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**FUNDAMENTAL STUDIES IN THE MOLECULAR BASIS
OF LASER INDUCED RETINAL DAMAGE**

Annual Report

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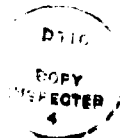
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A new unique method of investigating the effect of femtosecond pulses using nanosecond lasers that are common in most laser laboratories has been developed. This new method does not create femtosecond pulses by building a complex femtosecond laser but by modulating the energy spread in conventional lasers to generate pulses with ultrashort coherence times. <i>Keywords: Laser damage, eye damage; (KT)</i>					
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FOREWORD

In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Animal Resources, National Research Council (NIH Publication No. 86-23, Revised 1985).

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1. Introduction

The initial step in the process of visual excitation is the absorption of light in the chromophore. In all visual pigments, the chromophore is a derivative of retinal (Vitamin A aldehyde). Thus, a detailed study of the excited state properties of this retinylidene chromophore and related molecules is a crucial step in understanding the visual process.

Several spectroscopic techniques¹ have been applied to investigate the photophysical and photochemical properties of the retinylidene chromophore. But few nonlinear optical techniques have been used. Such methods can yield interesting information as has been demonstrated in other system.^{2,3} Second harmonic generation (SHG) is the lowest order nonlinear optical process, in which the second order polarizability of a material is responsible for the generation of light at the second harmonic frequency. Due to symmetry consideration, the SHG is forbidden in an isotropic medium in the electric dipole approximation, but allowed at the surface where the inversion symmetry is broken. It was recognized by Y. R. Shen et al.⁴ that this attribute can be used to probe adsorbed monolayers at surfaces with high spatial and temporal resolution. Of great importance is the fact that this technique has

been demonstrated to have submonolayer sensitivity^{5,6} and recently Th. Rasing et al.⁷ have proposed a simple and direct method to measure the second-order molecular polarizabilities of some organic molecules by using the surface second harmonic generation technique directly on a Langmuir-Blodgett trough.

Monolayers based on Langmuir-Blodgett troughs have attracted renewed interest during the last few years⁸ because such troughs allow the organization of appropriate molecules in a planned way so that the realization of artificial devices acting at the molecular scale becomes imaginable. The properties of biological membranes can readily be simulated by Langmuir-Blodgett (LB) monolayers. Second harmonic generation has the potential of being used as a direct, *in situ* surface probe of such monolayers. Using this probe, detailed information can be obtained on the physical, chemical and biological properties of these monolayers.

In this paper we apply SHG to compare the second harmonic properties of monolayers of retinal, retinylidene *n*-butylamine Schiff base (NRB) and protonated NRB (PNRB). The results have yielded new insight into the structure and dipolar properties of these molecules.

2. Experimental Procedures

A Q-switched frequency doubled Nd:YAG laser with a 10Hz repetition rate and 10nsec pulsewidth was used. The Nd:YAG laser was adjusted to give a green 532nm beam of less than 20mW which was focused onto a 3mm

diameter spot on the sample surface after passing through a half-wave plate, a Glan-Thompson laser prism polarizer and a KG-5 color glass filter. The reflected second harmonic signal at a wavelength 266nm was passed through a UG-5 color glass filter which blocked the fundamental at 532nm. The UG-5 filter was followed by a UV Glan-Taylor polarizer and a Schoeffel 0.2m double monochromator. The SH signal was detected by a cooled RCA C31034 photomultiplier and averaged by a boxcar integrator.

All-trans retinal purchased from Sigma was dissolved in n-hexane without further purification. High pressure liquid chromatography indicated that the all-trans retinal was greater than 95% pure. All-trans retinylidene butylamine Schiff base (NRB) was prepared as described before⁹. The molar concentrations of retinal, unprotonated and protonated NRB in methanol solutions were determined by the published extinction coefficients $\epsilon_{380} = 4.2884 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$ ^{10a}, $\epsilon_{365} = 5.23 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$ and $\epsilon_{445} = 4.98 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$ ^{10b} respectively. The molecules were spread on the surface of deionized distilled pure water subphase at pH 6.4.

The Langmuir trough was made out of teflon. A movable barrier controlled the surface density of molecules and a #40 filter paper with dimension 0.7cm x 1.0cm was attached to a Gould UC-3 force transducer to measure the surface pressure. During detection of the second harmonic signal, the movable barrier was controlled by a stepping motor such that the surface pressure of the film was kept at constant value.

Molecular orientation was determined by using the Heinz's nulling technique¹¹. The incident pump beam was arranged to satisfy the following condition:

$$[e_{\parallel}(\omega)/e_{\perp}(\omega)]^2 = 2 \quad 1$$

where $e(\omega)$ is the product of the polarization vector and the Fresnel factor for the pump field in the monolayer¹². The subscripts denote the components of $e(\omega)$ on or perpendicular to the layer plane. This condition can be met by adjusting the pump beam to have an angle of incidence of 45° and a polarization of 35.3° away from the incident plane. The molecular orientation was determined by the second harmonic extinction direction, as indicated by the UV polarizer.

