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TITLE: Is the Regenerative Capacity of the Mammary Gland  
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## INTRODUCTION:

**SUBJECT:** Because transplanted mammary epithelial cells can regenerate typical ductal and alveolar structures in the host mammary fat pad, the existence of a mammary stem cell population has been postulated (1). Using a progesterone receptor (PR) knockout (PRKO) mouse model, we demonstrated that functional PR is essential for pregnancy-induced mammary epithelial ductal proliferation and lobuloalveolar differentiation but not for immediate post pubertal ductal morphogenesis (2). Furthermore, recent transgenic studies have demonstrated that targeted overexpression of either the A or B isoforms of the PR to the mammary gland results in excessive ductal branching for the PR-A transgenic and extensive alveolar growth for the PR-B transgenic (3,4). Collectively, these observations implied that P-signaling pathways may specify the fate of a mammary epithelial cell to become either a ductal or alveolar cell type in the adult. Based on the above, we hypothesized that mammary-specific stem cells are present as a subgroup within those epithelial cells that express both ER and PR.

**PURPOSE AND SCOPE:** To determine whether stem cells represent a subgroup of cells within the PR expressing cell population (PR<sup>+</sup>), a recently generated PR-LacZ mouse model in combination with fluorescent-activated cell sorting (FACS) was utilized to separate lacZ<sup>+</sup> (or PR<sup>+</sup>) from lacZ<sup>-</sup> (or PR<sup>-</sup>) mammary cells. The final stage of this proposal was to evaluate the regenerative capacity of both isolated cell populations through transplantation into host mammary fat pads devoid of an endogenous epithelial cellular compartment. Our hypothesis predicts that, unlike the PR<sup>-</sup> population, PR<sup>+</sup> cells would regenerate the mammary ductal tree.

## BODY:

Murine mammary epithelial cells were isolated from 16-week-old nulliparous PR-LacZ females according to the procedures described in (5). Panel A in Figure 1 shows a typical lacZ stained mammary gland whole mount from these mice; panel B represents a higher magnification of panel A revealing the nonuniform organization of epithelial cells that express the PR. Panel C depicts a section through a ductal structure revealing that PR expression is limited to the luminal epithelial component of the gland; see black arrow. Because the success of this "CONCEPT" proposal is predicated upon isolating viable PR<sup>+</sup> cells that still exhibit lacZ expression in culture, whole mammary cell cultures derived from the PR-lacZ mouse were initially evaluated for lacZ expression. Panel D demonstrates that cellular lacZ expression is maintained in culture and furthermore that this expression is preserved following multiple passaging of the cells (see white arrows).

Exploiting lacZ encoded  $\beta$ -galactosidase activity within PR<sup>+</sup> epithelial cells, FACS analysis was employed to isolate PR<sup>+</sup> from PR<sup>-</sup> mammary epithelial cells. Briefly, from 5 mice (both inguinal glands/mouse with lymph nodes removed), epithelial cells were isolated and hypotonically permeabilized with the following staining solution: (in 100  $\mu$ l) 1X PBS containing 4% (v/v) fetal calf serum, 10 mM HEPES, pH 7.2 and the fluorogenic substrate: 2mM fluorescein di- $\beta$ -D-digalactopyranoside (FDG; (F-1179) Molecular Probes, Inc.) and incubated at 37°C for 1 minute. The incubation was terminated by incubating on ice for 5 min before being diluted to 1 ml with the above staining solution. Due to the severity of the loading method, cells were rapidly pelleted by centrifugation

and resuspended in 2ml of staining solution containing propidium iodide (5ug/ml) to later identify those cells that did not survive the procedure.

