

AD-A203 478

SEC

REPORT DOCUMENTATION PAGE			Form Approved OMB No. 0704-0188	
1a. REPORT SECURITY CLASSIFICATION (U)		1b. RESTRICTIVE MARKINGS NA		
2a. SECURITY CLASSIFICATION AUTHORITY NA		3. DISTRIBUTION / AVAILABILITY OF REPORT Distribution Unlimited		
2b. DECLASSIFICATION / DOWNGRADING SCHEDULE NA		5. MONITORING ORGANIZATION REPORT NUMBER(S) NA		
4. PERFORMING ORGANIZATION REPORT NUMBER(S) University of Pennsylvania		7a. NAME OF MONITORING ORGANIZATION Office of Naval Research		
6a. NAME OF PERFORMING ORGANIZATION University of Pennsylvania	6b. OFFICE SYMBOL (If applicable) NA	7b. ADDRESS (City, State, and ZIP Code) 800 N. Quincy Street Arlington VA 22217-5000		
6c. ADDRESS (City, State, and ZIP Code) Department of Pharmacology University of Pennsylvania School of Medicine 3841 Locust Walk, Philadelphia, PA 19104		9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER N00014-88-K-0048		
8a. NAME OF FUNDING / SPONSORING ORGANIZATION Office of Naval Research	8b. OFFICE SYMBOL (If applicable) ONR	10. SOURCE OF FUNDING NUMBERS		
8c. ADDRESS (City, State, and ZIP Code) 800 N. Quincy Street Arlington, VA 22217-5000		PROGRAM ELEMENT NO. 61153N	PROJECT NO. RR04108	TASK NO. 4414807
11. TITLE (Include Security Classification) Inhibition of ACTH release by peptide hormones: Molecular mechanisms and possible role as anti-stress				
12. PERSONAL AUTHOR(S) Terry Reisine				
13a. TYPE OF REPORT Annual	13b. TIME COVERED FROM 2/28/88 TO 12/1/88	14. DATE OF REPORT (Year, Month, Day) 1988; Dec. 27	15. PAGE COUNT	
16. SUPPLEMENTARY NOTATION				
17. COSATI CODES			18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)	
FIELD	GROUP	SUB-GROUP	Stress Physiology, Corticotrophs, Anterior pituitary	
08			Somatostatin receptor, Adrenocorticotropin, Cholecystic inhibitors, molecular biology, endocrinology	
19. ABSTRACT (Continue on reverse if necessary and identify by block number) The major objective of this proposal was to investigate whether somatostatin (SRIF) may be a non-steroid inhibitor of ACTH release from the anterior pituitary. The goals of the proposal were to; 1) investigate the structure of the SRIF receptor; 2) examine the effects of glucocorticoids on the expression of SRIF receptors in the pituitary; and 3) determine the physical differences between functionally distinct subtypes of SRIF receptors in the pituitary. In the past year, we showed that the pituitary and brain SRIF receptor is a glycoprotein of 55 to 60 kilodaltons. Furthermore, we were able to purify the SRIF receptor from brain by affinity chromatography. Presently we are attempting to sequence the receptor in order to attempt to clone the gene coding for this protein. In addition, we showed that glucocorticoids downregulate SRIF receptors in the anterior pituitary cell line AtT-20, which consists of a homogeneous population of ACTH secreting cells. Interestingly, some of the biological actions of SRIF on AtT-20 cells (inhibition of adenylate cyclase activity) are not affected by glucocorticoid treatment. Finally, we have observed that minor structural differences in the SRIF receptor appear to be responsible for the vastly different functional properties of the SRIF receptor subtypes in the pituitary. Attempts are under way to establish the basis of these minor structural differences. Reisine et al.				
20. DISTRIBUTION / AVAILABILITY OF ABSTRACT ☒ UNCLASSIFIED/UNLIMITED ☐ SAME AS RPT. ☐ DTIC USERS			21. ABSTRACT SECURITY CLASSIFICATION (U)	
22a. NAME OF RESPONSIBLE INDIVIDUAL Dr. J.A. Maide			22b. TELEPHONE (Include Area Code) 202-696-4055	22c. OFFICE SYMBOL ONR

DD Form 1473, JUN 86

Previous editions are obsolete.

SECURITY CLASSIFICATION OF THIS PAGE

S/N 0102-LF-014-6603

**ANNUAL REPORT ON CONTRACT ONR N00014-88-K-0048
R and T Code 4414807**

PRINCIPAL INVESTIGATOR Terry Reisine

CONTRACT TITLE: Inhibition of ACTH release by peptide hormones: Molecular mechanisms and possible role as anti-stress factors.

Contract Period: Nov. 1, 1987 to Oct, 30, 1988.

