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1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE	3. REPORT TYPE AND DATES COVERED		
	7 May. 96	technical; 01JUNE 95 to 30SEPT96		
4. TITLE AND SUBTITLE			5. FUNDING NUMBERS	
Enzymatic Polymerization of Phenolic Biomonomers Derived from Cashew Nut Shell Liquid (CNSL)			C: N00014-90-J-1148	
6. AUTHOR(S)			R&T Code: 4132016	
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7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)			8. PERFORMING ORGANIZATION REPORT NUMBER	
University of Massachusetts Lowell Department of Chemistry 1 University Avenue Lowell, MA 01854			1148-95-05	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
Office of Naval Research-Physical S&T Division, ONR 331 Department of the Navy Arlington, Virginia 22217-5000				
11. SUPPLEMENTARY NOTES			19960517 171	
12a. DISTRIBUTION / AVAILABILITY STATEMENT			12b. DISTRIBUTION CODE	
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13. ABSTRACT (Maximum 200 Words)				
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14. SUBJECT TERMS			15. NUMBER OF PAGES	
Cashew nut shell liquid(CNSL), enzymatic polymerization, horseradish peroxidase (HRP)			20	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT	18. SECURITY CLASSIFICATION OF THIS PAGE	19. SECURITY CLASSIFICATION OF ABSTRACT	20. LIMITATION OF ABSTRACT	
UNCLASSIFIED	UNCLASSIFIED	UNCLASSIFIED	UL	

Standard Form 298 (Rev. 2-89)  
Prescribed by ANSI Std. Z39-18  
298-102

**OFFICE OF NAVAL RESEARCH**

GRANT N00014-90-J-1148

R&T Code 4132016

Scientific Officer: Dr. Kenneth Wynne

Technical Report No. 95-05

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Submitted to

**Journal of Macromolecular Science-Pure and Applied Chemistry**

**University of Massachusetts Lowell  
Department of Chemistry  
Lowell, Massachusetts**

May 7, 1996

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# Enzymatic Polymerization of Phenolic Biomonomers Derived from Cashew Nut Shell Liquid (CNSL)

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## Abstract

Enzymatic polymerization of alkyl phenols derived from a renewable resource, viz. cashew nut shell, has been carried out in dioxane-water mixture. The reaction and reaction products have been characterized by a number of spectroscopic, microscopic and thermometric methods. These monomers have also been copolymerized with various substituted phenols. The homopolymers are soluble in organic solvents while the copolymers are crosslinked, with negligible solubility. Enzymatic polymerization in dioxane-water solvent mixture results in spherical particles in the case of homopolymerization while structures without distinctive morphologies are obtained in the case of copolymerization. The effect of dioxane-water composition and solution pH on the polymerization reaction has also been explored.

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## Introduction

The use of enzymes in polymer synthesis has attracted the attention of many researchers in recent years. Enzymes are becoming increasingly popular in organic synthesis[1-8]. Horseradish peroxidase (HRP) has been used for the enzymatic synthesis of polyphenols and polyanilines[1-5]. In the presence of hydrogen peroxide, peroxidases catalyze the oxidation of electron rich systems such as phenols and aromatic amines that eventually give rise to high molecular weight polymers through oxidative free radical coupling. The molecular weight and properties of these biochemically formed polymers can be manipulated by controlling the experimental conditions, unlike the non-enzymatic reactions.

In recent years, the synthesis of polymers from renewable resources has attracted the attention of many research workers for a number of reasons[9,10]. Growing environmental concern in the expanded use of petrochemicals is one reason. Cashew nut shell liquid (CNSL) (from *Anacardium occidentale, L.*) is one of the renewable resources rich in phenolic compounds[10]. CNSL contains four major components: anacardic acid, cardanol, cardol and 6-methyl cardol, whose structures are furnished in Figure 1. Although a variety of R groups is possible, all have the meta substitution of a 15 carbon long hydrocarbon chain, with varying degrees of unsaturation with an average of two double bonds. Tyman and Jacobs[11] showed that the relative proportions of unsaturated components of anacardic acid are monoene (38.7%), diene (36.3%) and triene (45.1%). Cardanol upon hydrogenation in the presence of Pd/C gives hydrogenated cardanol where the C<sub>15</sub> hydrocarbon chain is saturated. Hence, cardanol is a potential renewable monomeric source for the synthesis of a variety of specialty polymers. In the present program, both cardanol and hydrogenated cardanol have been used for the HRP catalyzed polymerization in the presence of hydrogen peroxide, in dioxane water mixture.