Using the Flow Cytometry Core Facility at Baylor College of Medicine, cells were sorted using an argon laser (488nm) to excite the two fluorochromes: propidium iodide (to detect dead cells) and fluorescein (to detect lacZ<sup>+</sup> cells) and collected at band paths 610nm and 525 nm respectively. Panels E (wild type mice: negative control) and F (PR-LacZ mice) show a typical two-dimensional contour plot of scattered cells according to the fluorescence intensities of propidium iodide (PI) (quadrants M1 and M2-dead cells) and the final hydrolyzed product of FDG: fluorescein (quadrant M4); quadrant M3 denotes viable cells that are lacZ<sup>-</sup>. Panel E, representing wild type, shows that approximately 80% of the cellular population were viable; 15% of the cellular population did not survive the FDG loading procedure, whereas ~4% of the total cell population exhibited background fluorescence. In the case of PR-lacZ mice, ~56% of the total cell population were viable PR<sup>-</sup> mammary cells; approximately 19% of the cell population did not survive the treatment protocol whereas ~25% of the total cell population exhibited fluorescein-derived fluorescence. As indicated in Panel F, a region representing maximum viability and cell number were selected (gated) in quadrants M3 and M4 for subsequent cellular-transplantation into epithelial deficient mammary fat pads of host animals. Panels G and H represent cytopsin preparations from quadrants M3 and M4 respectively, shown in panel F. Due to cellular "clumping", panel H demonstrates that a pure lacZ<sup>+</sup> cellular population was not obtained, usually 70-80% this cellular population was LacZ<sup>+</sup>. Although lacZ<sup>-</sup> and lacZ<sup>+</sup> (70% enriched) cell populations were transplanted into donor fat-pads, mice harboring these recombinant glands, including most of the PR-lacZ colony, were lost to Tropical Storm "Allison" in June 2001; we are currently rebuilding the colony with a view to completing this experiment in the future.

## **KEY RESEARCH ACCOMPLISHMENTS:**

Exploiting a novel PR-lacZ mouse in combination with FACS analysis, we were able to isolate a significantly enriched mammary epithelial cellular population that expresses the PR. In addition to providing cellular material for the studies proposed in this CONCEPT AWARD, future studies may utilize these cell populations in conjunction with high density DNA microarray technology to detect signaling pathways downstream of the PR.

## **REPORTABLE OUTCOMES:**

The data described herein will be presented as preliminary data for an IDEA AWARD in 2002, sponsored by the Department of Defense.

