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Award Number: W81XWH-05-1-0248

TITLE: Developing a beta-Lapachone Prodrug for Therapy Against NQO1-Overexpressing Breast Cancers

PRINCIPAL INVESTIGATOR: Kathryn E. Reinicke, B.S.
David A. Boothman, Ph.D.

CONTRACTING ORGANIZATION: Case Western Reserve University
Cleveland, Ohio 44106

REPORT DATE: March 2006

TYPE OF REPORT: Annual Summary

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

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REPORT DOCUMENTATION PAGE				<i>Form Approved</i> OMB No. 0704-0188	
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1. REPORT DATE (DD-MM-YYYY) March 2006		2. REPORT TYPE Annual Summary		3. DATES COVERED (From - To) 1 Mar 05 – 28 Feb 06	
4. TITLE AND SUBTITLE Developing a beta-Lapachone Prodrug for Therapy Against NQO1-Overexpressing Breast Cancers				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-05-1-0248	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Kathryn E. Reinicke, B.S. David A. Boothman, Ph.D. E-Mail: ker5@po.cwru.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Case Western Reserve University Cleveland, Ohio 44106				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT We report the development of mono(arylimino) derivatives of β -lap as potential prodrugs (Reinicke et al. 2005). They undergo hydrolytic conversion to β -lap at rates dependent on the electron-withdrawing strength of their substituent groups and pH of the diluent. Once converted, they caused NQO1-dependent, μ -calpain-mediated cell death in human cancer cells identical to that caused by β -lap. NQO1 reduces β -lap to an unstable hydroquinone that rapidly undergoes a two-step oxidation back to the parent compound. The resultant futile cycling generates reactive oxygen species (ROS). We hypothesize that $O_2^{\bullet-}$ is generated by the NQO1-mediated metabolism of β -lap. Comet assay analyses and measurements of disulfide glutathione levels suggest that NQO1-dependent β -lap metabolism produces DNA damaging oxidative stress. Our data indicate that ROS formation is upstream of subsequent endoplasmic reticulum calcium store release, suggesting that β -lap-mediated ROS formation may be necessary, but not sufficient, to induce cell death.					
15. SUBJECT TERMS beta-lapachone, breast cancer, prodrug, therapeutics, NQO1/DT-diaphorase, pharmacokinetics					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			USAMRMC
			UU	18	19b. TELEPHONE NUMBER (include area code)

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Introduction

β -Lapachone (β -Lap, 3,4-dihydro-2,2-dimethyl-2H-naphtho[1,2b]pyran-5,6-dione) is a naturally occurring *o*-naphthoquinone present in the bark of the South American Lapacho tree. β -Lap induces a novel, caspase-independent, apoptotic pathway at relatively low concentrations in human breast cancer cell lines that is dependent on NAD(P)H:quinone oxidoreductase (NQO1) expression. NQO1 reduces β -lap to an unstable hydroquinone that presumably undergoes a two-step oxidation back to the parent compound. The resultant futile cycling generates reactive oxygen species (ROS) (Pink et al. 2000) whose role in this apoptotic pathway is currently unknown. We hypothesize that superoxide ($O_2^{\bullet-}$) is generated by the NQO1-mediated metabolism of β -lap, and that this cycling is taking place in a membranous cellular compartment(s) that is unavailable to water soluble free radical scavengers (e.g., N-acetyl-L-cysteine, NAC) or is so robust as to be unaffected by general water soluble free radical scavengers. In addition, NQO1-mediated β -lap futile cycling is thought to deplete the levels of NAD(P)H and ATP, thus causing Ca^{2+} release from the endoplasmic reticulum (ER) leading to cell death.

We report the development of mono(arylimino) derivatives of β -lap as potential prodrugs (Reinicke et al. 2005). They undergo hydrolytic conversion to β -lap at rates dependent on the electron-withdrawing strength of their substituent groups and pH of the diluent. Once converted, they caused NQO1-dependent, μ -calpain-mediated cell death in human cancer cells identical to that caused by β -lap.

β -Lap, a novel chemotherapeutic agent and known radiosensitizer, is able to target various breast cancer cells over normal cells due to their 5- to 20-fold elevated levels of NQO1 which “bioactivates” β -lap. A deficiency or inhibition of NQO1 renders cells resistant to β -lap. Furthermore, NQO1 is damage-inducible in mammalian cells after ionizing radiation or ultraviolet radiation and its levels can increase in response to tamoxifen, giving β -lap the potential to augment current breast cancer therapies. Therefore, β -lap has great potential for the treatment of specific breast cancers with elevated NQO1 levels.

Body

1. Development of β -lapachone prodrugs for therapy against human cancer cells with elevated NAD(P)H:quinone oxidoreductase 1 levels

We have developed mono(arylimino) derivatives of β -lap as potential prodrugs (Reinicke et al. 2005). These derivatives are relatively non-toxic and not substrates for NQO1 when initially diluted in water. In solution, however, they undergo hydrolytic conversion to β -lap at rates dependent on the electron-withdrawing strength of their substituent groups and pH of the diluent. NQO1 enzyme assays, UV-vis spectrophotometry, HPLC-ESI-MS, and NMR analyses confirmed and monitored conversion of each derivative to β -lap. Once converted, β -lap derivatives caused NQO1-dependent, μ -calpain-mediated cell death in human cancer cells identical to that caused by β -lap. Interestingly, co-administration of NAC prevented derivative-induced cytotoxicity, but did not affect β -lap lethality. NMR analyses indicated that prevention of β -lap derivative cytotoxicity was the result of direct modification of these derivatives by NAC, preventing their conversion to β -lap.

The use of β -lap, β -lap mono(arylimino) prodrug derivatives, or more specifically a derivative converted in a tumor-specific manner (i.e., in the acidic local environment of the tumor tissue), should reduce normal tissue toxicity while eliciting tumor-selective cell killing by NQO1 bioactivation. A more detailed description can be found in the appendix (Reinicke et al. 2005).

2. Reactive oxygen species generation is necessary, but not sufficient, for β -lapachone-mediated apoptosis

Comet assay analyses and measurements of disulfide glutathione (GSSG) levels suggest that NQO1-dependent β -lap metabolism produces DNA damaging oxidative stress. Increases in GSSG can be blocked by inhibiting NQO1 with dicoumarol or by using high concentrations of NAC. Thirty-minute exposures of β -lap in NQO1-containing cells are sufficient to induce comet tail formation similar to hydrogen peroxide treated controls. Our data indicate that ROS formation is upstream of subsequent ER calcium store release as pretreatment with the intracellular calcium chelator BAPTA-AM is able to block toxicity, but not GSSG accumulation. BAPTA-AM partially abrogates comet tail formation. This suggests that β -lap-mediated ROS formation may be necessary, but not sufficient, to induce cell death.

Key Accomplishments

- Mono(arylimino) derivatives of β -lap are potentially viable prodrugs
- Mono(arylimino) derivatives of β -lap are relatively non-toxic and are not substrates for NQO1 when initially diluted in water
- Mono(arylimino) derivatives of β -lap undergo hydrolytic conversion to the parent β -lap compound; the rate of this conversion is dependent upon the electron-withdrawing strength of the substituent group and the pH of the aqueous solution
- Once converted to the parent β -lap compound, the mono(arylimino) derivatives are just as toxic as β -lap
- NAC interacts with β -lap mono(arylimino) derivatives and renders them incapable of converting to the parent β -lap compound
- β -Lap causes NQO1-mediated oxygen consumption, oxidative stress and cell death
- Inhibition of NQO1 by dicoumarol prevents oxygen consumption, generation of oxidative stress and cell death after β -lap exposure
- High dose NAC is able to partially block cell death during/after β -lap exposure
- β -Lap exposure causes DNA damage
- BAPTA-AM pretreatment does not abrogate β -lap-induced oxidative stress, but partially abrogates β -lap-mediated DNA damage

Reportable Outcomes

- Publications:
 - Reinicke, K.E., Bey, E.A., Bentle, M.S., Pink, J.J., Ingalls, S.T., Hoppel, C.L., Misico, R.I., Arzac, G.M., Burton, G., Bornmann, W.G., Sutton, D., Gao, J., Boothman, D.A. Development of β -lapachone prodrugs for

therapy against human cancer cells with elevated NAD(P)H:quinone oxidoreductase 1 levels. *Clinical Cancer Research*, 2005; 11(8): 3055-64.

