

AD \_\_\_\_\_

Award Number: DAMD17-03-1-0768

TITLE: Effect of Depleting Tumor-Associated Macrophages on  
Breast Cancer Growth and Response to Chemotherapy

PRINCIPAL INVESTIGATOR: Min-Fu Tsan, M.D., Ph.D.

CONTRACTING ORGANIZATION: Institute for Clinical Research Inc.  
Washington, DC 20422-9745

REPORT DATE: October 2004

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;  
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

20050415 129

# REPORT DOCUMENTATION PAGE

Form Approved  
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE October 2004	3. REPORT TYPE AND DATES COVERED Annual (15 Sep 2003 - 14 Sep 2004)	
4. TITLE AND SUBTITLE Effect of Depleting Tumor-Associated Macrophages on Breast Cancer Growth and Response to Chemotherapy			5. FUNDING NUMBERS DAMD17-03-1-0768	
6. AUTHOR(S) Min-Fu Tsan, M.D., Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Institute for Clinical Research Inc. Washington, DC 20422-9745  E-Mail: min-fu.tsan2@med.va.gov			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited			12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200 Words)  Tumor-associated macrophages (TAM) may comprise up to 50% of the tumor mass in breast cancer and are capable of producing estrogen and angiogenic cytokines that regulate the growth and angiogenesis of breast cancer. The goal of this project is to determine whether depletion of TAM, achieved by intra-tumoral injection of liposome-encapsulated dichloromethylene diphosphonate (clodronate), can slow tumor growth and improve the outcome of breast cancer treatment. Preliminary studies so far have revealed that daily intra-tumoral injection of liposome-encapsulated clodronate had no effect on the growth of subcutaneous breast cancer (4T1) model in mice. Whether liposome-encapsulated clodronate depletes tumor-associated macrophages in this model is currently under investigation.				
14. SUBJECT TERMS Macrophages, Breast Cancer, Chemotherapy			15. NUMBER OF PAGES 7	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)  
Prescribed by ANSI Std. Z39-18  
298-102

## Table of Contents

<b>Cover.....</b>	<b>1</b>
<b>SF 298.....</b>	<b>2</b>
<b>Table of Content.....</b>	<b>3</b>
<b>Introduction.....</b>	<b>4</b>
<b>Body.....</b>	<b>4- 5</b>
<b>Key Research Accomplishments.....</b>	<b>6</b>
<b>Reportable Outcomes.....</b>	<b>6</b>
<b>Conclusions.....</b>	<b>6</b>
<b>References.....</b>	<b>6-7</b>
<b>Appendices.....</b>	<b>7</b>

## INTRODUCTION

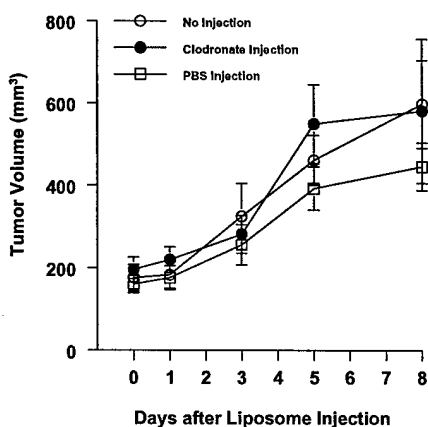
Tumor-associated macrophages (TAM) are one major component of tumor stroma. In breast cancer, TAM may comprise up to 50% of the tumor mass (1) and are capable of producing estrogen and angiogenic cytokines such as vascular endothelial growth factor and tumor necrosis factor alpha, that regulate the growth and angiogenesis of breast cancer (1-3). There is a positive correlation between high vascular grade and increased macrophage index, and a strong relationship between increased macrophage counts and reduced relapse-free survival as well as reduced overall survival in patients with breast cancers (3). In addition, the accumulation of TAM with thymidine phosphorylase is a potent prognostic indicator of early relapse in primary breast cancer (4). Despite the potential importance of TAM, current treatments of breast cancer have been primarily directed at breast cancer cells. Depletion of TAM may significantly improve the outcome of breast cancer therapy. The purpose of this study is to investigate whether depletion of TAM can slow breast tumor growth and improve the outcome of breast cancer treatment. Depletion of breast cancer TAM will be achieved using intra-tumoral injection of liposome-encapsulated dichloromethylene diphosphonate (clodronate), a specific and potent macrophage-depleting agent (5-8). Using a well established murine 4T1 breast cancer model (9), our specific objectives are: 1) To determine whether intra-tumoral injection of liposome-encapsulated clodronate can deplete TAM, 2) To determine the effect of depleting TAM on the tumor growth and angiogenesis, and 3) To determine the effect of depleting TAM on the outcome of chemotherapy.

