

AD \_\_\_\_\_

Grant Number DAMD17-96-1-6119

TITLE: MR Measurement of Breast Tissue's Anisotropic Mechanical Properties

PRINCIPAL INVESTIGATOR: John B. Weaver, Ph.D.

CONTRACTING ORGANIZATION: Dartmouth College  
Hanover, New Hampshire 03755

REPORT DATE: July 1997

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for public release;  
distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

**DTIC QUALITY INSPECTED 2**

**19971210 045**

# REPORT DOCUMENTATION PAGE

*Form Approved*  
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

<b>1. AGENCY USE ONLY (Leave blank)</b>	<b>2. REPORT DATE</b> July 1997	<b>3. REPORT TYPE AND DATES COVERED</b> Annual (1 Jul 96 - 30 Jun 97)	
<b>4. TITLE AND SUBTITLE</b>  MR Measurement of Breast Tissue's Anisotropic Mechanical Properties		<b>5. FUNDING NUMBERS</b> DAMD17-96-1-6119	
<b>6. AUTHOR(S)</b> John B. Weaver, Ph.D.			
<b>7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)</b> Dartmouth College Hanover, New Hampshire 03755		<b>8. PERFORMING ORGANIZATION REPORT NUMBER</b>	
<b>9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES)</b> U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012		<b>10. SPONSORING/MONITORING AGENCY REPORT NUMBER</b>	
<b>11. SUPPLEMENTARY NOTES</b>			
<b>12a. DISTRIBUTION / AVAILABILITY STATEMENT</b>  Approved for public release; distribution unlimited		<b>12b. DISTRIBUTION CODE</b>	
<b>13. ABSTRACT (Maximum 200)</b>  We are progressing well toward the original goals delineated in the grant. The MR sequences have been written to measure vibrations in all three directions. No attenuation of the vibration can be observed across a homogeneous sample. No vibration is seen in directions perpendicular to the induced vibration. These two results suggest that a simple linear model is appropriate. A linear model allows the elasticity to be calculated directly without elaborate calculations. This is encouraging. We have written and tested code to calculate the elasticity from measured phase changes assuming a linear model for the vibration. However, the phase changes used to calculate the vibrational displacements are relatively small using our current apparatus. Therefore, we are concentrating on increasing the phase changes by increasing the amplitude of the vibration and the strength of the magnetic field gradients. We have made significant progress toward the original objectives of this grant and we have accomplished some of the objectives for the second year of the grant such as measuring attenuation and scatter.			
<b>14. SUBJECT TERMS</b> Breast Cancer		<b>15. NUMBER OF PAGES</b> 12	
		<b>16. PRICE CODE</b>	
<b>17. SECURITY CLASSIFICATION OF REPORT</b> Unclassified	<b>18. SECURITY CLASSIFICATION OF THIS PAGE</b> Unclassified	<b>19. SECURITY CLASSIFICATION OF ABSTRACT</b> Unclassified	<b>20. LIMITATION OF ABSTRACT</b> Unlimited

FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

Y Where copyrighted material is quoted, permission has been obtained to use such material.

Y Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

Y Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

NA Y In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Resources, National Research Council (NIH Publication No. 86-23, Revised 1985).

NA Y For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

NA Y In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

NA Y In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

NA Y In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

John W. Wain 6/30/97  
PW - Signature Date

**Table of Contents**

	Page Number
Front Cover .....	1
Form SF 298 .....	2
Foreword.....	3
Table of Contents .....	4
Introduction .....	5
Body.....	7
Conclusions .....	9
References.....	10

## **MR Measurement of Breast Tissue's Anisotropic Mechanical Properties: Breast Cancer Detection and Classification**

### **Introduction:**

We are developing a magnetic resonance (MR) method of measuring the mechanical properties of tissue, including the hardness, quantified by the three dimensional (3D) modulus of elasticity, and the density. Tissue is vibrated at low frequency and the displacement of the tissue will be measured with MR imaging. The mechanical properties of the breast will be calculated from the measured displacements via a differential equation describing the motion. We will measure the elasticity in all three directions by vibrating the tissue and measuring the resulting three dimensional displacement. Measuring the 3D elasticity is important because the anisotropic distribution of fibrous tissue in the breast suggests that the elasticity will be anisotropic. In addition, we will test for nonlinearities and make sure that the measured displacements are consistent with the linear lossless wave equation; if it is not we will find the non-linearities most consistent with the data.