Our detection sensitivity was calibrated against the second harmonic intensity from an x-cut quartz plate, observed under the same experimental conditions. The quartz plate was rotated 28.75° about the y-axis of the crystal to obtain one of the maxima of the Maker's fringes. The polarizations of the fundamental and second harmonic beams were chosen to be p-polarized relative to the plane of incidence. In this case, the two beams are the extraordinary waves of the quartz crystal. Using the known nonlinear second order susceptibility $d_{11} = 2.4 \times 10^{-9}$ esu¹³ and the index of refraction of quartz at 532nm and 266nm, we obtained a ratio of¹⁴

$$I_{p \rightarrow p}(2\omega)/[I_p(\omega)]^2 = 3.96 \times 10^{-26} \text{ cm}^2 \text{ erg}^{-1} \quad 2$$

The potential energy barrier between the water subphase and the NRB monolayer is quite small. This is particularly true for the NRB on an acid (pH 2.8) water subphase. We have observed that the unprotonated NRB molecules leak through the movable barrier on the acid water subphase during the compression stage. In addition, the proton on the Schiff base nitrogen of protonated NRB was found to be able to detach from the nitrogen. In order to avoid these problems, the second harmonic intensities from unprotonated and protonated NRB monolayers were compared by using spin-coated monolayers on glass substrates.¹⁵ The monolayers were carefully prepared to make sure that they have the same surface density of molecules. Several steps were taken to prevent the molecules on the glass substrates from isomerization. First, the average power of the pump laser was limited to below 15mW when the spin-coated samples were measured. Second, the data collection time was reduced to 20sec and third, the samples were mounted on a spinning DC motor to minimize the exposure time of molecules to the pumping laser beam.

3. Results and Discussion

Correlation of Mechanical and Structural Properties of Retinal and Schiff-Base Monolayers

The molecular structures of all-trans retinal and its Schiff bases are shown in Figure 1. The initial characterization of the all-trans retinal unprotonated retinylidene n-butylamine Schiff base (NRB) Langmuir-Blodgett films were made by measuring surface pressure/surface area (π -A) isotherms. This allowed the mechanical properties of the films to be assessed. These measurements are shown in Figure 2, where the curve (a) and (b) are for retinal and unprotonated NRB respectively. The limiting areas $A_{\pi \rightarrow 0}$ obtained from extrapolation of the linear part to zero pressure,¹⁶ were found to be $58\text{\AA}^2$ for retinal and $69\text{\AA}^2$ for unprotonated NRB. Based on the linear dimensions estimated from space-filling models of retinal, $18\text{\AA}$, these limiting areas suggest the molecular long axes must tilt away from the water subphase surface. This observation is consistent with the results of the molecular orientation measurement, which are shown in Figure 3. The only difference between the chemical structures of retinal and unprotonated NRB is that the oxygen atom in retinal is replaced by a butylamine group (see Figure 1). The additional butyl group will require the NRB molecule to occupy a larger surface area than retinal does under the same surface pressure. However this does not have any appreciable effect on the molecular orientation (see Figure 3). In this measurement the molecular directions of retinal and NRB are studied as a function of

compressing the surface area and increasing the surface pressure. As is seen in this Figure, for each of these molecules at low surface pressure a molecule direction of $\approx 70^\circ$ relative to the surface normal is approached whereas at high surface pressure this angle reaches $\approx 50^\circ$.

The cross section of the β -ionone ring on a plane perpendicular to the molecular long axis has dimensions of $5.2\text{\AA}$ by $7.7\text{\AA}$. If most of the steric interaction among molecules comes from the β -ionone ring, it would be expected that the LB films become closely packed at a surface area of $40\text{\AA}^2$. Indeed from Figure 2, we observe that the isotherms of these two species level-off below $32\text{\AA}^2$. The slight difference of $8\text{\AA}^2$ can be attributed to the dynamic property of the molecules on the water surface. In order to examine the structure of the film, we have measured the second harmonic intensity under different surface pressures. The results are shown in Figure 4. It was observed that the SH p-polarized intensity $I_{s \rightarrow p}$ increase with the surface pressure in a nonlinear fashion. This is understandable by considering that the SH intensity is proportional to the absolute square of the second order molecular hyperpolarizability, β , by a coordinate transformation that depends on the molecular orientation:⁴

$$(\chi^{(2)})_{ijk} = N_s \langle G_{ijk}^{\xi\eta\zeta}(\theta, \phi, \psi) \rangle \beta \quad 3$$