## BODY

Two tasks are defined in the study: Task 1, Establish optimum conditions for depleting TAM using liposome encapsulated clodronate; and Task 2, Determine effects of macrophage depletion on breast cancer growth and angiogenesis, and the outcome of chemotherapy. Shortly after the funding of this project in September 2003, there was an unexpected turnover of the laboratory personnel, as a result the research in this project did not get started until after April 2004. A request for a no cost, one-year extension of the project was granted in September 2004. Thus, this report represents the first annual report, rather than the final report.

There are 4 stages proposed to accomplish Task 1: a, Establish the culture of 4T1 breast cancer cells and breast cancer model in BALBc mice; b, Prepare liposome-encapsulated clodronate; c, Treat (intra-tumoral injection) tumor-bearing mice with liposome-encapsulated clodronate; and d, Process tissue for immunohistochemical staining of macrophage-specific marker and quantify macrophages using Chalkley point counting method.

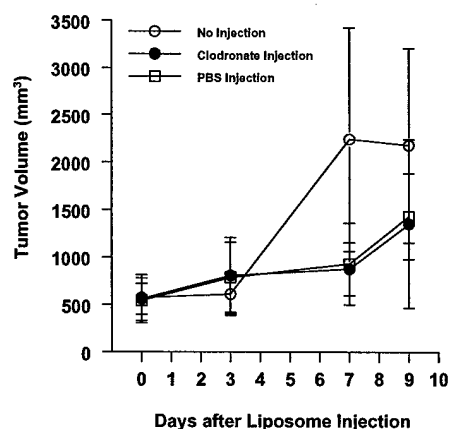
Two experiments have been carried out to accomplish Task 1. In the first experiment, 45 female mice at 12 weeks of age were obtained and quarantined for 2 week before sc injection of  $10^5$  cells in 100  $\mu$ l. The body weight of the animals was recorded every 2-3 days. Tumors became visible around 8 days after 4T1 cell inoculation. The tumor size was measured with a caliper every 2-3 days, and the tumor volume was estimated using the formula  $(L \times W^2)/2$  (10-12). Two weeks after the inoculation of 4T1 cells, tumors were well established in 40 mice. The animals were divided into 3 groups: a group with no injection as control, a group with injection of 10  $\mu$ l liposome-encapsulated clodronate, and a group with injection of 10  $\mu$ l liposome-encapsulated PBS (empty liposome) into the tumors. At intervals of 1, 3, 5, and 8 days after liposome injection, tumors were removed, fixed in buffered formalin for immuno-staining with macrophage-specific marker to quantify the number of macrophages. Results of tumor volume measurements revealed no significant difference between control and clodronate-injected group that was treated with 10  $\mu$ l liposome-encapsulated clodronate (Figure 1).



**Figure 1. Tumor growth after injection of Clodronate-encapsulated liposome.** 40 tumor-bearing mice were divided into 3 groups and treated with no injection, clodronate injection, PBS injection, respectively. Day 0 marks the day of injection. Tumor volume was determined as described in the text. Numbers of animals in each group at each data collection day are shown in **Table 1**. Results are presented with mean  $\pm$  S.D.

**Table 1. Numbers of animals in each group**

	n				
	Day 0	Day 1	Day 3	Day 5	Day 8
No Injection	14	14	10	7	4
Clodronate Injection	13	13	10	7	4
PBS Injection	13	13	8	5	2



**Figure 2. Tumor growth after injection of Clodronate-encapsulated liposome.** 28 tumor-bearing mice were divided into 3 groups and treated with no injection, clodronate injection, PBS injection, respectively. Day 0 marks the day of injection. Tumor volume was determined as described in the text. Numbers of animals in each group at each data collection day are shown in **Table 2**. Results are presented with mean  $\pm$  S.D.

**Table 2. Numbers of animals in each group**

	n			
	Day 0	Day 3	Day 7	Day 9
No Injection	9	9	6	3
Clodronate Injection	9	9	6	3
PBS Injection	10	10	7	4

In the second experiment, a total of 28 mice were obtained and processed as in the first experiment except that 30  $\mu$ l (15  $\mu$ l at 2 sites) of liposome-encapsulated clodronate or empty liposome was injected intra-tumorally. Similar to the first experiment, tumors became visible around a week and well established 2 weeks after inoculation. At intervals of 3, 7, and 9 days after liposome injection, tumors were removed, fixed in buffered formalin for immuno-staining with macrophage-specific marker to quantify the number of macrophages. Results of tumor volume measurement showed no significant difference between clodronate-injected group and PBS-injected group (Figure 2). The difference between injected and non-injected group had no statistical significance.