- Associate Member, American Association for Cancer Research
- University poster presentations:
 - Investigating the role of reactive oxygen species in β -lapachone-mediated cell death. Kathryn E. Reinicke¹, Marie Varnes², Doug Spitz, Ph.D³, John J. Pink, Ph.D², and David A. Boothman, Ph.D²
¹Department of Biochemistry, ²Department of Radiation Oncology, Case Western Reserve University, Cleveland, OH 44106; ³Department of Radiation Oncology, University of Iowa, Iowa City, IA 52242.
Biochemistry Department Retreat, Case Western Reserve University, October 2005
 - Reactive oxygen species generation is necessary, but not sufficient, for β -lapachone-mediated apoptosis. Kathryn E. Reinicke^{1,2}, John J. Pink^{1,3}, William G. Bornmann⁵, Douglas R. Spitz⁶, and David A. Boothman^{1,3,4,8}
¹Laboratory of Molecular Stress Responses, Departments of ²Biochemistry, ³Radiation Oncology, and ⁴Pharmacology, Case Western Reserve University, Cleveland, OH 44106-4942; ⁵Department of Molecular Imaging, MD Anderson University, Houston, TX 77030; and ⁶Free Radical and Radiation Biology Program, Department of Radiation Oncology, University of Iowa, Iowa City, IA 52242.
Annual Graduate Student Symposium, Case Western Reserve University, April 2005
- National meeting poster presentation:
 - Investigating the role of reactive oxygen species in β -lapachone-mediated cell death. Kathryn E. Reinicke¹, Marie Varnes², Doug Spitz, Ph.D³, John J. Pink, Ph.D², and David A. Boothman, Ph.D²
¹Department of Biochemistry, ²Department of Radiation Oncology, Case Western Reserve University, Cleveland, OH 44106; ³Department of Radiation Oncology, University of Iowa, Iowa City, IA 52242.
Department of Defense Era of Hope Breast Cancer Research Conference, June 2005

Conclusions

The rationale underlying development of this class of mono(arylimino) prodrugs for the delivery of β -lap for chemo- and/or radiotherapy is based on delayed, pH-dependent activation (Yao et al. 1994; Page 1996; Hoffman et al. 1997; Solovskii et al. 2002; Nishi and Jayakrishnan 2004). After systemic administration of a potent β -lap mono(arylimino) prodrug, the chemotherapeutic agent must traverse the body through zones of neutral pH via the blood stream and lymphatic circulation, eventually reaching its target, acidic solid tumor tissue (Loeffler et al. 1991; Engin et al. 1995; Stubbs et al. 2000). We believe that mono(arylimino) derivatives of β -lap would cause minimal damage to normal tissues with neutral pH microenvironments during circulation. Ideally, these β -lap mono(arylimino) prodrugs will remain in inactive forms during the initial systemic distribution, and will not become active until they successfully reach the breast tumor, which commonly has an acidic microenvironment (Loeffler et al. 1991; Engin et

al. 1995; Stubbs et al. 2000). The resultant β -lap produced is then 'bioactivated' by NQO1 (Pink et al. 2000), an enzyme found in human tissues at low levels (Jaiswal et al. 1988; Chen et al. 1995; Rauth et al. 1997), but expressed at high levels in many human breast cancers (Malkinson et al. 1992; Belinsky and Jaiswal 1993; Joseph et al. 1994; Marin et al. 1997).

Collectively, the data presented in this paper strongly suggest that some form of β -lap mono(arylimino) derivative can be developed for use as a prodrug. The observation that the electron-withdrawing group on the arylimino substitution group, as well as the pH of the surrounding aqueous environment, can directly influence conversion of the prodrugs to β -lap should be invaluable in developing these active chemotherapeutic agents. Analyses of this class of β -lap prodrugs could lead to potent, breast tumor-selective, antitumor agents.

The motivation behind investigating the role of ROS in β -lap-mediated cell death is to obtain a better understanding of this drug's mechanism of action for the purpose of improved application. Due to the low level of expression of NQO1 in human tissues, there is a possibility that systemic administration of this drug could lead to damaging effects in normal tissues. Co-administration of a chemical free radical scavenger may be encouraged to dampen any free radical-mediated damage to non-target tissues. Ideally, a low dose of scavenger may be sufficient to protect normal tissues, while having little to no effect on β -lap's ability to kill breast tumor tissue.

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Development of β -Lapachone Prodrugs for Therapy Against Human Cancer Cells with Elevated NAD(P)H:Quinone Oxidoreductase 1 Levels

Kathryn E. Reinicke,^{1,2} Erik A. Bey,^{1,3} Melissa S. Bentle,^{1,4} John J. Pink,^{1,3} Stephen T. Ingalls,^{4,6} Charles L. Hoppel,^{4,6,7} Rosana I. Misico,⁸ Gisela M. Arzac,⁸ Gerardo Burton,⁸ William G. Bornmann,⁹ Damon Sutton,⁵ Jinming Gao,⁵ and David A. Boothman^{1,3,4}

Abstract β -Lapachone, an *o*-naphthoquinone, induces a novel caspase- and p53-independent apoptotic pathway dependent on NAD(P)H:quinone oxidoreductase 1 (NQO1). NQO1 reduces β -lapachone to an unstable hydroquinone that rapidly undergoes a two-step oxidation back to the parent compound, perpetuating a futile redox cycle. A deficiency or inhibition of NQO1 rendered cells resistant to β -lapachone. Thus, β -lapachone has great potential for the treatment of specific cancers with elevated NQO1 levels (e.g., breast, non-small cell lung, pancreatic, colon, and prostate cancers). We report the development of mono(arylimino) derivatives of β -lapachone as potential prodrugs. These derivatives are relatively nontoxic and not substrates for NQO1 when initially diluted in water. In solution, however, they undergo hydrolytic conversion to β -lapachone at rates dependent on the electron-withdrawing strength of their substituent groups and pH of the diluent. NQO1 enzyme assays, UV-visible spectrophotometry, high-performance liquid chromatography-electrospray ionization-mass spectrometry, and nuclear magnetic resonance analyses confirmed and monitored conversion of each derivative to β -lapachone. Once converted, β -lapachone derivatives caused NQO1-dependent, μ -calpain-mediated cell death in human cancer cells identical to that caused by β -lapachone. Interestingly, coadministration of *N*-acetyl-L-cysteine prevented derivative-induced cytotoxicity but did not affect β -lapachone lethality. Nuclear magnetic resonance analyses indicated that prevention of β -lapachone derivative cytotoxicity was the result of direct modification of these derivatives by *N*-acetyl-L-cysteine, preventing their conversion to β -lapachone. The use of β -lapachone mono(arylimino) prodrug derivatives, or more specifically a derivative converted in a tumor-specific manner (i.e., in the acidic local environment of the tumor tissue), should reduce normal tissue toxicity while eliciting tumor-selective cell killing by NQO1 bioactivation.

β -Lapachone (3,4-dihydro-2,2-dimethyl-2H-naphtho[1,2-*b*]pyran-5,6-dione) has demonstrated significant antitumor activity against sarcoma 180 cells *in vitro* (1). Further studies show that

β -lapachone elicits significant antitumor activity against Yoshida sarcoma, Walker 256 carcinoma cells (2), and Rauscher leukemia (3) in mice. Data showing activity against a variety of tumors have revitalized interest in this drug (1, 3–5).

Previous studies from our laboratory revealed that β -lapachone was “bioactivated” by the two-electron oxidoreductase, NAD(P)H:quinone oxidoreductase 1 (NQO1; EC 1.6.99.2; ref. 6). Structural similarities between β -lapachone and other members of the naphthoquinone family, such as menadione (vitamin K3, 2-methyl-1,4-naphthoquinone), suggested that NQO1 may be involved in the activation or detoxification of this drug (7–11). Furthermore, the ionizing radiation-inducible properties of NQO1 (i.e., xip3) were consistent with this compound’s ability to radiosensitize various human cancer cells (12). We subsequently discovered that NQO1 was the primary intracellular determinant of β -lapachone cytotoxicity (6, 13) and noted that all NQO1⁺ cell lines examined were more sensitive to β -lapachone. Furthermore, the cytotoxicity of β -lapachone was prevented by coadministration of dicoumarol (a NQO1 inhibitor), and cells lacking NQO1 were inherently more resistant than NQO1-expressing cells. In contrast, NQO1-expressing cancer cells were resistant to menadione, and coadministration of dicoumarol potentiated menadione lethality.

Authors’ Affiliations: ¹Laboratory of Molecular Stress Responses and Departments of ²Biochemistry, ³Radiation Oncology, ⁴Pharmacology, ⁵Biomedical Engineering, and ⁶Medicine, Case Western Reserve University; ⁷Medical Research Service, Louis Stokes Veterans Affairs Medical Center, Cleveland, Ohio; ⁸Departamento de Química Orgánica, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Buenos Aires, Argentina; and ⁹Department of Molecular Imaging, M.D. Anderson University, Houston, Texas

Received 10/27/04; revised 1/18/05; accepted 1/27/05.

Grant support: NIH/National Cancer Institute grant R01 CA-92250 (D.A. Boothman), NIH grant 5T32 CA59366 and Department of Defense grant BC043335 (K.E. Reinicke), National Research Council of Argentina (CONICET-Argentina; G. Burton), and University of Buenos Aires fellowship (G.M. Arzac).

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Note: Supplementary data for this article are available at Clinical Cancer Research Online (<http://clincancerres.aacrjournals.org/>).

Requests for reprints: David A. Boothman, Laboratory of Molecular Stress Responses, Department of Radiation Oncology and Pharmacology, Case Comprehensive Cancer Center, 2103 Cornell Avenue, WRB-3531 West, Cleveland, OH 44106-7285. Phone: 216-368-0840; Fax: 216-368-8919; E-mail: DAB30@case.edu.

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We showed previously that β -lapachone killed cells through a unique apoptotic pathway. Cell death caused by many chemotherapeutic agents typically involves activation of cysteine proteases (e.g., caspases; refs. 14, 15). Apoptosis induced by β -lapachone in NQO1⁺ human breast or prostate cancer cell lines was unique, causing a pattern of poly(ADP-ribose) polymerase (PARP) and p53 proteolysis distinct from that caused by other agents (e.g., staurosporine) that caused cell death through activation of caspases (6, 13, 16–18). We noted concomitant atypical PARP (yielding an ~60-kDa fragment) and p53 (yielding an ~40-kDa fragment) cleavage in β -lapachone-treated NQO1⁺ cells caused by activation of the neutral Ca²⁺-dependent cysteine protease, μ -calpain (18–20). Certain aspects of β -lapachone cytotoxicity were unique (e.g., atypical PARP and p53 proteolysis), whereas other aspects conformed to classic apoptosis (e.g., lamin B proteolysis, DNA condensation, terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling-positive cells, sub-G₀-G₁ cells, and trypan blue exclusion; ref. 13).