Here, $\langle G_{ijk}^{\xi\eta\zeta} \rangle$ is the geometric factor specified by the coordinate transformation from the molecular coordinates (ξ, η, ζ) to the lab coordinates (i, j, k) , with the Euler angles (θ, ϕ, ψ) describing the molecular orien-

tation and the angular brackets indicating an average over all the molecules. From Figure 3, we found $\langle G_{ijk}^{\xi\eta\zeta} \rangle$ can depend on the surface pressure by way of the molecular orientation. Furthermore, the surface density N_s is not a linear function of the surface pressure as indicated in Figure 2: The SH intensity $I_{p \rightarrow p}$ on the other hand monotonically decreases with the surface pressure. It drops to zero at 14 dynes/cm. The second harmonic p-polarized electric field strength $E_{p \rightarrow p}(2\omega)$ is obtained in terms of the incident p-polarized field:¹⁷

$$E_{p \rightarrow p}(2\omega) \propto \langle \beta \rangle_{zxx} (\epsilon_w - \sin^2 \alpha) \left(1 - \frac{2\epsilon_2}{\epsilon_1}\right) + \langle \beta \rangle_{zzz} \sin^2 \alpha \left(\frac{\epsilon_w}{\epsilon_1}\right)^2 \quad 4$$

where ω is the angular frequency of the incident radiation, α is the angle of incidence (45° in our case), ϵ_w , ϵ_1 and ϵ_2 are the dielectric constants of the water subphase and of the film at ω and 2ω respectively. $\langle \beta \rangle_{zxx}$, $\langle \beta \rangle_{zzz}$ are the two distinct nonvanishing average molecular hyperpolarizability components for the isotropic film considered here. For a rod shape molecule like retinal, the second-order molecular hyperpolarizability tensor is dominated by the one component corresponding to the incident and second-harmonic electric fields lying along the axis of the molecule. If the molecular axes are inclined at an average angle θ to the subphase normal but have a random distribution of azimuthal angle, then

$$\begin{aligned} \langle \beta \rangle_{zzz} &= \beta \cos^3 \theta \\ \langle \beta \rangle_{zxx} &= \frac{1}{2} \beta \sin^2 \theta \cos \theta \end{aligned} \quad 5$$

The vanishing value of $E_{p \rightarrow p}(2\omega)$ implies that the inclination angle of

the retinal molecule has a value of 59° . This is consistent with the molecular orientation measurement by the Heinz's nulling technique (as described above and shown in Figure 3).

We have also observed the relaxation phenomenon of the all-trans retinal LB film. The LB film was compressed to have 11 dynes/cm of surface pressure and then was kept at the constant area condition. The time course of the molecular orientation and surface pressure were recorded during the film relaxation. We found that the molecular directions closely follow the surface pressure (see Figure 5). This suggests that the force which keeps the molecules standing up from the water surface mainly comes from the steric interaction between molecules and the same force takes part in generating the surface pressure.

Correlation of Second Harmonic Generation with Dipolar Properties of Retinal and Retinal Schiff Bases

Recent studies of nonlinear second order optical processes in organic molecules have shown that two factors are important for a molecule to have large nonlinear optical response to an external applied field.¹⁸ The first is that molecules must have a highly polarizable π -electron system and the second is that the π -electron system has to be distorted by interaction with strong electron donor and acceptor groups. Although retinal does not have explicit electron donor and acceptor groups, the charge transfer by way of the π -electron system can occur during the electronic excitation ${}^1B_u \leftarrow S_0$. Quantum-

chemical calculations¹⁹ have demonstrated that there is a dramatic change in charge distribution from the β -ionone ring to the carbonyl group such that the oxygen is more negative in the first excited singlet $\pi\pi^*$ state. Mathies et al. has used an electrochromic technique to monitor the electronic dipole moments of retinal, unprotonated and protonated NRB during the electronic excitation $^1B_u \leftarrow S_0$.²⁰ Their data confirmed that retinal's condensed-phase dipole moment increases substantially (14.7 Debyes) upon excitation with the unprotonated NRB having slightly less change (10.2 Debyes) in dipole compared to retinal. In protonated NRB, the proton attached to the Schiff base nitrogen encourages the charge-transfer process and therefore increases the dipole moment change to 15.2 Debyes. Charge-transfer of this type should provide a substantial contribution to the second-order molecular hyperpolarizability. A simple two-level model has been used²¹ to express this term as:

$$\beta = \frac{3e^2\hbar^2}{2m} \frac{W}{[W^2 - (2\hbar\omega)^2][W^2 - (\hbar\omega)^2]} f\Delta\mu_{ex} \quad 6$$

where $\Delta\mu_{ex}$ is the difference between excited- and ground-state dipole moments, f is the oscillator strength of transition, $\hbar\omega$ is the fundamental photon energy, and W is the energy of transition. If the charge-transfer process is the major mechanism in the second-order nonlinear optical response of retinal, unprotonated and protonated NRB, the SH intensities from these three species should show the same increasing tendency from unprotonated NRB to retinal to protonated NRB as seen in the data of Mathies and Stryer.²⁰ Table I summarizes our SH measurements for these molecules. We have found that the SH intensity from an

unprotonated NRB LB film at a surface pressure of 13 dynes/cm is 70% of the SH intensity observed from a retinal LB film under the same conditions. By correcting for oscillator strength and energy dispersion factors, we have obtained 0.73 for the ratio of the dipole moment changes between unprotonated NRB and retinal, close to the 0.69 value from the data of Mathies and Stryer. The SH signal from protonated NRB film is 3.7 times stronger than that of unprotonated NRB. After the same correction procedure, we have a ratio 0.97 between protonated NRB and retinal whereas the Mathies and Stryer data gives a value of 1.03. The consistency of our data with the work by Mathies and Stryer using the very different Stark shift technique is most encouraging.