INTRODUCTION AND RESEARCH OBJECTIVES: The major objective of the research proposal was the identification of non-steroidal factors which inhibit ACTH release and may be useful in the treatment of stress. Previously, it was reported that the hypothalamic peptide somatostatin (SRIF) could inhibit CRF stimulated ACTH release from a tumor cell line of the anterior pituitary, AtT-20, which consists of a homogeneous population of ACTH secreting cells. While administration of SRIF to non-stressed, humans failed to alter plasma ACTH or cortisol levels, patients with adrenal insufficiency did respond to SRIF with a lowering of ACTH release. This latter finding, may indicate that SRIF has some role in the suppression of ACTH secretion in humans but that under normal conditions, glucocorticoids downregulate or diminish the effectiveness of SRIF in regulating ACTH secretion. In fact, several studies have suggested that glucocorticoids downregulate SRIF receptors in the anterior pituitary. Furthermore, Lamberts et al (Endocrinol. 118:2188, 1986), showed that SRIF could inhibit CRF stimulated ACTH release from anterior pituitary cells in culture grown in the presence of glucocorticoid antagonists such as RU 486. Thus, a major aspect of the proposal was to determine the mechanisms by which SRIF may regulate ACTH release and whether this effect was in fact tonically suppressed by glucocorticoids.

Three aspects of the proposal which have progressed since the start of funding are; 1) the structural analysis of the SRIF receptor, 2) the effect of glucocorticoid treatment on SRIF receptor expression; and 3) the structural analysis of SRIF receptor subtypes.

PROGRESS REPORT:

1. Physical properties of SRIF receptors: Concerning the structural analysis of the SRIF receptor, we have employed several approaches to investigate the physical properties of SRIF receptors in AtT-20 cells, rat anterior pituitary and brain. Using photocrosslinking techniques, SRIF receptors can be covalently labeled with the stable SRIF analogue [125I] CGP 23996 (Thermos and Reisine, Molecular Pharmacology, 33:370-377; Thermos et al. in press). [125I] CGP 23996 binds to SRIF receptors in membranes of AtT-20 cells, anterior pituitary and brain in a saturable and specific manner (Mahy et al., in press). With the photoactivated crosslinking agent HSAB, it was possible to covalently link [125I] CGP 23996 to the SRIF receptor in AtT-20 and GH3 cells as well as rat anterior pituitary and brain (Thermos and Reisine, Mol. Pharmacol. 33:370; Thermos et al. in press). The binding could be blocked completely by SRIF but not by peptides which do not directly interact with the SRIF receptor. The size of the SRIF receptor labeled in these tissues is between 55 to 60 kilodaltons. The size and charge of the labeled proteins as assessed by two-dimensional polyacrylamide gel electrophoresis (PAGE) in all of these tissues is similar. The SRIF receptor does not appear to contain disulfide bridges since reducing agents do not alter its migration in SDS-gels. The receptor

appears to be a glycoprotein since it is able to bind selectively to wheat germ agglutinin. The sugar moiety of the receptor may be responsible for heterogeneities in the structure and function of the receptor in various tissues.

To further examine the physical properties of the SRIF receptor, we solubilized and purified it from brain using affinity chromatography (He et al., in press). The purified receptor is 60 kilodaltons in size and can be selectively labeled by [125I] CGP 23996. The purified protein is essentially homogeneous since iodination of the eluate from the SRIF affinity column with Bolton-Hunter reagent revealed that 95% of the radioactivity is present in the 60 kilodalton protein. Presently, we are attempting to obtain partial sequence information of the purified SRIF receptor in order to develop procedures for cloning the gene coding for this protein.

2. Effect of glucocorticoids on SRIF receptor expression: Concerning the effects of glucocorticoids on SRIF receptors, treatment of AtT-20 cells with dexamethasone decreased [125I] CGP 23996 binding to AtT-20 cell membranes by over 50%. The effect was dependent on the concentration of dexamethasone used with significant decreases occurring with 1 nM of the steroid and maximal effects with 100 nM. The decrease in binding was also dependent on the time of dexamethasone treatment with significant decreases seen after only 4 hr of pretreatment. The steroid antagonist RU 486 blocked the effects of dexamethasone on AtT-20 cells. Treatment of AtT-20 cells with high concentrations of other steroids such as testosterone and estrogen did not alter [125I] CGP 23996 binding to the AtT-20 cells SRIF receptor. Interestingly, glucocorticoid treatment did not affect the ability of SRIF to inhibit adenylate cyclase activity in AtT-20 cell membranes. This finding may indicate that there are either "spare" SRIF receptors or subpopulations of receptors, some of which are not coupled to adenylate cyclase. Future studies will determine whether dexamethasone affects SRIF receptors in rat anterior pituitary cells in culture and alters other the functional characteristics of SRIF receptors such as their coupling to ACTH release.

3. Structural analysis of SRIF receptor subtypes: SRIF receptors in AtT-20 cells and the growth hormone secreting tumor cell line, GH3, are functionally distinct. The receptors in AtT-20 cells have higher affinity for somatostatin-28 than somatostatin-14 and desensitize following agonist treatment (Thermos and Reisine, Mol. Pharmacol. 33:370; Mahy et al., 1988). The SRIF receptors in GH3 cells have higher affinity for somatostatin-14 than somatostatin-28 and do not desensitize (Thermos and Reisine, Mol Pharmacol. 33:370). Using photocrosslinking techniques, we covalently labeled the SRIF receptors in both cell lines with [125I] CGP 23996. The size, charge and peptide maps of the receptors, as assessed by SDS-PAGE were identical. However, analysis of the peptide maps by two-dimensional PAGE revealed small variations in the properties of the receptor subtypes. Future studies will investigate whether these small structural variations in the SRIF receptor subtypes are due to variations in sugar composition or amino acid sequence.