Elasticity measurements can play several roles in breast cancer detection and in evaluating treatment effectiveness. Elasticity may help classify lesions identified with mammography which is sensitive but not specific; roughly two thirds of the lesions detected with mammography turn out, on biopsy, to be neither malignant nor pre-malignant [1]. Secondly, because mammography can not detect all palpable lesions [5,2], elasticity measurements could supplement the physical examination and mammography in screening programs. The sensitivity of mammography with current technology is between 85% and 90% [3]. Elasticity measurement should be used as part of the screening examination if it catches some significant fraction of the missed malignancies. Abnormalities such as architectural distortions which are often missed in mammography [4] should be well visualized with elasticity measurements. We are developing 3D elasticity measurements to determine its usefulness in classification and screening. Once tissue properties are measured for a variety of tumors and benign lesions, less expensive methods can be developed that are designed to measure the relevant tissue properties.

Several different types of imaging techniques are being explored to supplement mammography: Doppler ultrasound [5,6], MRI contrast studies [7,8,9], radio nuclide uptake [10], and electrical impedance imaging [11]. We feel attempts to measure the elasticity of breast tissue are the most promising. Hardness or elasticity has a strong correlation with cancer. The physical examination is an important part of breast cancer screening [12,13,14,15,16]. In the Breast Cancer Detection Demonstration Project, around a third of malignancies were found with physical examination and not with mammography [17]. It is reasonable to look for similar changes in smaller lesions. There have been four approaches to measuring elasticity using ultrasound. None is capable of measuring anisotropic elasticity and all have significant practical problems in implementation. The first technique changed the compression and measured the resulting distortion

[18,19]. The second technique uses low frequency sound to vibrate the tissue and Doppler ultrasound to identify regions with large peak velocities [20,21,22,23,24]. Preliminary measurements of the "sonoelasticity" of masses in rodents and prostatectomy specimens have indicated that it correlates well with malignancy [7,28,25]. The third technique, ultrasonic computed tomography [26], measures the speed or attenuation of ultrasound. The fourth technique measures low frequency shear wave amplitudes with MR [27]. Obtaining quantitative measurements of tissue elastic properties from this technique is problematic because of attenuation and dispersion of the shear wave [28, 29]. Low frequency transverse waves are not attenuated or significantly dispersed over the thickness of the breast.

However, little quantitative work has been done studying the mechanical properties or behavior of soft tissue. Heart wall, heart valves and cartilage show anisotropic elasticity [30,31,32,33]. Aortic valves have also shown viscoelastic properties [34]. However, no systematic studies have been made to identify the appropriate models for soft tissue such as breast tissue. Such information would be important for detecting and classifying lesions and in breast reconstructions.

Therefore, we have been developing methods of studying vibrational motion of breast tissue. We have implemented an MR imaging technique to measure the displacement of tissue during periodic vibration. The linear, elastic model can be used for anisotropic material if the elasticity is measured in all three directions. This is important because the elasticity of breast tissue is probably anisotropic; the fibrous tissues in the breast tend to be oriented. However, if viscous or attenuation effects become important, the measured value of elasticity will be different when measured in different environments. In this case, accurate estimates of the elasticity might still be made using the measured displacements and a finite element model of the correct equations of motion [35].