By calibrating the SH signal against a thin x-cut quartz plate, the second-order molecular hyperpolarizability of retinal is estimated to be 1.4×10^{-28} esu, which is over two times larger than 2-methyl-4-nitroaniline ($\beta = 4.5 \times 10^{-29}$ esu at $0.83 \mu\text{m}$). A value of 1.2×10^{-28} esu for unprotonated NRB and 2.3×10^{-28} esu for protonated NRB were found. The large second-order molecular hyperpolarizability that we have measured and the near 55° adsorbed direction insure that such molecules are efficient SH generator. Recently A. Gierulski et al.²² has demonstrated a novel technique to measure the pulsewidth of a subpicosecond laser based on the SH generation from a dye-coated substrate. The time resolution is found to be limited by the transverse relaxation time (T_2) of dye molecules. We have used a temporarily incoherent broadband Stilbene-420 dye laser with 68 femtosecond correlation time to measure the incoherent photon echoes²³ from protonated and unprotonated NRB. Our results show²⁴ that the T_2 of protonated NRB at 420nm is shorter

than 15fsec. Our experiment suggests that retinal, protonated and unprotonated NRB can be very efficient time-correlation devices for subpicosecond light pulse measurements.

4. Conclusion

In conclusion, we have measured the second harmonic generation of retinal, unprotonated, and protonated retinylidene n-butylamine Schiff bases at air-water and air-glass interfaces. Our data support the conclusions of other workers that the first excited singlet state dipole moments of retinal and its Schiff bases have a major contribution from the charge-transfer process which takes place from the β -ionone ring to the end groups of these molecules. The long-axes of the molecules tilt away from the surface normal by nearly 55° . The second-order molecular hyperpolarizability of protonated NRB is larger than unprotonated NRB and the increment can be explained by the stabilization of the π -electron system provided by the proton which is attached to the Schiff base nitrogen. We believe that this technique could readily be extended to other relevant biological and chemical applications which require the investigation of dipolar and structural relationships between related systems.

The results reported in this paper are currently being extended to determine the light induced alteration of the dipolar characteristic of the retinylidene chromophore in the membrane bound rhodopsin.²⁵

Table I. Comparisons of the second-order molecular hyperpolarizabilities and the dipole moment changes of the first excited $\pi\pi^*$ singlet state for all-trans retinal, unprotonated and protonated retinylidene n-butylamine Schiff bases

molecules	surface	second-order	excited-stated	
	susceptibility $\chi_{xxx}^{(2)}$ (esu) ($N_s = 2.5 \times 10^{14}$)	molecular hyperpolarizability β (esu)	dipole moment changes (relative to retinal) this work	Mathies's
all-trans retinal	6.6×10^{-15}	$(1.4 \pm 0.4) \times 10^{-28}$	1	1
unprotonated retinylidene n-butylamine Schiff Base (NRB)	5.8×10^{-15}	1.2×10^{-28}	0.73	0.69
protonated retinylidene n-butylamine Schiff Base ($N^+RB-HCl$)	1.1×10^{-14}	2.3×10^{-28}	0.97	1.03

References

- (1) For review see, Methods in Enzymology, Vol. 88, Part I, ed. L. P. Packer (Academic Press, New York, 1982).
- (2) Nestor, J. R.; J. Raman Spectros. 1978, 7, 90.
- (3) Levenson, M.D.; Introduction to Nonlinear Laser Spectroscopy (Academic Press, New York, 1982).
- (4) Shen, Y. R.; Ann. Rev. Mater. Sci. 1986, 16, 69.
- (5) Tom, H. W. K.; Mate, C. M.; Zhu, X. D.; Crowell, J. E.; Heinz, T. F.; Phys. Rev. Lett. 1984, 52, 348.
- (6) Tom, H. W. K.; Mate, C. M.; Zhu, X. D.; Crowell, J. E.; Shen, Y. R.; Somorjai, G. A.; Surf. Sci. 1986, 172, 466.
- (7) Rasing, Th.; Berkovic, G.; Shen, Y. R.; Grubb, S. G.; Kim, M. W.; Chem. Phys. Lett. 1986, 130, 1.
- (8) E.g.: Second International Conference on Langmuir-Blodgett Films, Schenectady, New York, 1985, ed. Zemel, J. N.; Thin Solid Films, 132, 133, 134.
- (9) Marcus, M. A.; Lemley, A. T.; Lewis, A.; J. Raman Spectros. 1979, 8, 22.
- (10) (a) Becker, R. S.; Freedman, K.; J. Am. Chem. Soc. 1985, 107, 1477.
(b) Becker, R. S.; Freedman, K.; Lenoble, C.; J. Phys. Chem. 1986, 90, 4334.
- (11) Heinz, T. F.; Tom, H. W. K.; Shen, Y. R.; Phys. Rev. A 1983, 28, 1983.
- (12) Dick, B.; Gierulski, A.; Marowsky, G., Reider, G. A.; Appl. Phys. B 1985, 38, 107.
- (13) Miller, R. C.; Appl. Phys. Lett. 1964, 5, 17.