## Experimental Procedures

Horseradish peroxidase was obtained from Sigma Chemicals Company, St. Louis, MO. All other chemicals were purchased from Aldrich Chemicals Company, Milwaukee, WI, and used as obtained.

Cashew nut shell liquid was obtained from Kerala Cashew Development Corporation, Quilon, India. Cardanol was obtained by double vacuum distillation of CNSL at 3-4 mm Hg and the fraction distilled at 230-240°C was collected. Hydrogenated cardanol was obtained by catalytic hydrogenation of cardanol. Hydrogen was passed over Pd/C catalyst at 25 psi at 60°C for a period of 6-8 hrs to saturate the side chains of the cardanol. Hydrogenated cardanol was extracted with ether, which was later evaporated off to obtain hydrogenated cardanol in the form of pale yellow solid.

Under atmospheric condition 5 mM of cardanol and 300 units of HRP were dissolved in 17 ml of 1,4-dioxane and 3 ml of phosphate buffer (pH 7.0). 0.25 ml of hydrogen peroxide (30%) was added to the reaction mixture every 15 minutes for 20 times with vigorous stirring at room temperature. After 24 hrs, the resulting polymer was isolated by evaporation of the solvent at reduced pressure followed by washing the residue with ice cold methanol and water followed by drying in vacuum.

Spectrophotometric experiments were carried out using Perkin-Elmer Lambda-9 UV-Vis-near IR spectrophotometer. Vibration spectra were recorded using a Perkin-Elmer FT-IR spectrometer 1760 with a room-temperature TGS detector. The pellets of the sample with KBr were used for FT-IR studies. TGA experiments were carried out using TGA 2950 (TA Instruments, Inc.,) thermobalance. The temperature was increased at a speed of 20°C/min under nitrogen atmosphere. SEM experiments were performed using an Amray 1400 Scanning Electron Microscope/Electron Probe

Microanalyzer. Gel permeation chromatographic (GPC) analysis was carried out on a GBR mixed bed linear column (Jordi Associates, Inc., Bellingham, MA). A UV detector at 270 nm was used to follow the elution. The eluent was DMF containing 1% lithium bromide. The flow rate was kept at 1 ml/min. NMR spectra were recorded using a Bruker 250MHz NMR spectrometer.

## Results and Discussion

Hydrogenation of liquid cardanol was followed by  $^{13}\text{C}$  NMR. Figure 2 gives a selected region of the  $^{13}\text{C}$  NMR spectra of the cardanol and hydrogenated cardanol in  $\text{CDCl}_3$ . It can be observed from the spectra that there is complete hydrogenation of the cardanol. The resonance peaks arising from the unsaturated hydrocarbons disappear upon hydrogenation.

The enzymatic polymerization of cardanol and hydrogenated cardanol (3-pentadecyl phenol) was carried out using horseradish peroxidase (HRP) in a mixture of dioxane and buffer (85:15) for 24 hrs at room temperature. The polymerization reaction was initiated through the addition of hydrogen peroxide. Hydrogen peroxide was added at 15 minutes time intervals for about 5 hours to continue the polymerization reaction and to minimize the inhibition of the enzyme by the substrate (hydrogen peroxide). The system becomes turbid almost instantaneously upon the addition of hydrogen peroxide; a visual indication of reaction progress. From the spectroscopic observation of the polymerization reaction at 250nm, it was determined that the reaction attains an equilibrium in about 2 minutes. On further addition of hydrogen peroxide, similar changes in absorbance were observed. The reaction was allowed to continue for about 24 hours with continued stirring to ensure completion of the reaction.