Although β -lapachone is ideally suited for use as an antitumor agent because it targets an enzyme elevated in various tumors (21–24), concerns over normal tissue toxicity remain. These concerns could be avoided by the development of a β -lapachone prodrug that would favor conversion to β -lapachone in the microenvironment of the tumor and less so in normal tissues. β -Lapachone holds great promise as an antitumor agent because it (a) is “bioactivated” by NQO1 through a futile redox cycle, and NQO1 levels are elevated in numerous cancers (21–24); (b) kills independent of cell cycle, caspase, pRb, or p53 status; and (c) synergistically kills cells in combination with chemotherapeutic agents as well as ionizing radiation (25). Ideally, β -lapachone prodrugs would become active in a tumor-selective or spontaneous manner with identical or more potent antitumor activity than β -lapachone. The active form of such prodrugs would kill by the same NQO1-dependent manner but avoid normal tissue toxicity depending on their mechanism and rate of activation.

We report that biologically inactive mono(arylimino) derivatives of β -lapachone seem to act as Schiff's bases and are converted to β -lapachone through a spontaneous hydrolytic reaction. The rates of hydrolysis of β -lapachone derivatives varied as a function of the strength of the electron-withdrawing substituent groups in the *para* position of the mono(arylimino) leaving group as well as the pH of the solution (Fig. 1, R group; Supplementary Fig. 1). Thus, mono(arylimino) derivatives may represent usable nontoxic, inactive, and pH-sensitive (26–30) precursors to β -lapachone.

Materials and Methods

Reagents. β -Lapachone was synthesized by us, confirmed by nuclear magnetic resonance (NMR), dissolved in DMSO at 40 mmol/L, and concentrations verified by spectrophotometry (13, 16). β -Lapachone stock solutions were stored at -80°C until used. β -Lapachone mono(arylimino) derivatives [phenylimino lapachone (PIL), 2,2-dimethyl-(Z)-6-phenylimino-3,4,5,6-tetrahydro-2H-naphtho[1,2-b]oxin-5-one; *p*-methylphenylimino lapachone (MePIL), 2,2-dimethyl-(Z)-6-(4-methyl-phenylimino)-3,4,5,6-tetrahydro-2H-naphtho[1,2-b]oxin-5-one; *p*-methoxyphenylimino lapachone (MPIL), 2,2-dimethyl-(Z)-6-(4-methoxyphenylimino)-3,4,5,6-tetrahydro-2H-naphtho[1,2-b]oxin-5-one; and *p*-nitrophenylimino lapachone (NPIL), 2,2-dimethyl-(Z)-6-(4-nitrophenylimino)-3,4,5,6-tetrahydro-2H-naphtho[1,2-

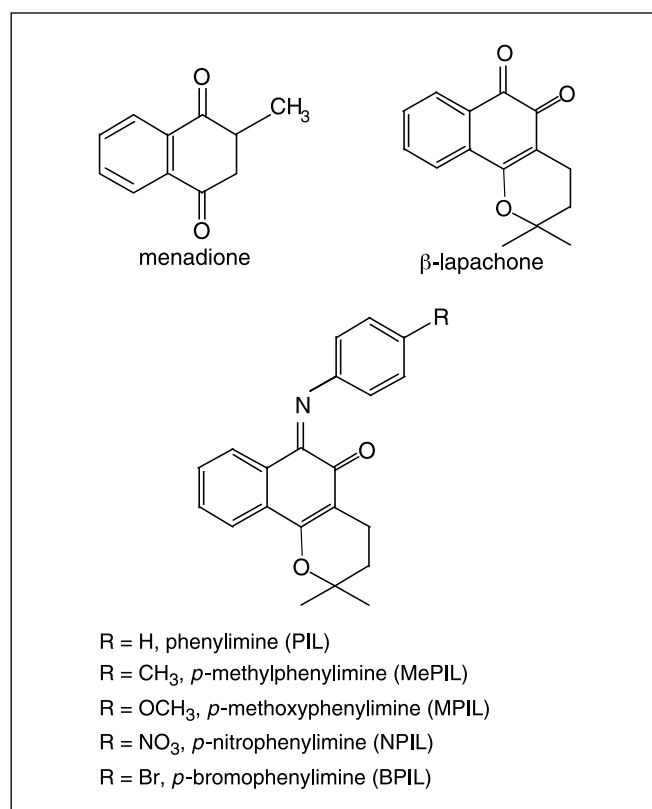


Fig. 1. Structures of β -lapachone derivatives. Structures for menadione, β -lapachone, PIL, MePIL, MPIL, NPIL, and BPIL.

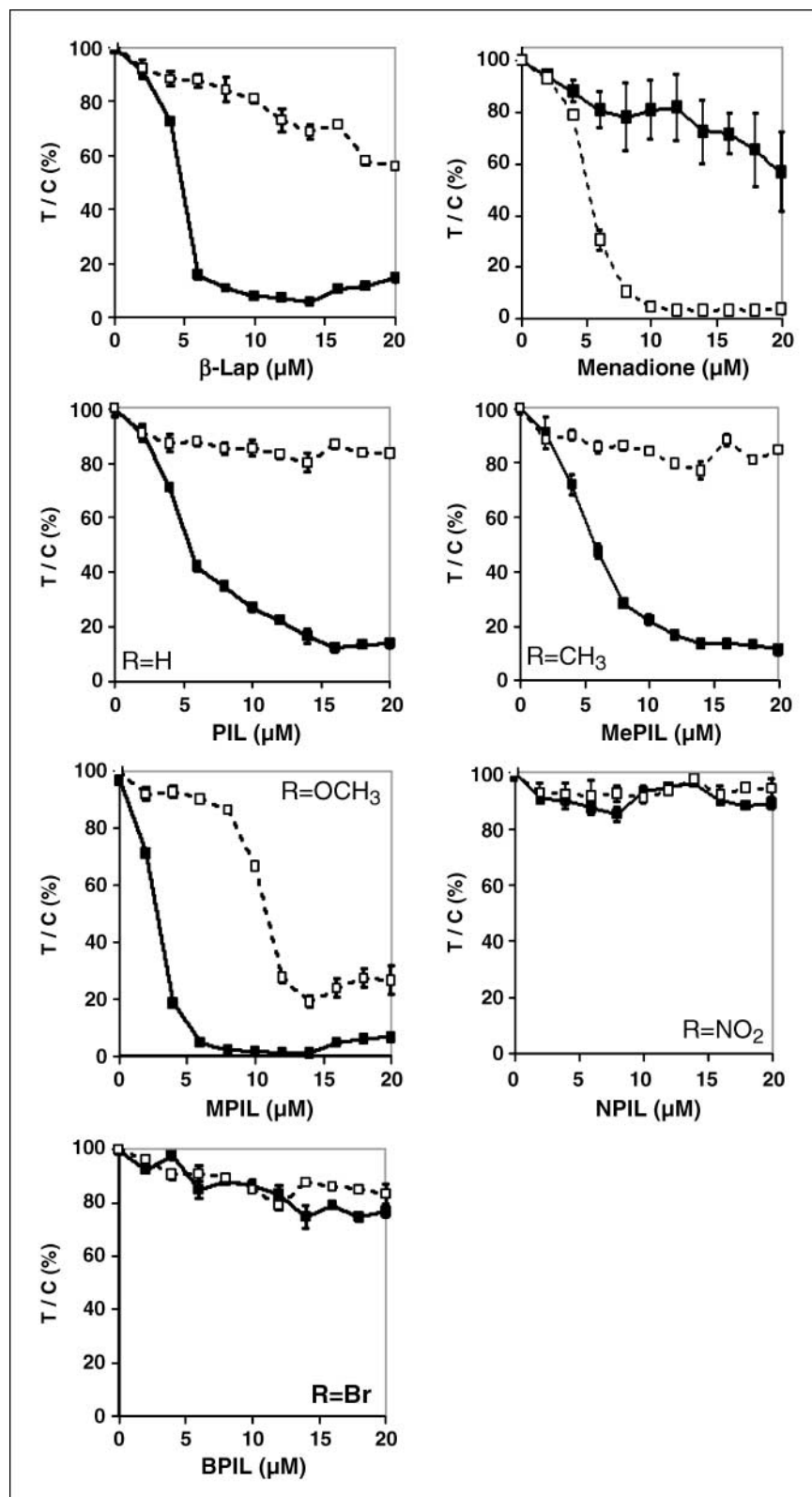
b]oxin-5-one] were synthesized as described (31). *p*-Bromophenylimino lapachone (BPIL), 2,2-dimethyl-(Z)-6-(4-bromophenylimino)-3,4,5,6-tetrahydro-2H-naphtho[1,2-b]oxin-5-one, was prepared from 4-bromoaniline and β -lapachone as described previously (31), with 90% yield as a brown solid that was recrystallized from cyclohexane. ¹H NMR (CDCl₃, 200 MHz): 8.19 (dd, *J* = 7.1 and 1.7, 1H, H-7), 7.83 (dd, *J* = 7.1 and 1.7, 1H, H-10), 7.50 (m, 2H, H-8, 9), 7.43 (d, *J* = 8.8, 2H, H-2'), 6.62 (d, *J* = 8.8, 2H, H-3'), 2.41 (t, *J* = 6.7, 2 H, H-4), 1.79 (t, *J* = 6.7, 2 H, H-3), 1.44 (s, 6 H, 2-CH₃); ¹³C NMR (CDCl₃, 50 MHz): 177.5 (C-5), 161.6 (C-10b), 152.1 (C-1'), 150.9 (C-6), 132.4 (C-10a), 131.7 (C-3'), 131.3 (C-8), 130.2 (C-9), 130.0 (C-6a), 127.0 (C-7), 123.3 (C-10), 117.6 (C-2'), 115.5 (C-4'), 111.9 (C-4a), 78.5 (C-2), 31.7 (C-3), 26.7 (2-CH₃), 16.0 (C-4). Seventy EV electron impact ionization mass spectroscopy, *m/z* (%): 397 (M⁺ + 2, 43), 395 (M⁺, 43), 316 (48), 260 (93), 232 (100), 203 (35), 76 (32), 41 (37). Aryliminoquinones were dissolved in DMSO at 50 mmol/L. Menadione was obtained from Sigma Chemical Co. (St. Louis, MO), dissolved in DMSO at 100 mmol/L, and stored at -80°C . Drugs [β -lapachone, menadione, and β -lapachone mono(arylimino) derivatives] were added to medium at a 1:1,000 dilution immediately before administration to cells; control cells received $\leq 0.1\%$ DMSO for vehicle alone treatment. Dicoumarol (Sigma Chemical) was used as described (6). *N*-acetyl-L-cysteine (NAC; Sigma Chemical) was dissolved in water, pH adjusted to 7.4 using NaOH, and stored at 4°C . Acetonitrile (high-performance liquid chromatography grade), methanol (high-performance liquid chromatography grade), and DMSO (ACS Certified Spectranalyzed grade) were purchased from Fisher Scientific (Pittsburgh, PA). Ammonium formate was purchased from Sigma Chemical. Medipure-grade liquid nitrogen was obtained in high pressure, gas-withdrawal containers from Praxair, Inc. (Cleveland, OH) for use as the sheath and auxiliary gas in the electrospray ion source of the mass spectrometer.