- (14) (a) Jerphagnon J.; Kurtz, S. K.; J. Appl. Phys. 1970, 41, 1667.
- (b) Miller, R. C.; Phys. Rev. 1963, 131, 95.
- (15) Heinz, T. F.; Chen, C. K.; Ricard, D.; Shen, Y. R.; Phys. Rev. Lett. 1982, 48, 478.
- (16) Nakahara, H.; Fukuda, K.; Möbius, D.; Kuhn, H.; J. Phys. Chem. 1986, 90, 6144.
- (17) Girling, I. R.; Cada, N. A.; Kolinsky, P. V.; Montgomery, C. M.; Electron. Lett. 1985, 21, 169.
- (18) Garito, A. F.; Singer, K. D.; Teng, C. C.; in Nonlinear Optical Properties of Organic Materials, ed. Williams, D. J. (ACS, Washington, D.C., 1983).
- (19) Birge, R. R.; Hubbard, L. M.; J. Am. Chem. Soc. 1980, 102, 2195.
- (20) (a) Mathies, R.; Stryer, L.; Proc. Natl. Acad. Sci. USA 1976, 73, 2169.
- (b) Ponder, M.; Mathies, R.; J. Phys. Chem. 1983, 87, 5090.
- (c) Davidsson, A.; Johansson, L. B. A.; J. Phys. Chem. 1984, 88, 1094.
- (21) Frazier, C. C.; Harrey, M. A.; Cockerham, M. P.; Hand, H. M.; Chauchard, E. A.; Lee, C. H.; J. Phys. Chem. 1986, 90, 5703; Oudar, J. L.; Chemla, D. S.; J. Chem. Phys. 1977, 66, 2664.
- (22) Gierulski, A.; Marowsky, G.; Nikolaus, B.; Vorobev, N.; Appl. Phys. B 1985, 36, 133.
- (23) Nakatsuka, H.; Tomita, M.; Fujiwara, M.; Asaka, S.; Optics Commun. 1984, 52, 150.
- (24) Huang, J.; Lewis, A.; unpublished results.

(25) Rasing, Th., Huang, J.; Lewis, A.; Stehlin, T.; Shen, Y. R.;
to be published.

Figure Captions

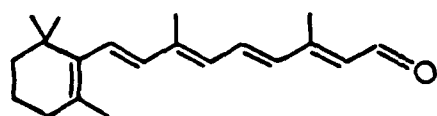
Fig. 1 The chemical structures of (a) all-*trans* retinal, (b) all-*trans* retinylidene n-butylamine Schiff base (NRB) and (c) protonated all-*trans* retinylidene n-butylamine Schiff base ($N^+RB-HCl$).

Fig. 2 The surface pressure-surface area (π -A) isotherms of (a) all-*trans* retinal (b) unprotonated retinylidene n-butylamine Schiff base (NRB) Langmuir-Blodgett monolayers on a pure water subphase. The dashed lines indicate the extrapolations of the linear parts to zero pressure. The limiting areas are $58\text{\AA}^2$ for retinal and $69\text{\AA}^2$ for NRB respectively. The cross point of these two isotherms is at the point where the surface pressure is 13 dynes/cm and the surface area is $38\text{\AA}^2/\text{molecule}$.

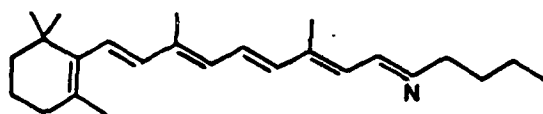
Fig. 3 The molecular orientation at different surface pressures. The open circles and open boxes are for all-*trans* retinal and retinal Schiff base (NRB) monolayers respectively. The open triangles are for NRB on a 4mM $CdCl_2$ water subphase. The inset figure indicates the orientation parameter that was measured.

Fig. 4 The second harmonic (SH) intensity of retinal monolayer at different surface pressures. The SH p-polarized intensity with an s-polarized pump beam, $I_{s \rightarrow p}$, which is proportional to the absolute square of the second-order susceptibility $\langle \beta \rangle_{zxx}$ of the molecular monolayer whose surface coincides with the xy-plane, is shown in open circles. The solid circles are for $I_{p \rightarrow p}$ SH intensity, which drops to zero when surface pressure is above 14 dynes/cm. The solid lines through the points are only intended to guide the reader.

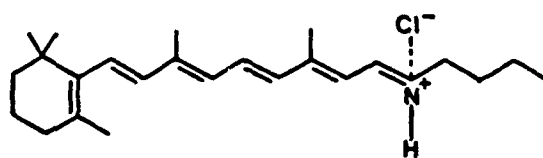
Fig. 5 The relaxation behavior of the molecular orientation (open circles) and surface pressure (solid line) of the all-trans retinal LB monolayer. The LB film was first compressed to 11 dynes/cm and then let it relax under the constant area condition.



