abbreviations for declassification/downgrading schedules listed in paragraph 4-207 of DoD 5200.1-R.

Block 16. Distribution Statement of the Report. Insert here the applicable distribution statement of the report from DoD Directive 5200.20, "Distribution Statements on Technical Documents."

Block 17. Distribution Statement (of the abstract entered in Block 20, if different from the distribution statement of the report). Insert here the applicable distribution statement of the abstract from DoD Directive 5200.20, "Distribution Statements on Technical Documents."

Block 18. Supplementary Notes. Enter information not included elsewhere but useful, such as: Prepared in cooperation with . . . Translation of (or by) . . . Presented at conference of . . . To be published in . . .

Block 19. Key Words. Select terms or short phrases that identify the principal subjects covered in the report, and are sufficiently specific and precise to be used as index entries for cataloging, conforming to standard terminology. The DoD "Thesaurus of Engineering and Scientific Terms" (TEST), AD-672 000, can be helpful.

Block 20. Abstract. The abstract should be a brief (not to exceed 200 words) factual summary of the most significant information contained in the report. If possible, the abstract of a classified report should be unclassified and the abstract to an unclassified report should consist of publicly-releasable information. If the report contains a significant bibliography or literature survey, mention it here. For information on preparing abstracts see "Abstracting Scientific and Technical Reports of Defense-Sponsored RDT&E," AD-667 000.

PROGRESS REPORT - ANNUAL

For the past three years with the support of this contract we have recovered an immunosuppressive polypeptide in trace amounts from pooled normal human serum. This peptide fraction, which we have called immunoregulatory alphaglobulin (IRA), is probably carried non-covalently bound to an alphaglobulin carrier or carriers at normal pH. It is released from its loosely bound state under conditions of high ionic strength and acidic pH. The IRA peptide has a molecular weight of 1000-2000 and chemically appears to be entirely a polypeptide by the quantitative Biuret test. It contains no nucleic acids, neutral sugars, sialic acid or lipids. It also contains no cortisol, prostaglandins E_1 or E_2 or ribonuclease activity.

We have shown that IRA inhibits a wide variety of T-cell mediated immune responses including the rejection of skin allografts in mice, the rejection of renal allotransplants in rats, immunity to syngeneic tumors in mice, the production of MIF by specifically sensitized guinea pig T-cells, the proliferative response of human and animal lymphocytes in vitro to T-cell mitogens, such as phytohemagglutinin (PHA) and to specific antigens to which the lymphocyte donor is known to be sensitized. We have shown that IRA is non-toxic to lymphocytes or experimental animals. It appears to act by preventing the recognition of antigen by potentially responsive T-lymphocytes. It is ineffective if it is administered after the antigen to be studied. Lymphocytes exposed to IRA will respond normally in tissue culture to antigenic and mitogenic stimuli if they are thoroughly washed and then recultured in medium lacking IRA. IRA has no effect on B-cell immune responses, however, it markedly inhibits antibody formation to antigens which require T-helper function, such as the plaque-forming cell response of mice to sheep erythrocytes (SRBC).

During the past two years we have studied a group of 117 patients who have been subjected to major trauma, including major surgery, or who have sustained burns. We have found that varying percentages of this group of patients have immunosuppressive serum, defined as serum which will inhibit by 50% or more the in vitro stimulation of normal human peripheral blood lymphocytes by phytohemagglutinin (PHA), when compared to normal human AB serum and/or autologous serum. The percentage of patients having immunosuppressive serum has been found to vary with the severity of the trauma and with the number of complications sustained by the patient. Patients who have septic complications are particularly likely to have suppressive serum. The immunosuppressive activity in trauma patients' serum does not appear to be caused by anesthetic agents since patients undergoing general anesthesia for minor surgery do not develop suppressive serum.