Our long term hypothesis is that elasticity measurements will 1) contribute toward the accurate classification of lesions detected with mammography and 2) detect a significant number of the malignancies missed by mammography. The hypothesis we are testing in this proposal is that we can measure the mechanical properties of tissue with MR using a linear, lossless motion model. We are also establishing that the properties measured completely describe the vibration of tissue so there is no other information to be gained from measurements of vibration. This is important to establish before trials with patients are started so the correct measurements are made.

The specific technical objectives we planned for the first year of this project were: 1) Refine the MR measurement of three dimensional displacement during vibration. 2) Compare the elasticity calculated from the MR displacements with a linear model to known elasticity's for isotropic and anisotropic materials.

Some of the objectives from the second year were also accomplished: Measure motion perpendicular to the direction of forced vibration. Measure viscous losses by the attenuation of the vibration across the material.

**Body:**

The first objective of this grant was to refine the MR measurement of three dimensional displacement during vibration. The first element of the objective was to rewrite the MR sequence we used to obtain displacements. We improved the timing and added motion sensitizing gradient pulses in all three directions so we can measure displacement in all three directions. We are also moving from a GE 4.8 machine to a GE 5.x Epic machine which required rewriting the sequences.

The second element necessary for the first objective was reducing the noise fed into the MR from the audio system that powers the piezoelectric crystal vibrating the sample. We have been able to reduce the noise to levels below the ambient noise without reducing image quality; this is a reduction of almost an order of magnitude. The signal to noise ratio (SNR) of motion sensitized images is now the same as that of standard images using the same imaging parameters. A series of incremental improvements were required. A better audio amplifier reduced the noise somewhat. The transformers used to couple the amplifier to the crystal can be used to filter out most of the remaining audio amplifier noise. Shielding the leads of the secondary winding of the transformer reduced the noise further. The final step was isolating the coaxial shields from the audio circuit and grounding them to the room itself. Grounding always seems to be somewhat of an art. It is much simpler to put a shield around the crystal itself but the shield produces eddy current artifacts in the images which are difficult to isolate from the sample.

The final element of the first objective was increasing the power of the system which was relatively easy. We have increased the voltage and power of the system.

The second objective we set was to compare the elasticity calculated from the MR displacements with a linear model with known elasticity's for isotropic and anisotropic materials.

The first element of this objective was to measure displacement amplitudes along to the direction of the induced motion and perpendicular to the direction of the induced motion. The displacements were measured and they are consistent with a linear model. We do not observe any change in the magnitude of the displacement across a homogeneous sample. This is a good indication that the system is indeed linear. We see no motion in directions perpendicular to the induced vibration. To measure the displacement with sufficient accuracy, we had to average over many pixels. To study the amplitude of the displacement along the direction of the induced vibration, we averaged pixels along lines perpendicular to the induced motion. To look for motion perpendicular to the induced motion, we averaged pixels along lines parallel to the induced motion. We need to increase our sensitivity.

The second element of the second objective was to calculate the elasticity from MR displacement measurements. Code was written in MATLAB to calculate the elasticity from a series of images taken with different lags between the motion and the motion sensitizing gradients.

At least two lags are needed to calculate the displacement; we use more at present to increase SNR's. Simulations were run to check the code and study sensitivity.

The phase differences induced by vibration are used to calculate the elasticity. Those phase differences are proportional to the product of the displacement and the gradient strength. The combination of gradients and displacement we are now able to generate experimentally produces approximately one degree phase changes. By averaging several hundred pixels we can obtain reasonable accuracy but averaging can not be used for inhomogeneous materials and limits the accuracy of the results for homogeneous samples.

Therefore, we are working to increase both the gradient field strength and the displacement. We have limited amounts of data using an add on head coil with twice the gradient strength we have on the stock MR. The coil is designed for functional magnetic resonance of the brain by Advanced NMR Systems. The data is better but not good enough. Therefore, we are in the process of porting the MR sequence to a small bore MR imager made by SMIS that has gradients that are 20 times as strong as those on the clinical system. We are making a vibrational system small enough to fit into the small bore system.