(a) All-trans Retinal

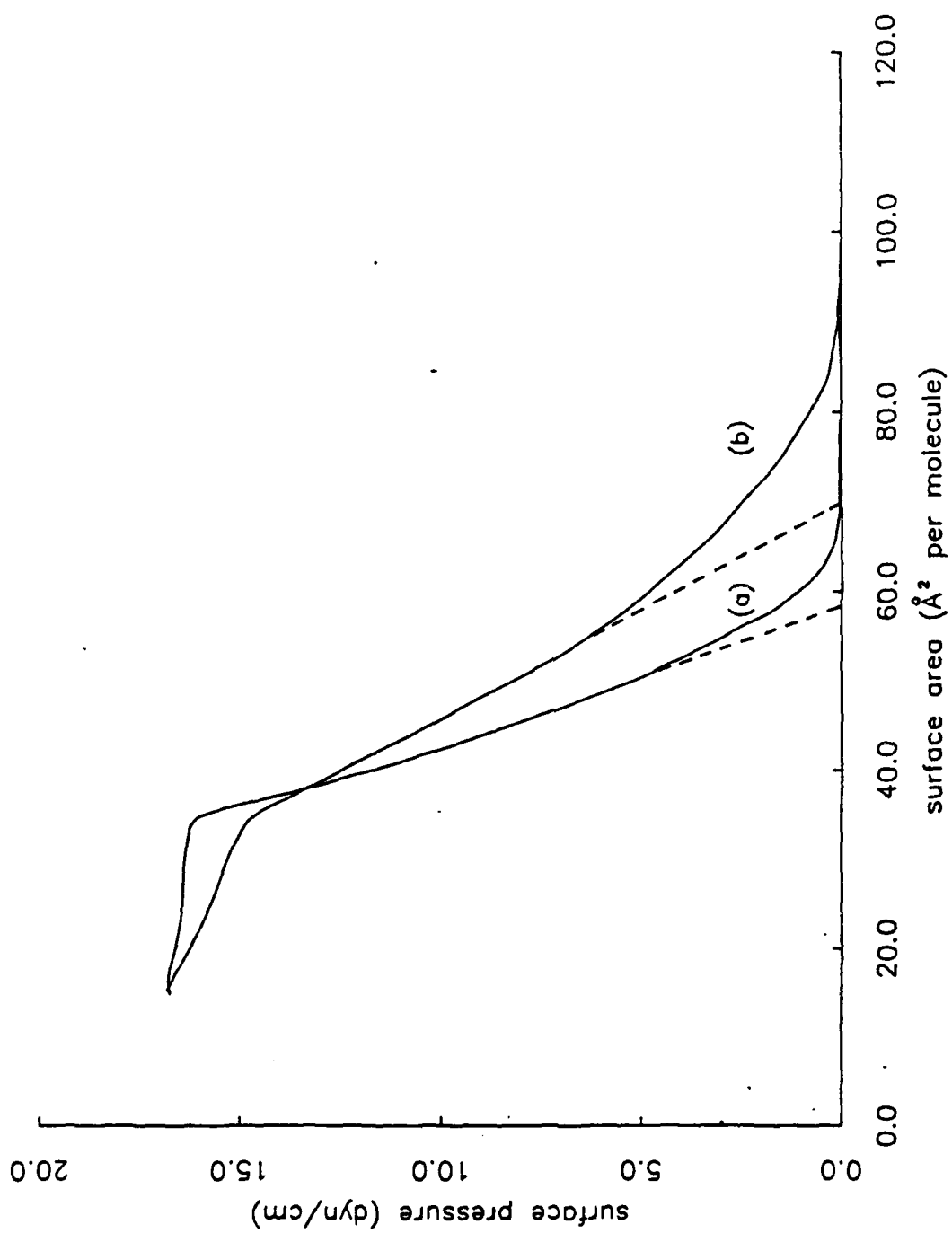


(b) All-trans Retinylidene n-Butylamine
Schiff Base (NRB)