The dioxane-water composition for the polymerization was optimized through

a set of experiments, which is summarized in Table 1. Upon increasing the dioxane concentration from 20 to 95 (v/v with buffer), the percentage yield of the polymer increases from very low to 72% at 85% of dioxane. On further increasing the dioxane concentration, the yield is lowered. At low dioxane concentrations, the enzymatically formed polymers precipitate out of the solution as oligomers. Mostly, dimers or trimers were formed when the reaction was carried out at low dioxane content.

The polymerization of cardanol was carried out in 85% dioxane - 15% water varying the pH from 6 to 7.5. The objective here was to optimize the pH condition for optimum yield. The results are summarized in Table 2. It is observed that with increasing pH from 6 to 7 the percentage of yield increases and with further increase of the pH, it decreases. The enzyme activity was maximum at pH 7.0.

Cardanol and hydrogenated cardanol (3-pentadecyl phenol) were copolymerized with phenol, p-ethyl phenol and p-phenyl phenol, the results of which are furnished in Table 3. The numbers in the parenthesis of the first column indicate the mass ratio of the cardanol (unsaturated and hydrogenated) and the co-monomer. Copolymerization in 85% dioxane-15% buffer resulted in the rapid change of color of the solution to brown or dark brown as soon as hydrogen peroxide was added indicating the formation of the polymer. The reactions were completed within 20-30 minutes. The percentage yield of the copolymers were above 90%. The polymers are insoluble in common solvents which limited the determination of molecular mass of these polymers.

The morphology of the polyphenols of cardanol and hydrogenated cardanol in the presence and absence of the co-monomer has been studied. Figure 3 shows the scanning electron micrographs of polymers (a) hydrogenated cardanol and (b) hydrogenated cardanol with phenol copolymer (in 1:5 ratio), polymerized in dioxane-water mixture. It can be observed from the figure that the homopolymer shows

spherical particle structures with the radius of the order of  $0.5\mu\text{m}$  while the copolymer does not show any ordered morphology. Recently, it has been reported that short chain derivatives of phenols form spherical particles upon polymerization in the presence of surfactants<sup>12</sup>. We speculate that the long hydrocarbon chain of the cardanol and hydrogenated cardanol organizes the monomers along the water dioxane interface in the dioxane-water microemulsion, creating a micellar environment. During polymerization, the positions of these monomers are fixed in the spherical form, forcing the chain curvature along the micellar curvature. However, addition of phenols or short chain derivatives of phenols lowers the effect of hydrocarbon chain in organizing the monomers resulting in featureless structures. Similar structures have also been observed in the case of polymer of cardanol and cardanol-phenol copolymer.

The spectroscopic characterization of the polymers was carried out using FT-IR and NMR spectroscopic techniques. The NMR of the copolymers could not be taken due to the insolubility of the polymers. The homopolymer of cardanol showed characteristic changes in the aromatic region of the  $^1\text{H}$  spectrum. The aliphatic region of the spectrum resembled the monomer spectrum. In the aromatic region, there were new peaks appearing due to the polymerization, which were down shifted compared to the monomer peaks. The monomer shows aromatic peaks at 6.7, 6.8 and 7.2 ppm. The polymeric peaks appear at 6.9, 7.6, 7.8 and 8.1 ppm. The appearance of more than one peak due to the polymerization can be attributed to the complex nature of the polymer structure. The FT-IR spectra of monomers and polymers were compared. Figure 4 shows the FT-IR spectra of hydrogenated cardanol and its polymer. It can be observed from the figure that the line widths of the peaks in the polymer have broadened as compared to that of the monomer. The peak at  $950\text{ cm}^{-1}$  present only in the polymer spectrum corresponds to the phenyl-phenyl C-C bond. Similar observations were also made in the case of copolymers.