Argon (5.0 ultrahigh purity grade) was obtained from Praxair for use as the collision gas during tandem mass spectrometric experiments.

Cell culture. NQO1-deficient (NQO1⁻) MDA-MB-468 and MDA-MB-231 cells were obtained from the American Type Culture Collection (Manassas, VA) and stably transfected with the cytomegalovirus-driven

human NQO1 cDNA in the pcDNA3 mammalian expression vector as described (6). NQO1⁺ 468-NQ3 and 231-NQ6 as well as NQO1⁻ vector-alone 468-Vec4 and 231-NQ2 clones were isolated; 231-NQ2 did not express NQO1 protein or enzyme levels. MCF-7:WS8 (MCF-7) cells were obtained and grown as described (6). A549 cells were obtained

Fig. 2. β -Lapachone (β -Lap) mono(arylimino) derivatives resemble β -lapachone in mediating NQO1-dependent cytotoxicity. NQO1⁺ 231-NQ6 cells were seeded into 48-well plates (1×10^4 cells per well) and allowed to attach overnight. Medium containing various drug concentrations (β -lapachone, menadione, PIL, MePIL, MPIL, NPIL, or BPIL as indicated) either alone (■ and solid lines) or coadministered with 40 μ M/L dicoumarol (□ and dashed lines) was added for 4 hours. Medium was then removed, fresh drug-free medium was added, and cells were monitored for changes in relative survival. Relative DNA content per well was determined by Hoechst dye fluorescence, and relative growth (T/C) was plotted as described in Materials and Methods. Points, mean of three independent wells from three independent experiments; bars, SE.



from Dr. Steven Dubinett (University of California-Los Angeles Medical Center, Los Angeles, CA) and grown as described below. PC-3 cells were obtained and grown as described (32). All cells were grown in RPMI 1640 supplemented with 200 µg/mL geneticin (except for parental MCF-7 and A549 cells), 5% fetal bovine serum, 2 mmol/L L-glutamine, 100 units/mL penicillin, and 100 mg/mL streptomycin (13, 16) in a 37°C humidified incubator with a 5% CO₂, 95% air atmosphere (13, 16). All cells were routinely monitored for, and were free of, *Mycoplasma* contamination (6).

Cell growth assays. Cell growth was assessed using DNA content (6, 13). Briefly, PC-3 (seeded at 5×10^3 per well), A549, MCF-7, 231-NQ6, 231-NQ2 (seeded at 1×10^4 per well), 468-Vec4, or 468-NQ3 (seeded at 4×10^4 per well) cells were plated in 48-well plates and allowed to attach overnight. Cells were then treated with 4-hour pulses of drug [0–20 µmol/L β-lapachone, menadione, or β-lapachone mono(arylimino) derivatives; Fig. 2] alone or in conjunction with 40 µmol/L dicoumarol or 5.0 mmol/L NAC; 40 µmol/L dicoumarol prevented NQO1 activity in all cells examined, and NAC was used to scavenge reactive oxygen and nitrogen species (33). Medium was removed, fresh medium was added, and cells were grown for at least 7 days. DNA content was determined by fluorescence using a Perkin-Elmer HTS 7000 Bio-Assay Reader microtiter plate reader (Boston, MA) (6), and data were expressed as relative growth [treated versus control (T/C)] from three wells per treatment. Each experiment was repeated at least thrice, and data were expressed as mean ± SE. Prior analyses with β-lapachone and other β-lapachone analogues showed that cytotoxicity monitored by growth assays correlated directly with changes monitored by colony forming ability assays (6).

UV-visible spectrophotometric analyses. UV-visible spectrophotometric analyses of β-lapachone mono(arylimino) derivative hydrolysis in water were done in the presence or absence of 5 mmol/L NAC. Analyses of MPIL hydrolysis at varied pH were done in 50 mmol/L Tris-HCl buffer at pH 6, 7, or 8 and the ratio of peak absorbance at 258 and 289 nm was plotted over time as a measure of MPIL hydrolysis to β-lapachone. Analyses were made using a Beckman DU 640 spectrophotometer (Beckman Coulter, Fullerton, CA) scanning 200 to 500 nm wavelengths. Stock solutions (50 mmol/L) of each β-lapachone mono(arylimino) derivative dissolved in DMSO were diluted to 1 mL with water and subjected to UV-visible wavelength scans every 30 minutes for 4 hours. Scans were compared with β-lapachone.

NAD(P)H:quinone oxidoreductase 1 enzyme assays. S9 supernatants were prepared as described (6), aliquoted, and stored at –80°C before analysis. NQO1 enzymatic activities were measured using cytochrome *c* (practical grade; Sigma Chemical) in Tris-HCl buffer (50 mmol/L, pH 7.5; ref. 6). NADH (200 µmol/L) was the immediate electron donor, and β-lapachone, menadione, or β-lapachone mono(arylimino) derivatives (10 µmol/L) were intermediate electron acceptors as monitored using a Beckman DU 640 spectrophotometer. Activities attributed to NQO1 were levels inhibited by 10 µmol/L dicoumarol (34) and calculated as nmol cytochrome *c* reduced/min/µg protein based on initial rate of change in absorbance at 550 nm and an extinction coefficient for cytochrome *c* of 21.1 mmol/L/cm. Results are average enzyme activities for three separate cell extracts ± SD.

NADH recycling assays. NADH recycling assays were done using NQO1-containing S9 extracts from MCF-7 cells (6). NADH oxidation was monitored by changes in absorbance at 340 nm measured for 5 minutes with a Beckman DU 640 spectrophotometer. Protein concentrations were determined by Bradford assays (6). Enzyme activities were expressed as T/C where mol NADH reduced in 5 min/mol substrate [β-lapachone, menadione, or mono(arylimino) derivative] added per µg protein are compared with mol NADH reduced in 5 min/mol menadione added per µg protein.

Western blot analyses. Western blot analyses of DMSO control or drug-treated MCF-7 cells were done as described (13, 16, 18). Loading equivalence and transfer efficiency were monitored by Western blot analyses of proteins not altered in cells following β-lapachone exposure as described (13). Dilutions of 1:2,000 for α-PARP SC-8007 antibody (Santa Cruz Biotechnology, Santa Cruz, CA) and 1:2,000 for

α-p53 D01 antibody (Santa Cruz Biotechnology) were used to monitor apoptotic proteolyses of these proteins.

¹H nuclear magnetic resonance spectroscopy. For pure compound spectra, MPIL or NAC (1.0 mg) was dissolved in deuterated DMSO (DMSO-d₆) before acquiring NMR spectra. For 1:1 molar ratio MPIL:NAC spectra, MPIL (2.9 mol) was dissolved in DMSO-d₆, diluted into water containing 2.9 mol NAC, and incubated at room temperature for ≥4 hours. For MPIL hydrolysis analysis, MPIL (1 mg) was dissolved in DMSO-d₆, diluted into water, and allowed to incubate at room temperature for ≥4 hours. The water was evaporated to a volume of 7 mL. All samples were filtered (0.2 µm nylon syringe filter), freeze-dried in a Labconco Freezone 4.5 lyophilizer (Kansas City, MO), and redissolved in 1 mL DMSO-d₆ before acquiring NMR spectra. All ¹H NMR spectra were recorded on a 200 MHz Varian Gemini Fourier transform NMR spectrometer (Palo Alto, CA).