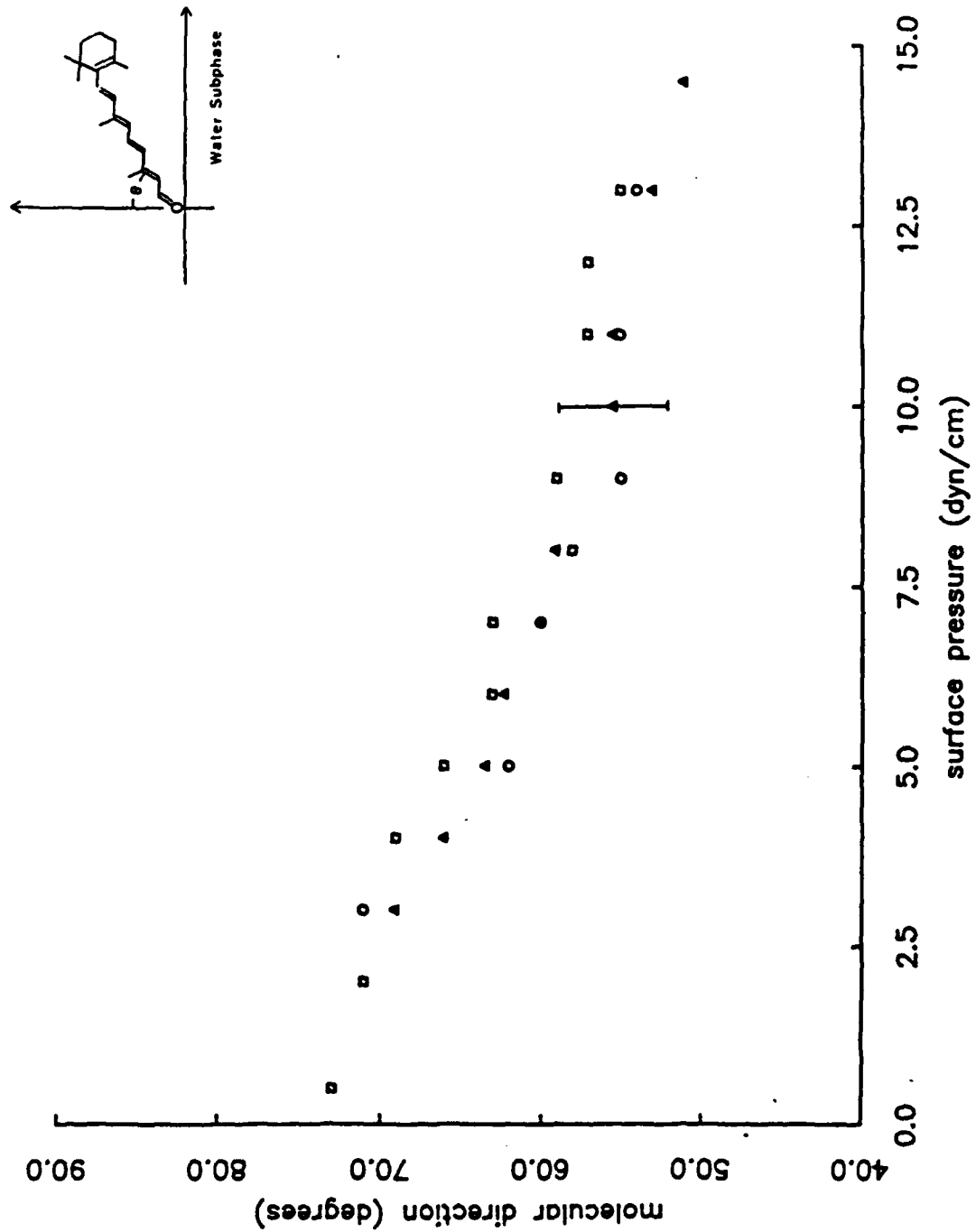


(c) Protonated All-trans Retinylidene
n-Butylamine Schiff Base (NRB-HCl)