The TG thermogram of the copolymer of hydrogenated cardanol with p-phenyl phenol is shown in Figure 5. The characteristic thermal data of the copolymers prepared from cardanol and hydrogenated cardanol with substituted phenols are provided in Table 4 with the weight loss at different temperatures. A 2 to 10% weight loss occurs in case of all the copolymers when heated upto 200°C. This may be attributed to the moisture retained in the sample. About 50 -70% weight loss occurs in the region 450-500°C in the case of all the copolymers and 80% weight loss occurs at 800°C. In case of the copolymer prepared from hydrogenated cardanol and p-phenyl phenol, 20% weight loss occurs at 400°C and thereafter degradation is rapid. Kobayashi et al. have reported the TG analysis of the polyphenol prepared using HRP-hydrogen peroxide catalyst[13]. Comparing the thermal data of the polyphenol with cardanol-phenol or hydrogenated cardanol-phenol copolymer, it is evident that the homopolymer has greater thermal stability than the copolymers. Under nitrogen, 10% weight loss was observed at 387°C and 57% weight loss occurs at 1000°C. The difference in thermal stability is due to the structural difference of these two types of compounds. In the case of cardanol or hydrogenated cardanol, there is a long hydrocarbon chain at the meta position. The degradation of the long hydrocarbon chain occurs at a lower temperature. Similar observations have been reported earlier by Nayak et al, while studying the degradation pattern of cardanol-formaldehyde and phenol-formaldehyde resins[14].

Further, comparing the thermal stability of the three copolymers prepared from hydrogenated cardanol with phenol, p-ethyl phenol and p-phenyl phenol, it is observed that the copolymer from p-phenyl phenol is more stable upto 400°C (~20% weight loss) than the other two copolymers. The enhancement in stability is due to the rigidity of the copolymer because of the presence of a phenyl group at the para position. On the other hand, after 400°C, the degradation of this copolymer becomes rapid because of the fact that the breakage of the carbon-carbon bond of the phenyl

group at the para position might take place along with the degradation of the long hydrocarbon chain at the meta position.

## **Conclusions**

Polymers of monomers derived from cashew nut shell liquid have been enzymatically synthesized. The copolymers with various derivatives of phenols are insoluble in common solvents, while the homopolymers are low molecular weight oligomers. The polymer yield was observed to be dependent on the nature of the comonomer. The interfacial polymerization reaction has been observed to be very fast which required continuous stirring to achieve increased polymer chain length. The monomers organize in the micellar form with the polymer chain growing in the micellar curvature. The SEM studies indicate that the homopolymer forms a spherical particle while the copolymers with phenols show featureless structures. The surfactant property of the monomer aids the formation of these spherical particles. The thermal properties of the copolymers are dependent on the co-substrates. The copolymers degrade at lower temperature as compared to the homopolymeric polyphenols.

## **Acknowledgments**

Partial financial support from ONR and NSF is gratefully acknowledged. Thanks are due to D-W. Cheong, H. C. Wang and S. Balasubramanian for their assistance in some of the experiments. PLN thanks the Department of Higher Education, Govt. of Orissa, India for providing a leave of absence.

## Figure captions

Figure 1: Structures of the components of cashew nut shell liquid.

Figure 2:  $^{13}\text{C}$  NMR spectra of (a) cardanol and (b) hydrogenated cardanol in chloroform-d.

Figure 3: Scanning Electron micrographs of polymers of (a) cardanol and (b) cardanol with phenol.

Figure 4: FT-IR spectra of (a) hydrogenated cardanol and (b) polymer of hydrogenated cardanol.

Figure 5: Thermogram of copolymer of hydrogenated cardanol with p-phenyl phenol.

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Table 1: Effect of solvent/buffer ratios on the enzyme activity in the polymerization of cardanol.

Synthesis medium Dioxane:buffer <sup>a</sup>	Polymer yield (Percentage)	
	cardanol	hydrogenated cardanol
20:80	Negligible	Negligible
40:60	10	12
60:40	18	25
70:30	35	35
80:20	52	55
85:15	75 ( $M_n = 2200$ )	85
95:5	37	35

<sup>a</sup>-phosphate buffer pH 7.0

Table 2: Effect of pH on the enzyme activity in the polymerization of cardanol

<b>pH</b>	<b>Percentage yield</b>
6	Negligible
6.5	45
7	75
7.5	30

Table 3: Effect of substituents on the copolymerization of hydrogenated cardanol (3-pentadecyl phenol) with substituted phenols. The reaction was carried out in 85:15 diaoxane - buffer mixture at pH 7.0.