EXPENDITURES

Supplies: expenditures normal

Equipment: ONR funds were used to purchase a gamma counter which was requested in the original proposal.

Travel: Travel funds were used by the PI to attend the Soc. for Neuroscience Meeting in Toronto.

For

I

on

on/

ty Code

and/or

Special

Dist

A-1

Personnel: Henry Wang: Research Assistant (100% support).

Women or minorities- 0; Non-citizens - 1.

Publications

Abstracts:

Thermos, K., He, H.T., Margolis, N. and Reisine, T.: Biochemical properties of brain Somatostatin receptors. Soc. Neurosci. Abst 14:14, 1988.

Manuscripts:

Mahy, N., Woolkalis, M., Manning, D. and Reisine, T.: Characterization of somatostatin desensitization in the pituitary tumor cell line AtT-20. J. Pharmacol Expt. Therap. 247:390-396, 1988.

Reisine, T.: Phorbol esters and corticotropin releasing factor stimulate calcium influx in the anterior pituitary tumor cell line AtT-20 through different intracellular sites of action. J. Pharmacol. Expt. Therap. (in press).

He, H.T., Johnson, K., Thermos, K. and Reisine, T.: Purification of a putative brain somatostatin receptor. Proc. Natl. Acad. Sci. (in press).

Thermos, K., He, H.T., Wang, H., Margolis, N. and Reisine, T.: Biochemical properties of brain somatostatin receptors. Neuroscience (in press).

2ND YEAR WORK PLAN: The objectives of year 2 of funding are 1) to obtain partial sequence information of the purified SRIF receptor so as to begin cloning of the gene coding for this protein; 2) submit the studies on the glucocorticoid regulation of the SRIF receptor for publication and to determine whether such regulation occurs in normal anterior pituitary corticotrophs; and 3) to determine what are the structural differences in SRIF receptor subtypes in AtT-20 and GH3 cells.

DISTRIBUTION LIST

Stress Neurochemistry Program

Annual, Final and Technical Reports (one copy each)

INVESTIGATORS

Dr. H. Elliott Albers
Lab. Neuroendocrin. & Behavior
Depts. of Biology & Psychology
Georgia State University
Atlanta, GA 30303

Dr. Gwen V. Childs
Dept. of Anatomy & Neuroscience
Univ. of Texas Medical Branch
Galveston, TX 77550

Dr. Carl E. Creutz
Dept. of Pharmacology
University of Virginia
Charlottesville, VA 22908

Dr. Mary F. Dallman
Dept. of Physiology
University of California, Box 0444
San Francisco, CA 94143-0444

Dr. Caleb E. Finch
Dept. of Neurobiology
Univ. of Southern California
Los Angeles, CA 90089-0191

Dr. Thackery S. Gray
Department of Anatomy
Loyola University Medical Center
216 South First Avenue
Maywood, IL 60153

Dr. Richard F. Ochillo
College of Pharmacy
Xavier Univ. of Louisiana
7325 Palmetto Street
New Orleans, LA 70125

Dr. Terry Reisine
Dept. of Pharmacology
Univ. of Pennsylvania
School of Medicine
36th and Hamilton Walk
Philadelphia, PA 19104

Dr. C. Frank Starmer
P.O. Box 3181
Duke Univ. Medical Center
Durham, NC 27710

Dr. Kent E. Vrana
Dept. of Biochemistry
West Virginia School
of Medicine
Morgantown, WV 26506

Stress Neurochemistry

Annual, Final and Technical Reports (one copy each except as noted)

ADMINISTRATORS

Scientific Officer, Physiology
Code 1141SB
Office of Naval Research
800 N. Quincy Street
Arlington, VA 22217-5000

Program Manager, Code 1213
Human Factors Biosciences
Division
Office of Naval Research
800 N. Quincy Street
Arlington, VA 22217-5000

Administrator (2 copies) (Enclose DTIC Form 50)
Defense Technical Information Center
Building 5, Cameron Station
Alexandria, VA 22314

Program Manager, Code 223
Support Technology
Directorate
Office of Naval Technology
800 N. Quincy Street
Arlington, VA 22217-5000

Administrative Contracting Officer
ONR Resident Representative
(address varies - obtain from business office)

Annual and Final Reports Only (one copy each)

DoD ACTIVITIES

Commanding Officer
Naval Medical Center
Washington, DC 20372

Commanding Officer, Code 404
Naval Medical Research & Development Command
National Naval Medical Center
Bethesda, MD 20814

Commander
Chemical and Biological Sciences Division
Army Research Office, P.O. Box 12211
Research Triangle Park, NC 27709

Directorate of Life Sciences
Air Force Office of Scientific Research
Bolling Air Force Base
Washington, DC 20332

Final and Technical Reports Only

Director, Naval Research Laboratory (6 copies)
Attn: Technical Information Division, Code 2627
Washington, DC 20375