High-performance liquid chromatography-electrospray ionization-mass spectrometry measurement of β-lapachone derivative hydrolysis. β-Lapachone mono(arylimino) derivative stock solutions in DMSO ($\sim 3 \times 10^{-3}$ mol/L) were diluted 1:100 (v/v) in methanol. Methanolic working solutions were diluted immediately 1:100 (v/v) in water contained by autosampler vials. Aliquots (5 µL) were injected into the instrument at precisely measured intervals of ~1 hour for 30 hours. Chromatographic separations were done using reverse-phase Polaris C18-A (3 µm particle diameter) medium contained in a 0.46 cm i.d. × 5.0 cm liquid column (Ansyl Metachem, Lake Forest, CA). The chromatographic mobile phase consisted of aqueous ammonium formate (2.5×10^{-2} mol/L, pH not adjusted)/acetonitrile (30:70, v/v). Column temperature and mobile phase flow rate were 40°C and 0.5 mL/min, respectively. The mass spectrometer was operated with heated capillary and analyzer manifold temperatures of 350°C and 70°C, respectively. Ion source sheath gas pressure was 80 p.s.i.g., and the auxiliary gas flow rate was set to 40 dimensionless units. The instrument was operated in positive ion mode, the ion source electrospray potential was 3.5 kV, and the collision cell argon pressure was 2.0×10^{-5} mTorr. β-Lapachone and β-lapachone mono(arylimino) derivatives ionized readily as positively charged proton adducts and underwent a characteristic collision-induced loss of *m/z* 56 in the mass spectrometer. Transitions were monitored [precursor ion (*m/z*), product ion (*m/z*), and collision cell offset potential (V)]: β-lapachone (243, 187, –24), PIL (318, 262, –25), MePIL (332, 276, –25), MPIL (348, 292, –25), NPIL (363, 307, –25), and BPIL (396, 340, –25). Acquisition dwell time and product ion scan width were 0.1 second and *m/z* 1.0, respectively, for all monitored transitions. Chromatographic peaks were observed for β-lapachone, PIL, MePIL, MPIL, NPIL, and BPIL at 2.0, 5.1, 6.3, 4.6, 5.1, and 7.8 minutes, respectively. β-Lapachone mono(arylimino) derivative concentration versus time data sets was modeled as an irreversible first-order process according to the equation:

$$C(t)_{\text{prodrug}} = C(0)_{\text{prodrug}} \exp(-k_1 * t),$$

where *k*₁ is the first-order rate constant and *C*(*t*)_{prodrug} and *C*(0)_{prodrug} are the concentrations of the prodrug at time *t* and 0, respectively. The production of β-lapachone during β-lapachone mono(arylimino) derivative hydrolysis was modeled as an irreversible first-order process according to the equation:

$$C(t)_{\beta\text{-lapachone}} = C(0)_{\text{prodrug}} (1 - \exp(-k_2 * t)),$$

where *k*₂ is the first-order formation constant of β-lapachone and *C*(*t*)_{β-lapachone} is the concentration of β-lapachone at time *t*, respectively.

Results

Phenylimine lapachone, *p*-methylphenylimine lapachone, and methoxyphenylimine lapachone β-lapachone mono(arylimino) derivatives mimic β-lapachone cytotoxicity. Various mono(arylimino) substituted β-lapachone derivatives were synthesized

Table 1. Comparison of lethality of β -lapachone analogues in NQO1⁺ and NQO1[−] MDA-MB-231 breast cancer cells by LD₅₀ values (μ mol/L)

Compound	Compound alone		Compound + dicoumarol		Compound + NAC	
	NQO1 ⁺ *	NQO1 [−] †	NQO1 ⁺	NQO1 [−]	NQO1 ⁺	NQO1 [−]
β -Lapachone	6.6 $\pm$ 1.4 [‡]	14.2 $\pm$ 2.1	26 $\pm$ 0.5	30 $\pm$ 0.5	5.5 $\pm$ 0.4	15.3 $\pm$ 2.1
PIL	5.9 $\pm$ 1.3	16.7 $\pm$ 1.8	21 $\pm$ 0.5	29 $\pm$ 0.5	25 $\pm$ 0.5	29 $\pm$ 0.5
MePIL	5.7 $\pm$ 1.0	15.3 $\pm$ 2.3	30 $\pm$ 1.0	40 $\pm$ 0.5	15 $\pm$ 0.5	40 $\pm$ 0.5
MPIL	3.7 $\pm$ 0.5	9.7 $\pm$ 1.3	17.8 $\pm$ 1.9	18.0 $\pm$ 1.6	8.5 $\pm$ 0.4	15.3 $\pm$ 1.6
NPIL	>50 [§]	>50	>50	>50	>50	>50
BPIL	>50	>50	>50	>50	>50	>50

NOTE: Log-phase cells were treated with a 4-hour pulse of compound (with or without 40 μ mol/L dicoumarol or 5 mmol/L NAC). Compound medium was replaced with fresh medium and the cells were allowed to grow for 7 days. Relative DNA per well was determined by Hoechst fluorescence, and relative growth (T/C DNA) was plotted as described in Materials and Methods.

*NQO1⁺ MDA-MB-231 cells generated as described in Materials and Methods.

†NQO1[−] MDA-MB-231 cells generated as described in Materials and Methods.

‡Values were determined from relative growth assays done at least thrice in triplicate.

§Values of >50 μ mol/L LD₅₀ indicate <50% lethality at 50 μ mol/L drug, the upper limit of toxic doses used in assaying that drug.

(31) with leaving groups of different electron-withdrawing strengths (Fig. 1). Because NQO1 reduces quinones (in either *para* or *ortho* position; ref. 31), mono(arylimino) derivatives of β -lapachone were not expected to be substrates for NQO1, nor were they expected to kill NQO1⁺ human cancer cells as noted with β -lapachone (6). Relative growth assays using NQO1⁺ (e.g., 468-NQ3, 231-NQ6, MCF-7, A549, and PC-3 cells) compared with NQO1[−] (e.g., 468-Vec4, and 231-NQ2) cells were done for each β -lapachone derivative. Log-phase cells were exposed for 4 hours to various concentrations of β -lapachone alone, menadione alone, each β -lapachone mono(arylimino) derivative alone, or each compound in combination with 40 μ mol/L dicoumarol (Fig. 2). β -Lapachone mono(arylimino) derivatives PIL, MePIL, and MPIL were as selectively toxic to NQO1⁺ breast cancer cells as β -lapachone, with comparable LD₅₀ values of 5.9 $\pm$ 1.3, 5.7 $\pm$ 1.0, 3.7 $\pm$ 0.5, and 6.6 $\pm$ 1.4 μ mol/L, respectively (Fig. 2; Table 1). In contrast, NPIL and BPIL were significantly less toxic, with LD₅₀ values of >50 μ mol/L (Fig. 2; Table 1). Dicoumarol prevented PIL, MePIL, and MPIL cytotoxicities in a manner similar to that observed with β -lapachone in NQO1⁺ breast cancer cells, increasing LD₅₀ values to 21 $\pm$ 0.5, 30 $\pm$ 1.0, 17.8 $\pm$ 1.9, and 26 $\pm$ 0.5 μ mol/L, respectively (Table 1). MPIL was not as toxic as β -lapachone in NQO1⁺ lung cancer (A549) cells, with LD₅₀ values of 8.1 $\pm$ 0.2 and 5.3 $\pm$ 0.6 μ mol/L, respectively (Supplementary Table 1). MPIL was also not as toxic as β -lapachone in NQO1⁺ prostate cancer (PC-3) cells, with LD₅₀ values of 6.5 $\pm$ 0.3 and 3.5 $\pm$ 0.5 μ mol/L, respectively (Supplementary Table 1). Dicoumarol prevented MPIL cytotoxicities in both A549 and PC-3 cells, increasing LD₅₀ values to >50 μ mol/L (Supplementary Table 1). NQO1[−] 468 and 231 cells were significantly more resistant to all drugs examined (Table 1). Overall, these data suggested that β -lapachone mono(arylimino) derivatives may be converted to β -lapachone or β -lapachone-like compounds to kill cancer cells in an NQO1-dependent manner.

Evidence for conversion of β -lapachone mono(arylimino) derivatives to β -lapachone. Direct data supporting the hypothesis that β -lapachone mono(arylimino) derivatives were

converted to β -lapachone were then sought. Stock solutions of β -lapachone mono(arylimino) derivatives in DMSO were diluted in water (15:10,000) and subjected to UV-visible spectrophotometric wavelength scans of 200 to 500 nm every 30 minutes for 4 hours, representing the time of drug exposure used in cytotoxicity experiments (Fig. 2). The UV-visible spectra of β -lapachone mono(arylimino) derivatives PIL, MePIL, and MPIL significantly changed, ultimately resembling the spectra of β -lapachone (Supplementary Fig. 2A-C). In contrast, the β -lapachone spectra remained unaltered over the 4-hour period. In contrast, scans of NPIL and BPIL [less toxic derivatives (Fig. 2)] showed a time-dependent change in their spectra, yet the resulting spectra did not resemble the spectra of β -lapachone (Supplementary Fig. 2D and E), suggesting that these compounds did not convert to β -lapachone.

To determine the effect of pH on the hydrolysis of β -lapachone mono(arylimino) derivatives, a stock solution of MPIL in DMSO was diluted in 50 mmol/L Tris-HCl buffers of pH 6, 7, and 8 (5:1,000), and UV-visible spectra measured every 30 minutes for 4 hours. The spectrum of β -lapachone is characterized by a major peak at 258 nm and a minor peak at 289 nm, giving a signature ratio of 2.7. The spectrum of unhydrolyzed MPIL also has a major peak at 258 and a minor peak at 289 nm; however, the ratio of these two peaks is 1.16. As MPIL hydrolyzes to β -lapachone, the ratio increases from 1.16 to 2.7. For each different pH, the ratio of peak absorbance for MPIL at 258 and 289 nm (258/289 nm) was plotted over time as a measure of hydrolysis of MPIL to β -lapachone (Supplementary Fig. 1). MPIL was converted to β -lapachone more rapidly at pH 6 (filled circles) than at pH 7 (filled squares) or pH 8 (filled triangles), demonstrating that MPIL converted to β -lapachone more efficiently in acidic conditions. The ratio at pH 6 does not reach 2.7 within 4 hours, indicating that MPIL does not convert to β -lapachone completely within 4 hours.