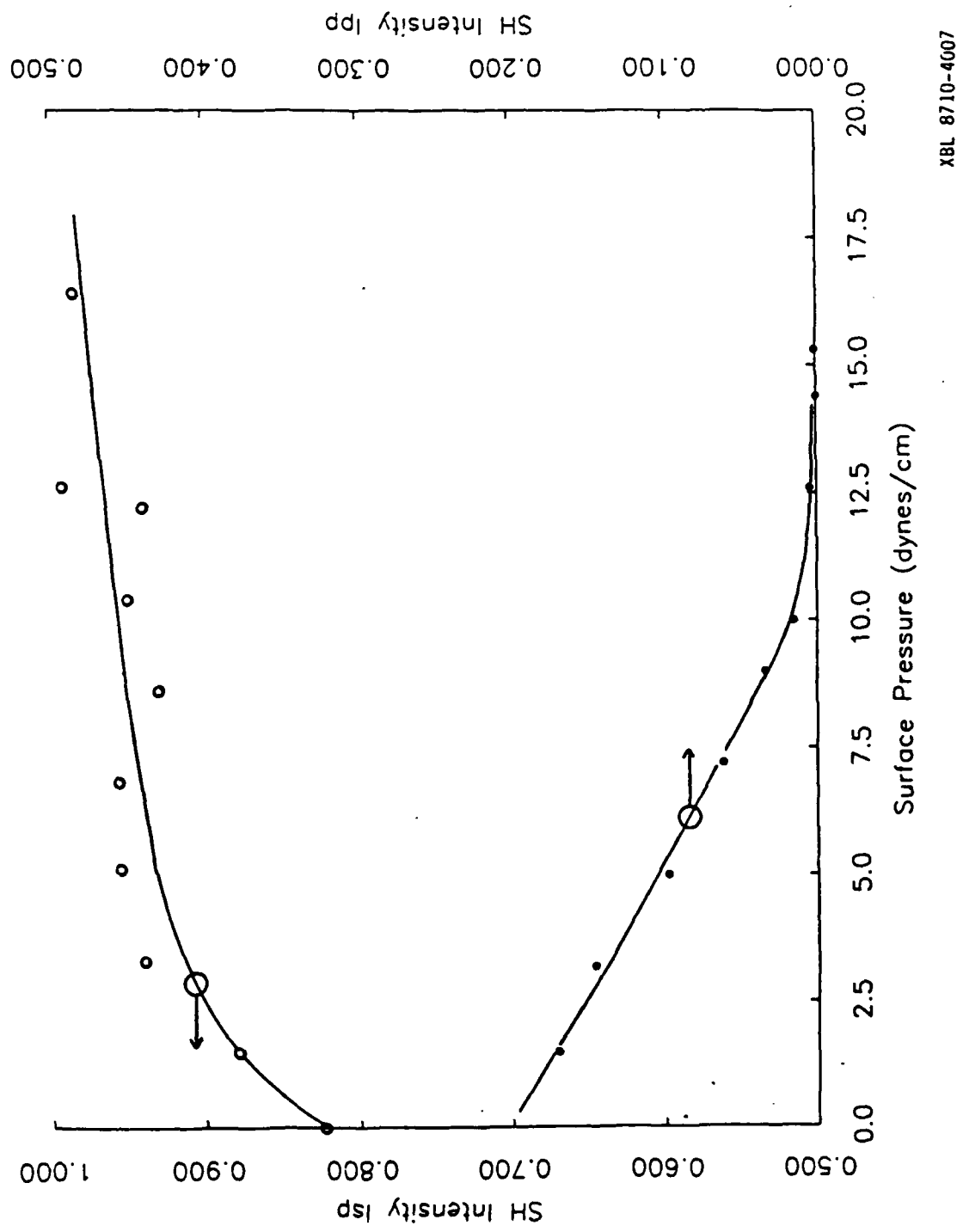
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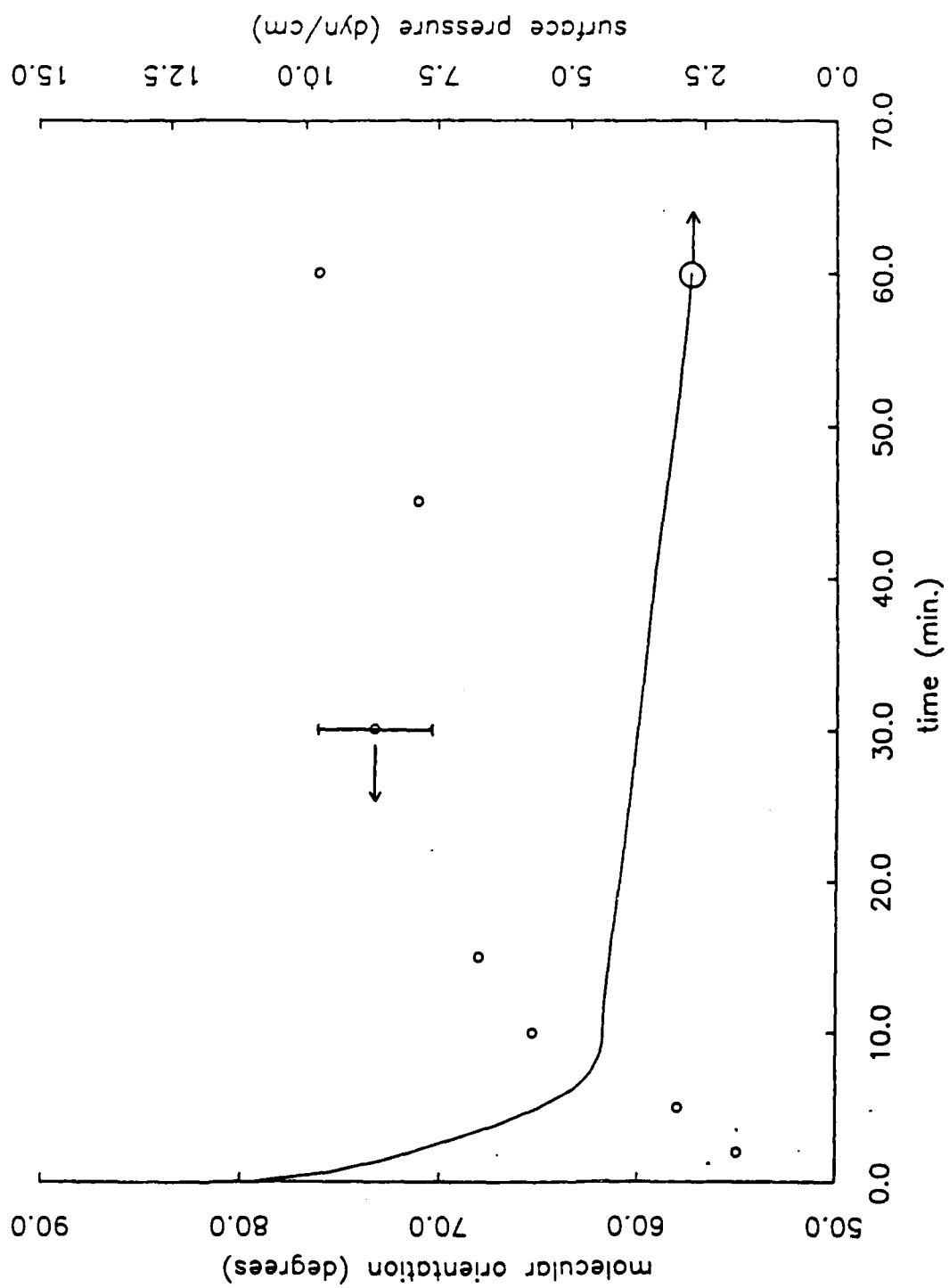


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