## **CONCLUSIONS:**

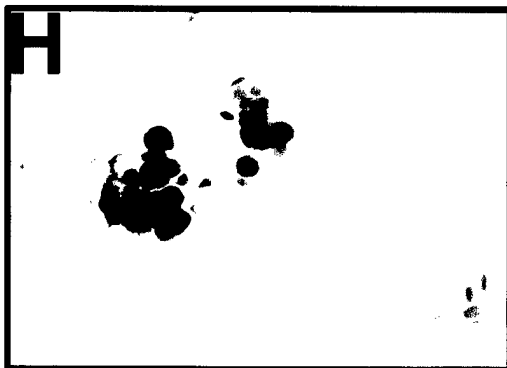
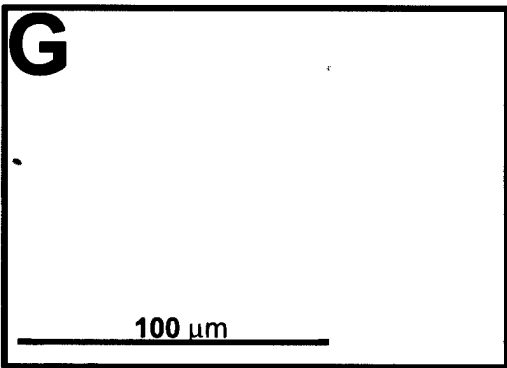
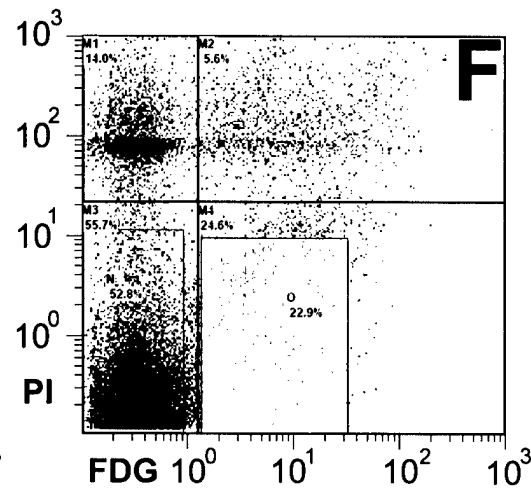
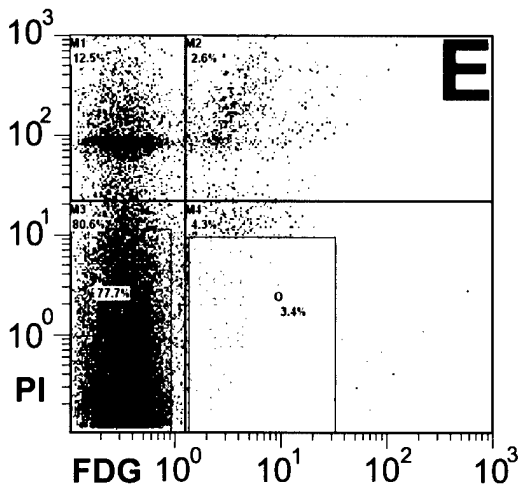
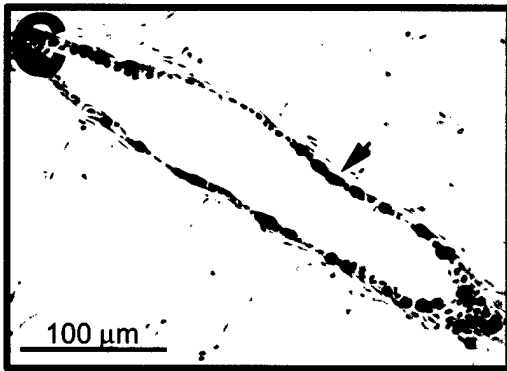
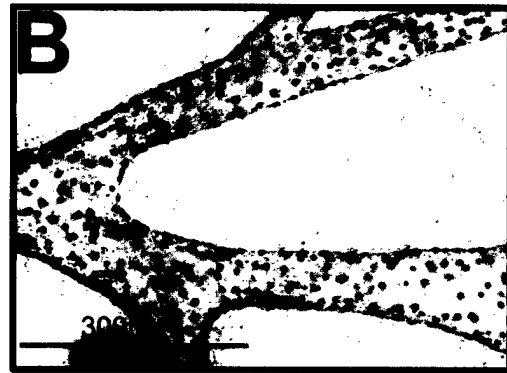
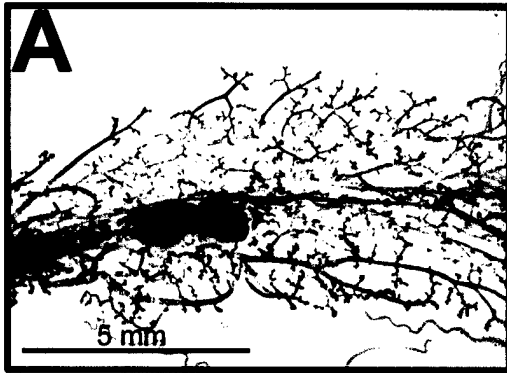
Due to extenuating circumstances (described in the **BODY**), it was not possible to address the final goal of this proposal, namely to evaluate the regenerative capacity of the individual PR-lacZ<sup>-</sup> and PR-lacZ<sup>+</sup> cell populations; however, the studies described herein provide support that the original CONCEPT is feasible. Finally, the targeted insertion of the autofluorescent green fluorescent protein into the PR gene may provide

an even more effective separation of PR<sup>+</sup> from PR<sup>-</sup> cells than obtained with the PR-lacZ mouse; this mouse is currently being generated.

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# APPENDIX



**FIGURE 1. FACS analysis of the PR-lacZ mammary gland.**

Panels A and B represent whole mounts of lacZ stained mammary glands from adult virgin mice; note the non-uniform expression pattern for PR in the main ducts (Panel B). Panel C represents a transverse section through a typical duct shown in (B); PR expression is limited to the luminal epithelial cellular component. Panel D represents lacZ staining of cultured mammary cells (epithelial, myoepithelial, and fibroblast cells) isolated from the PR-lacZ mammary gland; lacZ expression was maintained through a number of passaging steps. Panels E (wild type mice) and F (PR-lacZ mouse) represent two dimensional contour plots of scattered viable and nonviable PR<sup>-</sup> and PR<sup>+</sup> lacZ mammary epithelial cells. Panels G and H show lacZ stained mammary epithelial cells from the quadrants M3 and M4 shown in Panel F respectively; note the significant enrichment of PR<sup>+</sup> cells in Panel H.