Using high-performance liquid chromatography-electrospray ionization-mass spectrometry, the conversion in water of PIL, MePIL, MPIL, and NPIL to β -lapachone over time was monitored

(Supplementary Fig. 3; Supplementary Table 2). The rates of conversion of these β -lapachone derivatives from highest to lowest were MPIL > MePIL > PIL > NPIL, with rate constants of derivative loss ranging from -0.693 to -0.001 h^{-1} and rate constants of β -lapachone formation ranging from 0.221 to 0.034 h^{-1} . NMR spectra acquired before (Supplementary Fig. 4A) and after incubating MPIL with water for ≥ 4 hours (Supplementary Fig. 4C) confirmed disappearance of the mono(arylimino) derivative and a concomitant increase in β -lapachone formation. The appearance of the doublet at 7.9 ppm , as well as the loss of intensity of the 8.2 ppm doublet, suggests that MPIL was hydrolyzed to β -lapachone (Supplementary Fig. 4C). The rates of loss of parent compounds (Supplementary Table 2) correlated well with the relative cytotoxicities of the compounds (Fig. 2; Table 1; Supplementary Table 1). For example, the low rate of conversion of NPIL to β -lapachone was consistent with the lowered cytotoxicity of this set of compounds (e.g., NPIL and BPIL).

β -Lapachone mono(arylimino) derivatives become NAD(P)H: quinone oxidoreductase 1 substrates. Data from Fig. 2, Table 1, Supplementary Fig. 2, and Supplementary Table 1 suggest that β -lapachone mono(arylimino) derivatives can be converted to β -lapachone. Therefore, we did NQO1 enzyme activity assays after immediate ($t = 0$) or 1 or 4 hours of diluting each β -lapachone derivative in aqueous buffer. NQO1 activities were expected to reflect conversion of biologically inactive β -lapachone mono(arylimino) derivatives to β -lapachone. β -Lapachone and menadione were included as controls. S9 supernatants from MCF-7 cells were used as a source of NQO1 for enzymatic reactions as described in Materials and Methods. As expected at time 0, all β -lapachone mono(arylimino) derivatives examined were poor NQO1 substrates, reflected by low NQO1 activities (1.17 – $6.49\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$) compared with menadione ($21.23\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$) or β -lapachone ($13.34\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$; $t = 0$; Fig. 3A). However, after a 1-hour incubation in reaction buffer, PIL, MePIL, and MPIL became NQO1 substrates, resulting in NQO1 enzyme activities ranging from 9.48 to $13.49\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$ ($t = 1\text{ hour}$; Fig. 3A), similar to β -lapachone. In contrast, the relatively nontoxic derivatives, BPIL and NPIL, remained poor substrates for NQO1 (1.34 – $2.25\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$). After an incubation time of 4 hours, PIL, MePIL, and MPIL were better substrates for NQO1, eliciting approximately three to five times the NQO1 enzyme activity initially observed, rising to 14.7 to $18.6\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$ ($t = 4\text{ hours}$; Fig. 3A). NPIL elicited approximately twice the NQO1 enzyme activity ($2.9\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$), and BPIL elicited approximately thrice the NQO1 enzyme activity initially observed, rising to $4.6\text{ }\mu\text{mol cytochrome } c/\text{min}/\mu\text{g}$. Thus, biologically inactive β -lapachone mono(arylimino) derivatives were converted into NQO1 substrates at times that correlated with cytotoxic exposure times for each derivative in NQO1⁺ cells (Fig. 2) and hydrolytic rates determined by high-performance liquid chromatography-electrospray ionization-mass spectrometry (Supplementary Fig. 3; Supplementary Table 2).

When NQO1 metabolizes β -lapachone in the absence of an electron acceptor (e.g., cytochrome *c*), a futile cycle of NADH oxidation occurs (6). Up to 10 mol NADH may be oxidized per mol β -lapachone in 5 minutes (Fig. 3B). In

contrast, when menadione was used in the same reaction, ~ 1 mol NADH was oxidized per mol menadione in 5 minutes (Fig. 3B; ref. 35). When β -lapachone mono(arylimino) derivatives were examined immediately after dilution with water, these compounds were not efficient NQO1 substrates and therefore elicited minimal futile redox cycling in this enzymatic assay. Consistent with the finding that PIL, MePIL, and MPIL were converted to β -lapachone (a known NQO1 substrate; ref. 6), at 4 hours, these same compounds elicited significant NQO1 futile cycling; for example, 7.25- to 12.9-fold T/C were noted, equal to the futile cycling noted for β -lapachone (9.98-fold T/C). Consistent with the marginal cytotoxicity of NPIL and BPIL (Fig. 2; Table 1), these compounds elicited little or no NQO1-dependent recycling activities (2.37- to 2.73-fold T/C; Fig. 3B).

β -Lapachone mono(arylimino) derivatives induce atypical poly(ADP-ribose) polymerase and p53 cleavage in NAD(P)H: quinone oxidoreductase 1-positive MCF-7 cells. β -Lapachone-induced apoptosis was characterized by atypical proteolyses of PARP (resulting in an ~ 60 -kDa fragment; refs. 6, 16, 18, 32) and p53 (giving an ~ 40 -kDa fragment; ref. 18), indicative of μ -calpain-mediated cell death (Fig. 4A and C; ref. 18). NQO1⁺ MCF-7 cells were treated for 4 hours with either $5\text{ }\mu\text{mol/L}$ PIL, MePIL, or MPIL as well as $20\text{ }\mu\text{mol/L}$ NPIL or BPIL. Because NPIL and BPIL were relatively nontoxic at concentrations below $15\text{ }\mu\text{mol/L}$ (Fig. 2), a higher concentration of these compounds was used. PIL, MePIL, and MPIL induced atypical PARP (Fig. 4B; Supplementary Fig. 5A and B) and p53 (Fig. 4C and D) proteolyses. Proteolyses were abrogated by $40\text{ }\mu\text{mol/L}$ dicoumarol cotreatment as shown for β -lapachone (6) and were not observed in NQO1[−] 231 or 468 cells (data not shown). Unlike β -lapachone, PARP and p53 proteolyses induced by PIL, MePIL, or MPIL were blocked by cotreatment with 5 mmol/L NAC (Fig. 4; Supplementary Fig. 5), consistent with the protection afforded by NAC against PIL, MePIL, and MPIL cytotoxicities (Table 1). NPIL and BPIL did not induce atypical proteolyses of PARP or p53 (Fig. 4; Supplementary Fig. 5), consistent with their inability to induce cell death after a 4-hour exposure (Fig. 2; Table 1).

N-acetyl-L-cysteine prevents β -lapachone mono(arylimino) derivative conversion to β -lapachone and subsequent cytotoxicity. Coadministration of NAC did not affect the cytotoxicity of β -lapachone, suggesting that reactive oxygen species produced by this drug (36–38) may not play a role in its lethality or that the reactive oxygen species formed by treatment of NQO1⁺ cells with β -lapachone are compartmentalized (Table 1; Supplementary Table 1). Preliminary data suggest that NAC pretreatment and cotreatment of cells did not block reactive oxygen species formation induced by β -lapachone exposure.¹⁰ In contrast, cotreatments of NQO1⁺ breast cancer cells with 5 mmol/L NAC in combination with PIL, MePIL, or MPIL dramatically protected these cells (~ 3 -fold or greater) from lethality; LD₅₀ values increased to 25 ± 0.5 , 15 ± 0.5 , and $8.5 \pm 0.4\text{ }\mu\text{mol/L}$, respectively (Table 1). In NQO1[−] A549 and PC-3 cells, cotreatments with NAC also increased LD₅₀ values to 16.1 ± 0.2 and $11.3 \pm 0.4\text{ }\mu\text{mol/L}$, respectively (Supplementary Table 1). To determine if NAC was directly interacting with

¹⁰ K. E. Reinicke et al. Investigating the role of reactive oxygen species in β -lapachone-mediated cell death, unpublished data.