<b>Phenol</b>	<b>Percentage yield</b>
Phenol (3:1)	95
p-ethyl phenol (3:1)	90
p-phenyl phenol(2:1)	98
cardanol-phenol(1:2) <sup>a</sup>	95

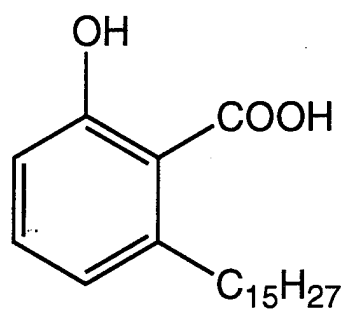
a- cardanol was used instead of hydrogenated cardanol.

Table 4: Thermal decomposition of the copolymers measured by thermogravimetric analysis

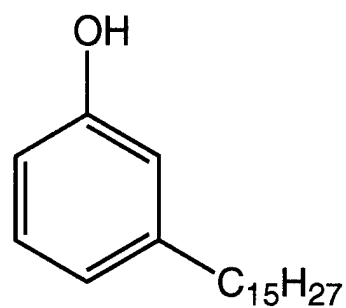
Polymer System	Percentage weight loss at various temperatures						
	200°C	300°C	400°C	500°C	600°C	700°C	800°C
card + phenol	8	17	35	55	62	68	75
cardH + phenol	12	23	38	48	-	-	-
cardH + p-ethyl phenol	5	12	28	58	67	76	81
cardH + p-phenyl phenol	3	8	22	78	81	85	88