We are studying three methods of increasing the vibrational displacement in the clinical MR: A bimorph piezoelectric element has sufficient displacement but the force is only sufficient for small samples (50 grams). A bimorph is a two piezoelectric elements bonded together with opposite polarities so when one is expanding the other is contracting. A positive voltage flexes the pair into an arc and a negative voltage flexes the pair into an arc the other direction. The second option is a stacked piezoelectric actuator with many elements; the displacement of each element is independent of thickness so many thin elements produce a much larger displacement. However, a lever arm would be needed to increase the displacement to desired levels. This kind of actuator has sufficient force and displacement if the lever arm can be made strong enough. The third option is a pair of rotating shafts on either side of the sample. Each shaft would have a slightly off center or bent segment that pushes the object back when they were rotated. The shafts would be rotated with an electric motor. This option provides sufficient force and displacement but the materials might have to be constructed from stainless steel which might cause eddy current artifacts that are too large. The size of the eddy current artifacts will depend on how we couple the shafts to the sample. We are doing experiments to see how strong we can make the lever for the piezoelectric actuator and how well we can isolate the shafts from the sample.

The lack of sufficient displacement with piezoelectric crystals can also be solved by working at resonance frequencies where the displacement is more than an order of magnitude larger than when driven at low frequencies. The currently used technique accumulates phase in the MR image by following the displacement with the gradient waveform. Because the gradients can not follow displacements that are

faster than 1kHz, we are looking at other mechanisms to detect the higher frequency displacement. We have simulated periodic displacement at higher frequencies during an RF pulse. The simulations show reduced amplitudes of magnetization that reaches the transverse plane resulting in reduced signal amplitude. The reduction is nearly independent of the frequency of the displacement. The reduction in signal is not linear with displacement but it is monotonic so the displacement can be estimated from the signal reduction. We are now designing RF pulses to increase the magnitude of the signal reduction to usable levels. These results suggest that high frequency displacement during the RF pulse is measurable. Therefore, piezoelectric crystals can be used to induce measurable displacements at resonance. The intriguing aspect of this technique is that the properties of the tissue as a function of frequency can be studied.

We are also working on a related project that is not being funded by this grant. We are developing an elastic image alignment algorithm. We are planning to use our image alignment algorithm to measure static displacements. This would allow the elasticity to be estimated using a few assumptions as other groups have been doing [17,18].

However, our primary efforts are directed toward measuring the low frequency displacement with the method described in the original proposal by increasing the gradients and displacement.

### **Conclusions:**

We are progressing well toward the original goals delineated in the grant. The issues have changed slightly as problems we did not foresee were more problematic than the problems we foresaw. However, we have made significant progress and accomplished some of the objectives for the second year of the grant such as measuring attenuation and scatter.

Our results suggest that a linear model is appropriate to calculate the elasticity from transverse vibration of materials at low frequencies. Attenuation of the displacement is negligible throughout the sample and there is no measurable vibration perpendicular to the induced vibration. Therefore, this method should provide a good way to measure elasticity simply and accurately because the linear model allows the elasticity to be calculated directly without elaborate analysis.

We are developing methods of generating larger displacements in the MR to increase the sensitivity of the method.