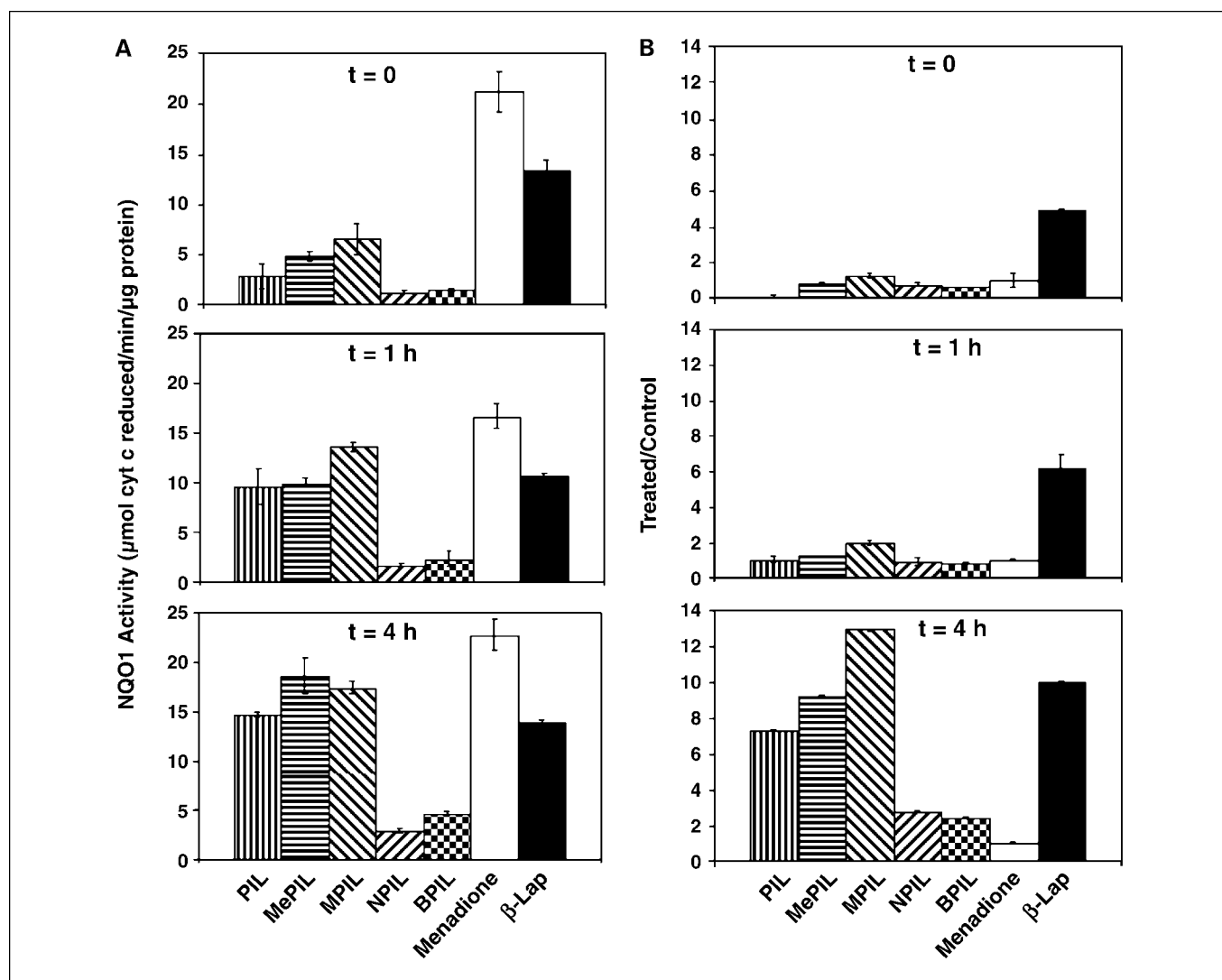


Fig. 3. β -Lapachone mono(arylimino) derivatives become NQO1 substrates and enter into futile cycling reactions of NADH oxidation similar to β -lapachone. **A**, NQO1 enzyme activity was assayed as described in Materials and Methods (1) after allowing the β -lapachone mono(arylimino) derivatives to solubilize in water for 0, 1, or 4 hours ($t = 0, 1$, or 4 hours). Enzyme activities were calculated as μ mol cytochrome c reduced/min/ μ g protein. Columns, average NQO1 enzyme activities for three separate cell extractions; bars, SD. **B**, futile cycling activity was assayed as described in Materials and Methods (6) after dissolving β -lapachone mono(arylimino) derivatives in water for 0, 1, or 4 hours ($t = 0, 1$, or 4 hours). NADH oxidation was monitored by changes in UV-visible absorbance at 340 nm. Results are expressed as T/C, where average mol NADH oxidized/mol substrate (compound)/5 min/ μ g protein $\pm$ SD are compared with mol NADH oxidized/mol menadione/5 min/ μ g protein $\pm$ SD. Averages were determined from experiments done thrice.

β -lapachone mono(arylimino) derivatives to prevent their conversion to β -lapachone, we collected UV-visible scans of PIL, MePIL, MPIL, and BPIL, with and without 5 mmol/L NAC, every 30 minutes for 4 hours (Supplementary Fig. 6). We observed that conversion of PIL, MePIL, MPIL, and BPIL to β -lapachone was blocked by NAC (Supplementary Fig. 6B-E), whereas scans of β -lapachone with or without 5 mmol/L NAC remained unchanged (Supplementary Fig. 6A). NMR analyses confirmed the direct interaction and derivitization of MPIL with NAC (Supplementary Fig. 7). A 1:1 molar ratio mixture of MPIL and NAC incubated at room temperature in aqueous solution showed the appearance of doublets in the 6.5 to 7.5 ppm range, suggesting chemical modification of MPIL near the imine moiety of the molecule (Supplementary Fig. 7C). The appearance of a multiplet at 4.35 ppm is indicative of participation of NAC in the chemical modification (Supplementary Fig. 7C).

Discussion

Our data support the hypothesis that mono(arylimino) derivatives of β -lapachone could be potentially useful prodrugs for therapy against human cancers with elevated NQO1 levels. Five derivatives with unique chemical structures (Fig. 1; ref. 31) were directly compared with β -lapachone and menadione for apoptotic responses and lethality. Three derivatives (PIL, MePIL, and MPIL) exhibited cytotoxicities equivalent to, or slightly greater than, β -lapachone in NQO1⁺ cells (Fig. 2; Table 1; Supplementary Table 1). Our results indicated that these biologically inactive β -lapachone derivatives hydrolyzed to β -lapachone (Fig. 5; ref. 39) in a manner consistent with Schiff's base hydrolysis. Relative cell growth assays supported the theory that these derivatives were converted to β -lapachone within 4 hours after diluting the compounds in aqueous solutions (Fig. 2;

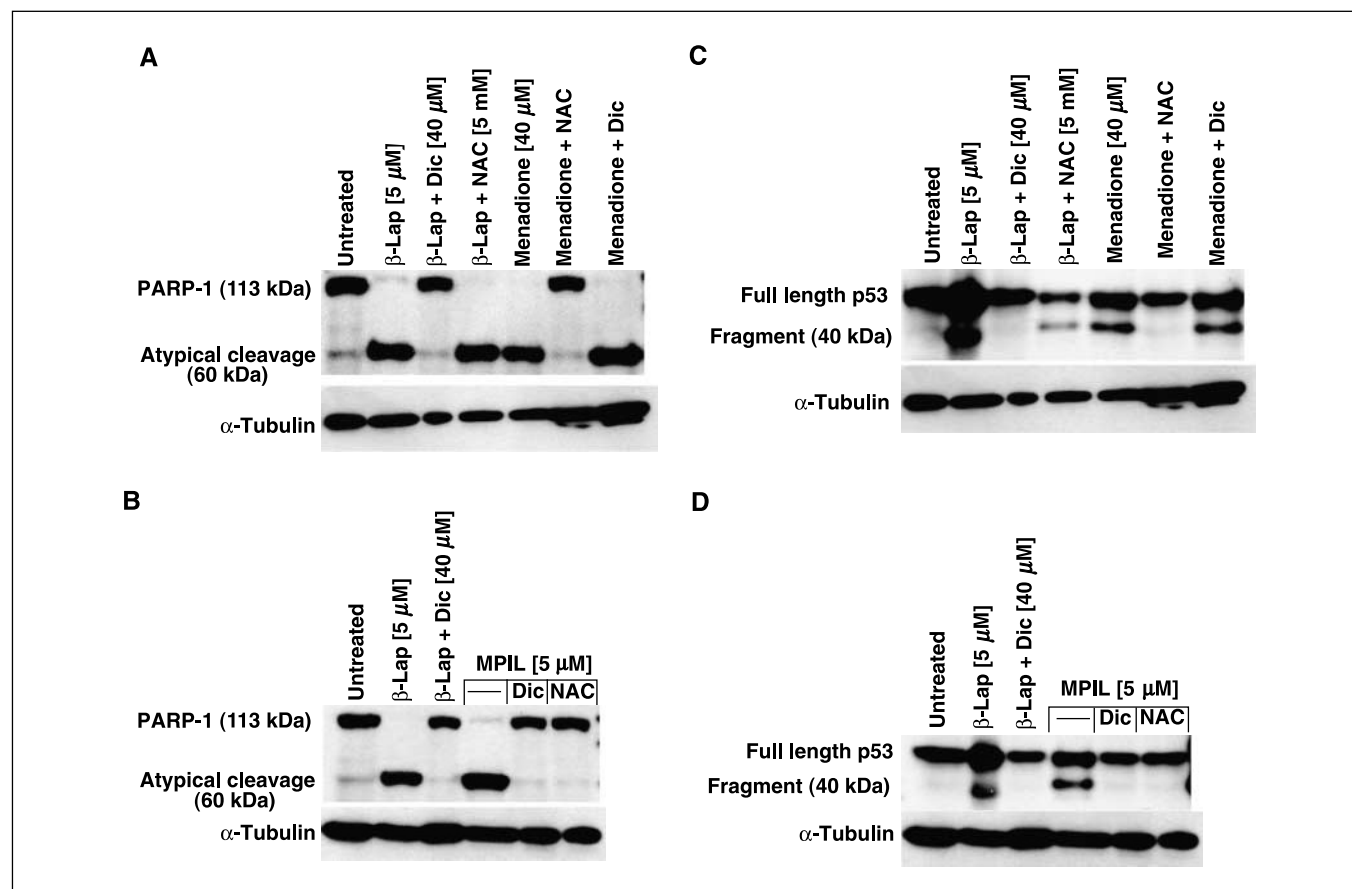


Fig. 4. β -Lapachone mono(arylimino) derivatives cause NQO1-dependent, atypical PARP and p53 apoptotic cleavage in MCF-7 cells similar to β -lapachone. MCF-7 cells were treated with 4-hour pulses of (A) 5 μ M β -lapachone or 40 μ M menadione or (B) 5 μ M MPIL $\pm$ 40 μ M dicoumarol (Dic) or 5 mmol/L NAC. A higher dose of menadione was necessary to induce apoptosis due to high levels of NQO1 in MCF-7 cells (6). Whole cell extracts were prepared at 24 hours and analyzed using standard Western analyses as described in Materials and Methods. Blots were probed with α -PARP antibody. Representative of experiments done at least thrice. MCF-7 cells were treated with 4-hour pulses of 5 μ M β -lapachone, 40 μ M menadione, or DMSO alone (C) or 5 μ M MPIL $\pm$ 40 μ M dicoumarol (Dic) or 5 mmol/L NAC (D). A higher dose of menadione was needed to induce apoptosis due to the elevated levels of NQO1 in MCF-7 cells (6). Whole cell extracts were prepared 24 hours post-treatment and analyzed using standard Western blot analyses as described in Materials and Methods. Blots were probed with α -p53 antibody. Representative of experiments done at least thrice.