card - cardanol

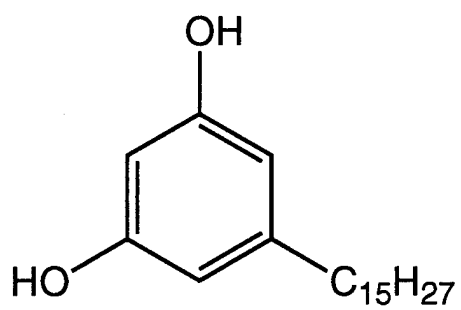
cardH - hydrogenated cardanol



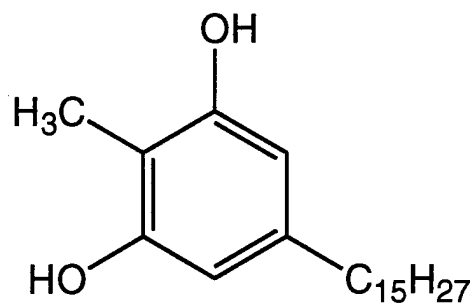
Anacardic acid



Cardanol



Cardol



6-methyl cardol

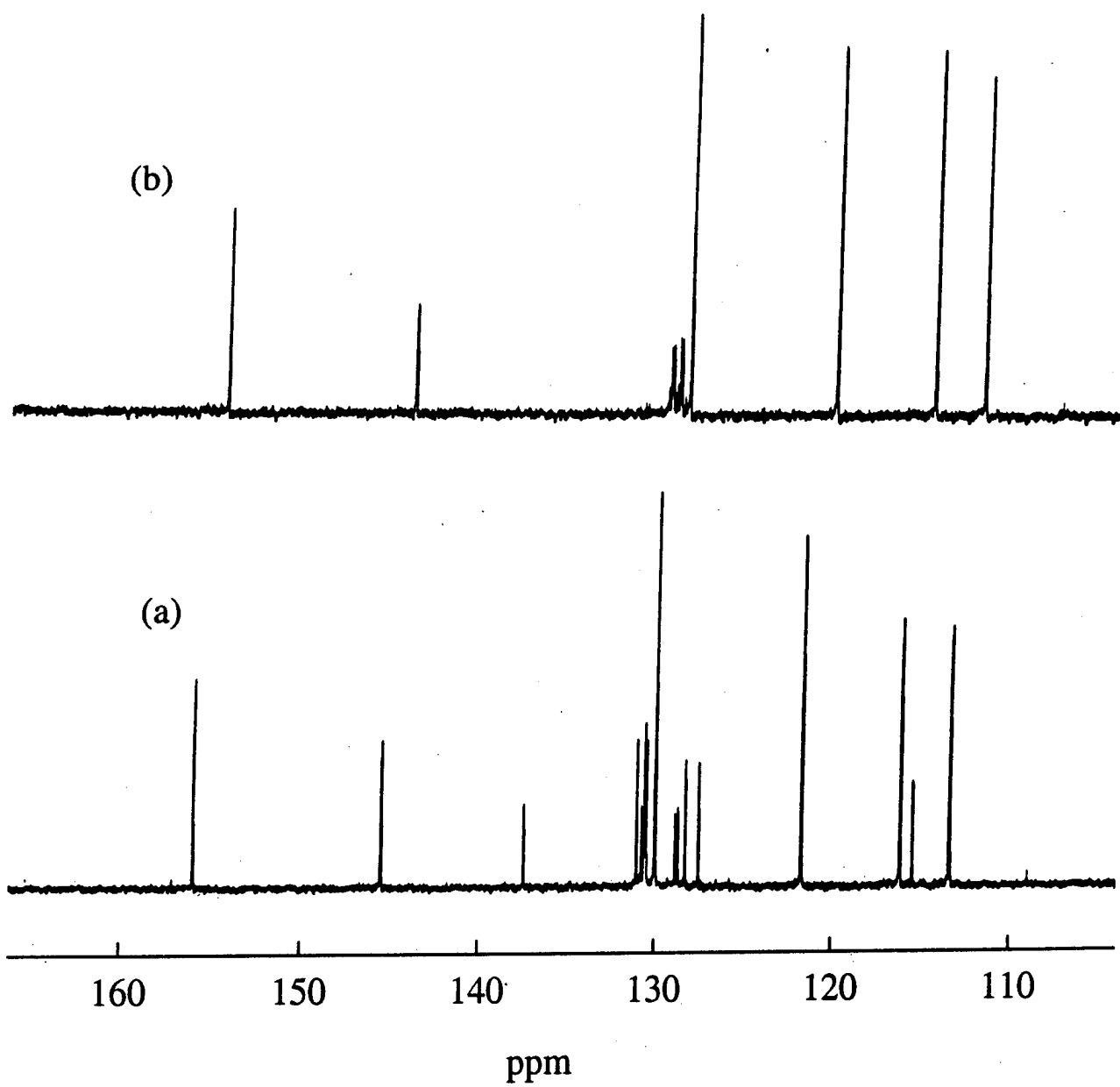
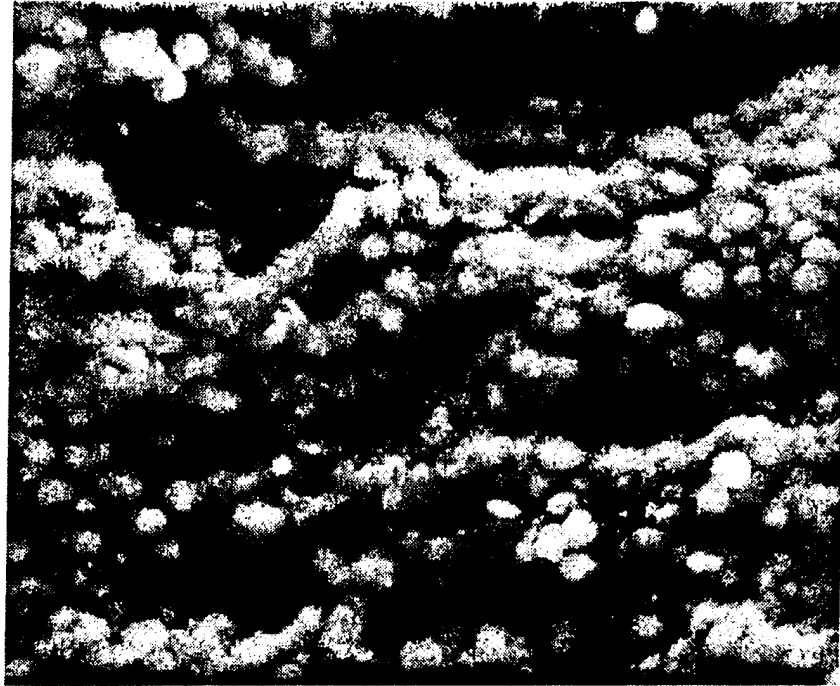