**References:**

- 1 E.A. Sickles, S.H. Ominsky, R.A. Sollitto, H.B. Galvin, D.L. Monticciolo: "Medical Audit of a Rapid-Throughput Mammography Screening Practice: Methodology and Results of 27,114 Examinations," *Radiology* 1990; 175:323-327.
- 2 D.B. Kopans: "Breast Imaging and the Standard of Care for the Symptomatic Patient." *Radiology* 1993, 187:608-11.
- 3 R.E. Bird: "Professional Quality Assurance for Mammography Screening Programs (letter)." *Radiology* 1990; 177:587.
- 4 R.E. Bird, T.W. Wallace, B.C. Yankaskas: "Analysis of Cancers Missed at Screening Mammography," *Radiology* 1992; 184:613-617.
- 5 P.D. Britton, R.A. Couleden: "The Use of Doppler Ultrasound in the Diagnosis of Breast Cancer," *Clinical Radiology* 1990, 42(6):399-401.
- 6 P.N. Burns, M. Halliwell, P.N.T. Wells, A.J. Webb: "Ultrasonic Doppler Studies of the Breast." *Ultrasound in Med. Biol.* 1982;8:127-143.
- 7 F.W. Flickinger, J.D. Allison, R.M. Sherry, J.C. Wright, "Differentiation of benign from malignant breast masses by time-intensity evaluation of contrast enhanced MRI." *Magnetic Resonance Imaging* 1993; 11(5):617-20.
- 8 S.E. Harms, D.P. Flamig, "MR imaging of the breast." *Journal of magnetic resonance imaging* 1993;3(1):277-83.
- 9 T.J. Turkat, B.D. Klein, R.L. Polan, R.H. Richman: "Dynamic MR mammography: a technique for potentially reducing the biopsy rate for benign breast disease." *Journal of magnetic resonance imaging JMRI* 1994 Jul-Aug;4(4):563-8.
- 10 I. Khalkhali, I. Mena, E. Jouanne, L. Diggles, R. Venegas, J. Block, K. Alle, S. Klein: "Prone scintimammography in patients with suspicion of carcinoma of the breast." *Journal of the American College of Surgeons* 1994, May;178(5):491-7.
- 11 G. Piperno, E.H. Frei, M. Moshitzky: "Breast Cancer Screening by Impedance Measurements," *Frontiers Med. Biol. Engng* 1990, 2(2):111-117.
- 12 C.J. Baines, A.B. Miller, A.A. Bassett: "Physical examination. Its role as a single screening modality in the Canadian National Breast Screening Study." *Cancer; Diagnosis, treatment, research* 1989 May 1;63(9):1816-22.
- 13 D.P. Winchester: "Physical examination of the breast." *Cancer; Diagnosis, treatment, research* 1992 Apr 1;69(7 Suppl):1947-9.
- 14 S.W. Fletcher, W. Black, R. Harris, B.K. Rimer, S. Shapiro: "Report of the International Workspop for Breast Cancer," *Journal of the National Cancer Institute* 1993, 85(20):1644-1656.
- 15 A.B. Miller, C.J. Baines, T. To, C. Wall: "Canadian National Breast Screening Study: 2. Breast cancer detection and death rates among women aged 50 to 59 years." *Canadian Medical Association, Canadian Medical Association Journal* 1992 Nov 15;147(10):1477-88.
- 16 R.S. Foster Jr., J.K. Worden, M.C. Costanza, L.J. Solomon: "Clinical breast examination and breast self-examination. Past and present