Table 1; Supplementary Table 1). β -Lapachone prodrug-induced cell death was significantly less in NQO1⁻ cells and was inhibited by dicoumarol in NQO1⁺ breast, prostate, or non-small cell lung cancer cells, similar to β -lapachone (6, 18, 32). These results were consistent with hollow fiber assays (31), indicating that mono(arylimino) derivatives of β -lapachone may have an advantage over β -lapachone.

Direct evidence that specific β -lapachone mono(arylimino) derivatives spontaneously convert to β -lapachone in aqueous solution was provided by UV-visible spectrophotometric analyses (Supplementary Fig. 2), NMR analyses (Supplementary Fig. 4), high-performance liquid chromatography-electrospray ionization-mass spectrometry analyses (Supplementary Fig. 3; Supplementary Table 2), and preferential lethality in NQO1⁺ cells (Table 1; Supplementary Table 1). The data presented in Fig. 3A support our hypothesis that these β -lapachone mono(arylimino) derivatives need to hydrolyze to β -lapachone for reduction by NQO1. The rate of hydrolysis was dependent on the electron-withdrawing strength of the unique substituent phenylimine group of the compounds (Figs. 1 and 3; Supplementary Figs. 2 and 3; Supplementary Table 2; ref. 39). The ability of these derivatives to induce NQO1-dependent atypical PARP and p53 cleavage (Fig. 4; Supplementary

Fig. 5) confirmed their conversion to the biologically active β -lapachone parent compound (6, 16, 18, 32).¹¹ Conversion of β -lapachone mono(arylimino) derivatives to β -lapachone was noted by Misico and Forzani (39), where the hydrolytic decay time constants of β -lapachone mono-(arylimino) derivatives in aqueous solution were measured by cyclic voltammetry. In these studies, increased conversion was noted at lower pH. However, the biological consequences of the proposed conversion were not evaluated. Here, we show that β -lapachone mono(arylimino) derivatives are converted to active β -lapachone. Because conversion is enhanced at lower pH (Supplementary Fig. 1; refs. 26–30, 39), use of these β -lapachone derivatives for increased tumor-selective conversion in the acidic microenvironment of tumors (40–42) could be achieved.

Interestingly, the biologically active β -lapachone mono(arylimino) derivatives were fundamentally different from β -lapachone, because their cytotoxicity was blocked by NAC (5 mmol/L) cotreatments (Table 1; Supplementary Table 1).

¹¹ R.I. Misico, G. Arzac, G. Burton, J.J. Pink, M. Beman, E. Hohler, D. Boothman. 23rd IUPAC International Symposium on the Chemistry of Natural Products; Florence, Italy; July 28 to August 2, 2002.

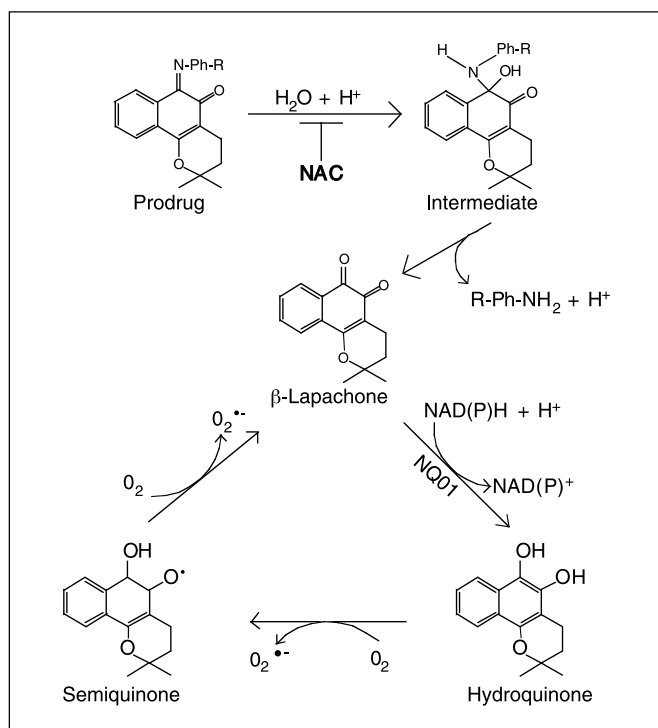


Fig. 5. Proposed mechanism of β-lapachone mono(arylimino) derivative conversion to β-lapachone followed by β-lapachone redox cycling. Our data strongly suggest that β-lapachone mono(arylimino) derivatives undergo a spontaneous Schiff's base hydrolytic reaction, converting the original nontoxic β-lapachone derivatives into the NQO1-dependent cytotoxic β-lapachone parent compound. The rate of this reaction seems to be dependent on the electron-withdrawing effect of the R group on the mono(arylimino) phenyl ring (Fig. 1). β-Lapachone undergoes a futile redox cycle with NQO1 wherein the hydroquinone form is unstable and through two one-electron oxidation steps converts back to the parent β-lapachone molecule. This cycling "bioactivates" β-lapachone and results in Ca²⁺-dependent, μ-calpain-mediated apoptotic cell death responses in which p53 and PARP are selectively cleaved (Fig. 4; Supplementary Fig. 5).

Consistent with the ability of NAC to inhibit the cytotoxicities of these β-lapachone derivatives, addition of NAC prevented spontaneous conversion to β-lapachone in the UV-visible spectrophotometric wavelength scans observed with these compounds in aqueous solutions (Supplementary Fig. 6). NMR data suggest that NAC directly interacts with these agents to create NAC-derivative conjugates (Supplementary Fig. 7), thereby preventing conversion to β-lapachone. Consequently, NAC exposure rendered the derivatives unable to interact with NQO1 and cause cell death. A similar phenomenon of lapachones undergoing sulfur-mediated Michael reactions was noted previously using L-cysteine and β-mercaptoethanol (43).

The rationale underlying development of this class of mono(arylimino) prodrugs for the delivery of β-lapachone for chemotherapy and/or radiotherapy is based on delayed, pH-dependent activation (26–30). After systemic administration of a potent β-lapachone mono(arylimino) prodrug, the chemotherapeutic agent must traverse the body through zones of neutral pH via the bloodstream and lymphatic

circulation, eventually reaching its target, acidic solid tumor tissue (40–42). We believe that mono(arylimino) derivatives of β-lapachone would cause minimal damage to normal tissues with neutral pH microenvironments during circulation. Ideally, these β-lapachone mono(arylimino) prodrugs will remain in inactive forms during the initial systemic distribution and will not become active until they successfully reach the tumor, which commonly has an acidic microenvironment (40–42). The resultant β-lapachone produced is then "bioactivated" by NQO1 (6), an enzyme found in human tissues at low levels (10, 44, 45) but expressed at high levels in many human cancers (21–24). Therefore, a mono(arylimino) prodrug of β-lapachone would remain inactive in circulation through neutral pH zones until it reached its target tissue (acidic tumors with elevated NQO1 levels). There, it would spontaneously become a high affinity substrate for NQO1 (6). Retention in tumor over normal tissue could be enhanced in NQO1-overexpressing cells due to futile cycling of the converted, and now active, β-lapachone compound.

The pharmacokinetics of a prodrug are expected to be vastly different from those of the parent drug. This was clearly noted in the development of CPT-11, a camptothecin derivative that undergoes de-esterification *in vivo* to yield SN-38 in the blood of mammals (46). We are in the process of analyzing the pharmacokinetics of β-lapachone compared with specific β-lapachone mono(arylimino) derivatives. The ideal β-lapachone prodrug may be one that requires a longer time to become active (e.g., NPIL or BPIL) and is possibly accelerated in an NQO1⁺ acidic tumor environment compared with a compound that is more rapidly activated (e.g., PIL, MePIL, or MPIL; ref. 39).

Collectively, the data presented in this article strongly suggest that some form of β-lapachone mono(arylimino) derivative can be developed for use as a prodrug. The observation that the electron-withdrawing group on the arylimino substitution group as well as the pH of the surrounding aqueous environment can directly influence conversion of the prodrugs to β-lapachone should be invaluable in developing these active chemotherapeutic agents. Analyses of this class of β-lapachone prodrugs could lead to potent, tumor-selective, antitumor agents. We are currently working on developing a β-lapachone mono(arylimino) compound with a leaving group that is specifically cleaved through a reaction catalyzed by a tumor-selective enzyme. Additionally, we are developing a derivative of β-lapachone with a cytotoxic leaving group that could further enhance the antitumor efficacy of these prodrugs, particularly in an acidic tumor microenvironment.

Acknowledgments

We thank Meghan Beman and Erin Hohler for preliminary work with these prodrugs, Norased Nasongkla and Fangjing Wang for help with the NMR analyses, and Sarah Hildebrand for encouragement and support of our research in the development of β-lapachone and its prodrugs.

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