We have fractionated the serum of many of these individuals by DEAE cellulose chromatography and have found that the majority of the immunosuppressive activity is recovered in Peak I while the immunosuppressive activity in normal patients' serum is recovered in later alphasglobulin rich peaks. We have also found that the immunosuppressive activity contained in Peak I from trauma patients can be recovered after acidification and diafiltration, as a peptide fraction of less than 2,000 molecular weight, which is very highly suppressive of T-cell function both in vivo and in vitro. It appears, therefore, that patients who have been recently traumatized have high levels of a circulating immunosuppressive peptide fraction which appears to be similar to IRA peptide. The peptide obtained has been investigated chemically to determine its true peptide content by the quantitative Biuret test and has been found to be entirely composed of polypeptide as far as can be determined by this assay. The presence of nucleic acids and carbohydrates have been ruled out by standard chemical techniques and no cortisol is present as determined by the competitive protein binding assay. Similarly, the peptide contains no prostaglandins E₁ and E₂ as determined by radioimmunoassay.

More recently we have attempted to fractionate the immunosuppressive peptide obtained from trauma serum by G25 sephadex gel filtration and finally by preparative high voltage electrophoresis. In initial studies it appears that a good share of the immunosuppressive activity in this peptide fraction is contained in the peptides at the basic end of the electrophoresis pattern. The active peptide in normal serum and in serum from cancer patients also appears to be located in the same area by preparative high voltage electrophoresis. We have recently been exploring a technique to fractionate the serum from trauma patients and cancer patients by high pressure liquid chromatography in an attempt to purify the suppressive peptide moiety. Attempts at amino acid analysis of the basic polypeptide obtained by high voltage electrophoresis have been impaired by the fact that the peptide must be eluted from paper and, therefore, the final product is contaminated by amino acids from the paper. The advantage of high pressure liquid chromatography, if the peptide can be isolated by this method, is that the product will be free of salt or amino acid contamination. The trauma serum peptide when isolated will be subjected to amino acid analysis and ultimately to amino acid sequencing. Its amino acid composition will then be compared with that of similar peptides obtained from cancer serum and normal serum.

During the past year we have continued a study of the lymphocytes of trauma patients with regard to their response to PHA stimulation in vitro and their ability to form rosettes with sheep red blood cells (SRBC). During the past 6 months 20 patients have been studied. As anticipated the lymphocytes of patients who have suffered major burns or the most severe trauma are suppressed with respect to their ability

Page Three

to respond to PHA stimulation and with regard to their ability to form rosettes with SRBC (E-rosettes) as compared with control lymphocytes from normal individuals, even when incubated in the same normal AB reference serum. In some trauma patients whose lymphocyte responsiveness in vitro is impaired, the response has returned to or toward normal after washing the lymphocytes 6 times as opposed to once in vitro in tissue culture medium prior to culture with PHA or exposure to SRBC. There is also a correlation between the hyporesponsiveness of lymphocytes from traumatized and burn patients and the suppressive activity of the serum from these individuals as tested against normal lymphocytes. However, this correlation is only rough at present and more patients must be studied. It appears that the more suppressive the serum the more likely it is that the patient's lymphocytes are hyporesponsive in normal serum after one washing in vitro. Thus it appears that one reason for the hyporesponsiveness of the lymphocytes of trauma patients as reported by a number of workers is likely to be lymphocyte surface coating with a substance which can be removed by multiple washings in tissue culture medium. We are currently studying the medium in which the lymphocytes have been washed in an attempt to obtain the suppressive material from this source.

DISTRIBUTION LIST

4 copies

HQDA (SGRD-AJ)
Washington DC 20314

12 copies

Defense Documentation Center (DDC)
ATTN: DDC-TCA
Cameron Station
Alexandria, Virginia 22314

1 copy

Superintendent
Academy of Health Sciences, US Army
ATTN: AHS-COM
Fort Sam Houston, Texas 78234

1 copy

Dean
School of Medicine
Uniformed Services University of the
Health Sciences
Office of the Secretary of Defense
6917 Arlington Road
Bethesda, Maryland 20014