- effect on breast cancer survival." *Cancer; Diagnosis, treatment, research* 1992 Apr 1;69(7 Suppl):1992-8.
- 17 C. Byrne, C.R. Smart, K.C. Chu, W.H. Hartmann: "Survival advantage differences by age. Evaluation of the extended follow-up of the Breast Cancer Detection Demonstration Project." *Cancer; Diagnosis, treatment, research* 1994, Jul 1;74(1 Suppl):301-10.
  - 18 I. Cespedes, J. Ophir, H. Ponnekanti, N. Maklad: "Elastography: Elasticity Imaging Using Ultrasound with Application to Muscle and Breast in vivo," *Ultrasonic Imaging* 1993, 15(2):73-88.
  - 19 J. Ophir, I. Cespedes, H. Ponnekanti, Y. Yazdi, X. Li: "Elastography: A Quantitative Method for Imaging the Elasticity of Biological Tissues," *Ultrasonic Imaging* 1991, 13:111-134.
  - 20 K.J. Parker, R.M. Lerner: "Sonoelasticity of organs: shear waves ring a bell." *Journal of Ultrasound in Medicine* 1992 Aug;11(8):387-92.
  - 21 F. Lee Jr, J.P. Bronson, R.M. Lerner, K.J. Parker, S.R. Huang, D.J. Roach: "Sonoelasticity imaging: results in in vitro tissue specimens." *Radiology* 1991 Oct;181(1):237-9.
  - 22 K.J. Parker, S.R. Huang, R.A. Musulin, R.M. Lerner: "Tissue response to mechanical vibrations for 'sonoelasticity imaging'." *Ultrasound Med Biol* 1990;16(3):241-6.
  - 23 R.M. Lerner, S.R. Huang, K.J. Parker: " 'Sonoelasticity' images derived from ultrasound signals in mechanically vibrated tissues." *Ultrasound Med Biol* 1990;16(3):231-9.
  - 24 Y.Yamakoshi, J. Sato, T. Sato: "Ultrasonic imaging of internal vibration of soft tissue under forced vibration." *IEEE Trans. Ultrason. Ferroelec. Frequency Contr.* 1990; 37:45-53.
  - 25 M.A. Hadley, D.J. Rubens, S.K. Alam, L. Goa, R.D. Mayer, K.J. Parker: "Sonoelasticity Imaging of Prostate Cancer." *RSNA Abstracts* 1993.
  - 26 A.L. Scherzinger, R.A. Belgam, P.L. Carson, C.R. Meyer, J.V. Sutherland, F.L. Bookstein, T.M. Silver: "Assessment of Ultrasonic Computed Tomography in Symptomatic Breast Patients by Discriminant Analysis." *Ultrasound Med Biol* 15(1):21-8, 1989.
  - 27 R. Muthupillai, D.J. Lomas, P.J. Rossman, J.F. Greenleaf, A. Manduca, and R.L. Ehman. *Science* 269, 1854, 1995.
  - 28 D.B. Plewes, G. Poole, M. Leitch, and S.N. Urchuk: "MR Assessment of Viscoelastic Properties of Tissue through Propagation of Transient Strain Waves." *Proceedings of the ISMRM Abstract* 476.
  - 29 R. Muthupillai, P.J. Rossman, J.F. Greenleaf, S.J. Riederer, and R.L. Ehman: "MR Imaging of Acoustic Strain Waves: Initial in Vivo Results." *Proceedings of the ISMRM Abstract* 475.
  - 30 YC Fung, SQ Liu: "Determination of the mechanical properties of the different layers of blood vessels in vivo." *Proceedings of the National Academy of Sciences*, 1995 Mar 14;92(6):2169-73.
  - 31 CO Olsen, DD Glower, KL Lee, PA McHale, JS Rankin: "Diastolic anisotropic properties of the left ventricle in the conscious dog." *Circulation research*, 1991 Sep;69(3):765-78.

- 32 P Zioupos, JC Barbenel, J Fisher: "Anisotropic elasticity and strength of glutaraldehyde fixed bovine pericardium for use in pericardial bioprosthetic valves." *Journal of biomedical materials research*, 1994 Jan;28(1):49-57.
- 33 T Farquhar, PR Dawson, PA Torzilli: "A microstructural model for the anisotropic drained stiffness of articular cartilage." *Journal of biomechanical engineering*, 1990 Nov;112(4):414-25.
- 34 JM Lee, DW Courtman DR Boughner: "The glutaraldehyde-stabilized porcine aortic valve xenograft. I. Tensile viscoelastic properties of the freest leaflet material" *Journal of Biomedical Materials Research*, 1984, 18:61-77.
- 35 KD Paulsen and H Jiang: "Spatially-varying optical property reconstructions using a finite element diffusion equation approximation," *Medical Physics* 22: 691-701, 1995.