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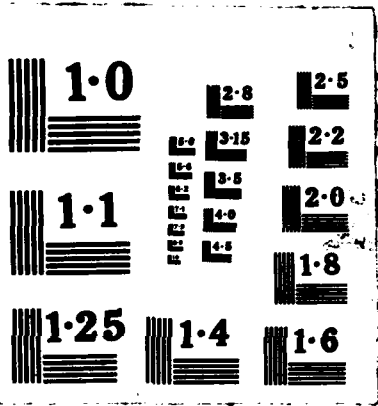
MECHANISMS OF TRANSMITTER RELEASE IN HIPPOCAMPUS
UNIVERSITY RESEARCH INSTRUMENTATION PROGRAM(U) BAYLOR
COLL OF MEDICINE HOUSTON TX D JOHNSTON 10 SEP 87
AFOSR-TR-87-1431 AFOSR-86-0214 F/G 6/4

1/1

UNCLASSIFIED

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REPORT DOCUMENTATION PAGE

1a. REPORT SECURITY
Unclassified
2a. SECURITY CLASSIF
2b. DECLASSIFICATION

AD-A187 454

1b. RESTRICTIVE MARKINGS

3. DISTRIBUTION/AVAILABILITY OF REPORT
Approved for public release; distribution unlimited.

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AFOSR-TR-87-1431

4. PERFORMING ORGANIZATION REPORT NUMBER(S)

5. MONITORING ORGANIZATION REPORT NUMBER(S)

6a. NAME OF PERFORMING ORGANIZATION
Baylor College of Medicine

6b. OFFICE SYMBOL
(if applicable)

7a. NAME OF MONITORING ORGANIZATION
AFOSR/NL

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Building 410
Bolling AFB, DC 20332-6448

8a. NAME OF FUNDING / SPONSORING ORGANIZATION
AFOSR

8b. OFFICE SYMBOL
(if applicable)
NL

9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER
AFOSR 86-0214

8c. ADDRESS (City, State, and ZIP Code)
Building 410
Bolling AFB, DC 20332

10 SOURCE OF FUNDING NUMBERS

PROGRAM ELEMENT NO.	PROJECT NO.	TASK NO.	WORK UNIT ACCESSION NO.
61102F	2917	A4	

11. TITLE (Include Security Classification)

Mechanisms of Transmitter Release in the Hippocampus (Unclassified)

12. PERSONAL AUTHOR(S)
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13a. TYPE OF REPORT
Final Report

13b. TIME COVERED
FROM 7/1/86 TO 6/30/87

14. DATE OF REPORT (Year, Month, Day)
87/09/10

15. PAGE COUNT
1

16. SUPPLEMENTARY NOTATION

17. COSATI CODES		
FIELD	GROUP	SUB-GROUP

18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)
mossy fiber synaptic terminals, hippocampus, LTP, subcellular fractionation and patch clamping facility

19. ABSTRACT (Continue on reverse if necessary and identify by block number)

This grant was made under the DoD University Research Instrumentation Program. Our request was for equipment to implement a subcellular fractionation and patch clamping facility. The equipment requested was for a Beckman L7-55 ultracentrifuge with a SW-41 rotor kit, a Beckman J2-21 centrifuge with JS-13.1 and JA-20 rotors, a Masscomp 5400 computer, a Zeiss IM microscope, and a Newport vibration isolation table. All items of equipment have been ordered, received, and have been in use for several months. The research for which this equipment is being used is that associated with AFOSR-85-0178. We have been isolating an enriched fraction of mossy fiber synaptic terminals from hippocampus of rat and investigating mechanisms for release of glutamate.

20. DISTRIBUTION/AVAILABILITY OF ABSTRACT
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21. ABSTRACT SECURITY CLASSIFICATION
Unclassified

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Mechanisms of Transmitter Release in Hippocampus
 University Research Instrumentation Program
 AFOSR-86-0124
 Final Report

→ This grant was for the purchase of equipment to establish a subcellular fractionation and patch clamping facility at Baylor College of Medicine. This equipment will be utilized for research projects associated with AFOSR Grant 85-0178, "Amine Neurotransmitter Regulation of Long-Term Synaptic Plasticity in Hippocampus." The equipment requested was: Beckman L7-55 ultracentrifuge with SW-41 rotor kit, Beckman J2-21 centrifuge with JS-13.1 and JA-20 rotors, Masscomp 5400 computer, Zeiss IM microscope, and a Newport vibration isolation table. All of the above items of equipment have been purchased, received, and have been in use for the last six months.

→ The experiments involve the isolation of an enriched fraction of mossy fiber synaptic terminals from adult rats. We have been investigating mechanisms of transmitter release, using biochemical and electrophysiological techniques. We have used the centrifuges successfully to develop this preparation of enriched mossy fiber synaptosomes. The computer, microscope, and isolation table are in use as a patch clamping facility to study the electrophysiological properties of these terminals.

We have successfully measured the potassium stimulated and calcium dependent release of endogenous glutamate from these terminals. We have found that several phorbol esters are able to potentiate this release of glutamate, and we are in the process of investigating the mechanisms underlying this enhanced release.

Our patch clamping has met with only limited success thus far. Although we have shown that the technique can be successfully applied to these small terminals, we have yet to make recordings of single calcium channels. The channels recorded thus far appear to be nonselective cation channels. We are currently in the process of altering our procedures and are hopeful that this aspect of the project will meet with more success in the very near future.

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