Fig. 2



2 $\mu$ m

10KeV

X5000

Fig 3(a)



2 $\mu$ m

10KeV

X5000

Fig 3(b)

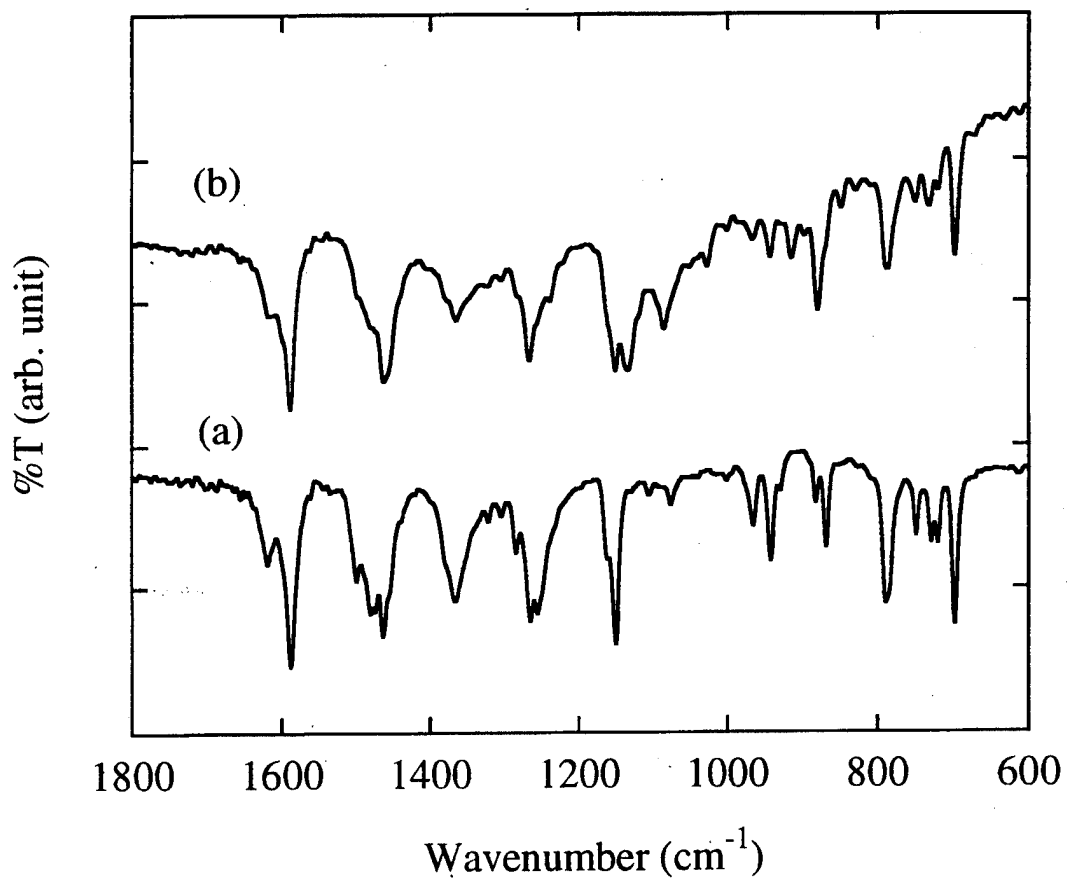


Fig 4

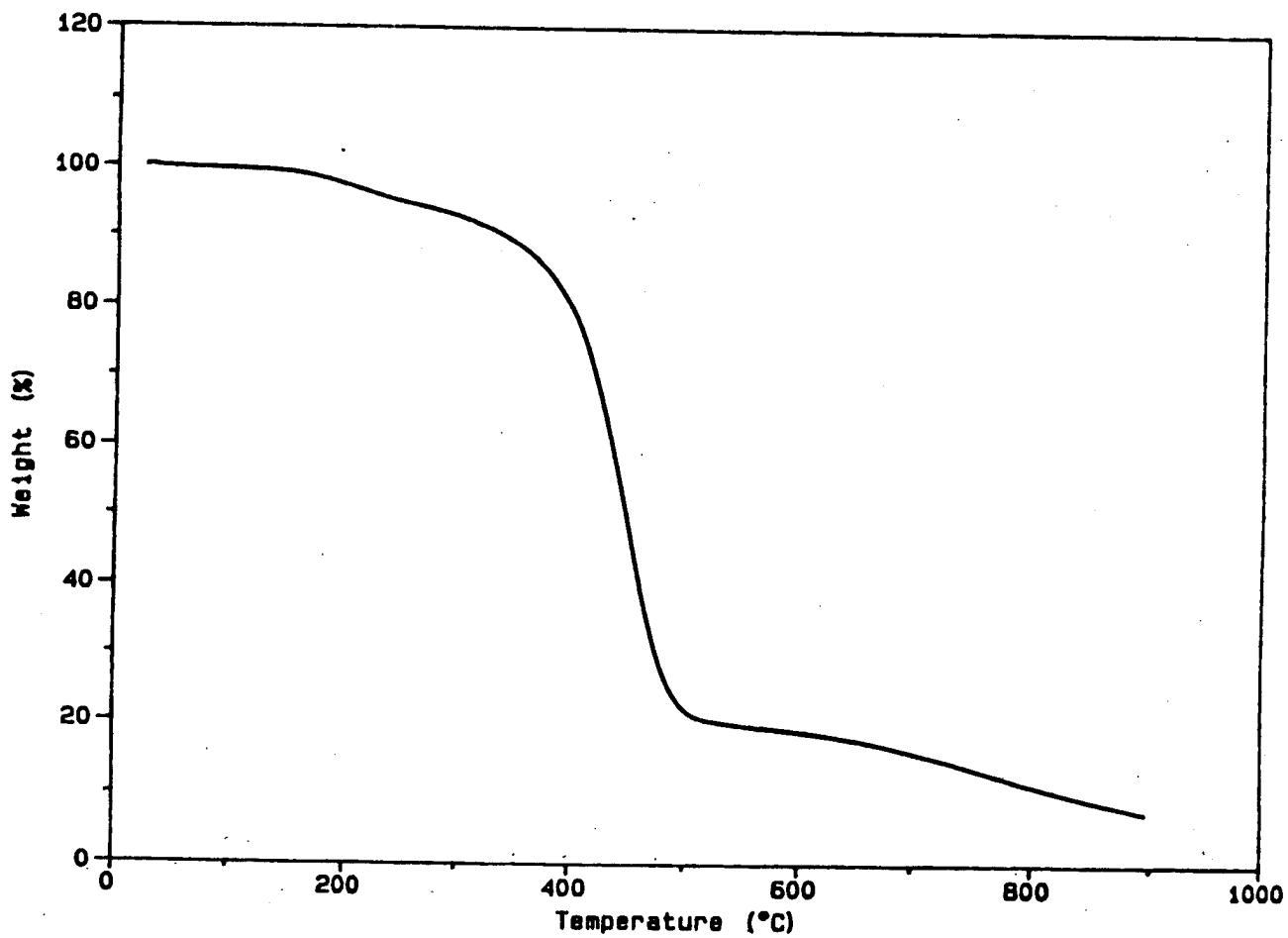


Fig 5