The tumor tissues collected from these two experiments were processed for immunochemical staining. Paraffin-embedded tumor tissues were sectioned into 4-6 $\mu$ m slices. The slides were deparaffinated, trypsinized or heated to activate the antigen, and processed in a DAKO Autostainer with an antibody to mouse macrophage-surface marker F4/80, HRP-conjugated secondary antibody, and DAB substrate. The slides were then counter stained with hematoxylin. So far, we have not been able to identify macrophages in these tumor sections. Currently, we are using a different antibody to macrophage markers (anti-Mac-1) attempting to improve macrophage detection.

## KEY RESEARCH ACCOMPLISHMENTS

- Established breast cancer model in BALBc mice.
- Treated tumor-bearing mice with liposome-encapsulated clodronate.
- Process tissue for immunohistochemical staining with macrophage-specific marker.

## REPORTABLE OUTCOMES

None.

## CONCLUSIONS

The purpose of this project is to determine whether in vivo depletion of tumor-associated macrophages (TAM) is feasible and if it is feasible, what is the effect of TAM-depletion on the growth of breast cancer and its response to cancer chemotherapy. Using a murine subcutaneous breast cancer model, we have demonstrated that intra-tumoral injection of up to 30  $\mu$ l of liposome-encapsulated clodronate had no effect on the growth of breast cancer. Whether it has any effect on the depletion of TAM is currently under investigation.

## REFERENCES

1. Mor G, Yue W, Santen RJ, Gutierrez L, Eliza M, Berstein LM, Harada N, Wang J, Lysiak J, Diano S, Naftolin F. Macrophages, estrogen and the microenvironment of breast cancer. *J Steroid Biochem Mol Biol* 67: 403-411, 1998.
2. Lewis CE, Leek R, Harris A, McGee JO'D. Cytokine regulation of angiogenesis in breast cancer: the role of tumor-associated macrophages. *J Leukoc Biol* 57: 747-751, 1995.
3. Leek RD, Lewis CE, Whitehouse R, Greenall M, Clarke J, Harris AL. Association of macrophage infiltration with angiogenesis and prognosis in invasive breast carcinoma. *Cancer Res* 56: 4625-4629, 1996.
4. Toi M, Ueno T, Matsumoto H, Saji H, Funata N, Koike K, Tominaga T. 1999. Significance of thymidine phosphorylase as a marker of protumor monocytes in breast cancer. *Clin Cancer Res* 5: 1131-1137.
5. Van Rooijen N. The liposome-mediated macrophage 'suicide' technique. *J Immunol methods* 124: 1-6, 1989.
6. Van Rooijen N, Classen E. In vivo elimination of macrophages in spleen and liver, using liposome-encapsulated drugs: methods and applications. In: *Liposomes as drug carriers*, edited by G. Gregoriadis. London: Wiley, pp 131-143, 1988.
7. Berg JT, Lee ST, Thepen T, Lee CY, Tsan MF. 1993. Depletion of alveolar macrophages by liposome-encapsulated dichloromethylene diphosphonate. *J Appl Physiol* 74: 2812-2819.
8. Bottino R, Fernandez LA, Ricordi C, Lehmann R, Tsan MF, Oliver R, Inverardi L. Transplantation of allogenic islets of Langerhans in the rat liver. Effect of macrophage depletion on graft survival and microenvironment activation. *Diabetes* 47: 316-323, 1998.
9. Bove K, Lincoln DW, Tsan MF. 2002. Effect of resveratrol on growth of 4T1 breast cancer cells in vitro and in vivo. *Biochem Biophys Res Commun* 291: 1001-1005.
10. Wang XY, Kazim L, Repasky EA, and Subjeck JR. Characterization of Heat Shock Protein 110 and Glucose-Regulated Protein 170 as Cancer Vaccines and the Effect of Fever-Range Hyperthermia on Vaccine Activity. *J Immunol* 166: 490-497, 2001.
11. Huang X, Wong MK, Yi H, Watkins S, Laird AD, Wolf SF, and Gorelik E. Combined Therapy of Local and Metastatic 4T1 Breast Tumor in Mice Using SU6668, an Inhibitor of Angiogenic Receptor Tyrosine Kinases, and the Immunostimulator B7.2-IgG Fusion Protein. *Can Res* 62: 5727-5735, 2002.

12. Manjili MH, Wang X-Y, Chen X, Martin T, Repasky EA, Henderson R, and Subjeck JR. HSP110-HER2/neu Chaperone Complex Vaccine Induces Protective Immunity Against Spontaneous Mammary Tumors in HER-2/neu Transgenic Mice. *J Immunol* 171: 4054-4061, 2003.
13. Fox SB, Leek RD, Weekes MP, Whitehouse RM, Gatter KC, Harris AL. Quantitation and prognostic value of breast cancer angiogenesis: Comparison of microvessel density, Chalkley count, and computer image analysis. *J Pathol* 177: 275-283, 1995.

## **APPENDICES**

None