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Award Number: DAMD17-03-1-0586

TITLE: Role of Rad51-Mediated Interactions in Recombination

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REPORT DATE: August 2005

TYPE OF REPORT: Annual Summary

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
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REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
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1. REPORT DATE (DD-MM-YYYY) 01-08-2005		2. REPORT TYPE Annual Summary		3. DATES COVERED (From - To) 1 Aug 2004 – 31 Jul 2005	
4. TITLE AND SUBTITLE Role of Rad51-Mediated Interactions in Recombination				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER DAMD17-03-1-0586	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Steven Raynard, Ph.D. E-mail: steven.raynard@yale.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Yale University New Haven, CT 06520-8047				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT Mutations in the BRCA2 gene are linked to familial and sporadic breast cancer, yet the molecular function of BRCA2 protein remains largely obscure. BRCA2 protein physically interacts with the Rad51 recombinase, a member of the RAD52 epistasis group of proteins that mediate homologous recombination (HR), a major mechanism that repairs chromosomes damaged by ionizing radiation and genotoxic agents. Accordingly, BRCA2 deficient cell lines exhibit impaired HR and sensitivity to genotoxic agents. To help define the molecular function of human BRCA2, we have expressed and purified a polypeptide that harbors the BRC3 and BRC4 repeat and also the DNA binding domain of this tumor suppressor. The BRC3/4-DBD polypeptide interacts with hRad51 and binds DNA with a distinct preference for ssDNA. Importantly, we have demonstrated by biochemical means that BRC3/4-DBD nucleates hRad51 onto ssDNA and enhances the homologous DNA pairing activity of hRad51. In isolation neither the BRC3-BRC4 repeats nor the DNA binding domain of BRCA2 performs these mediator functions. This biochemical system described in this study should be valuable for systematically dissecting the HR functions of BRCA2 in the context of the Rad51-mediated homologous DNA pairing reaction. Comprehending the manner in which BRCA2 modulates Rad51 activity and the functional integrity of the homologous recombination machinery could very well pave the way for devising new strategies in breast cancer diagnosis, prevention, and treatment.					
15. SUBJECT TERMS BRCA2, DNA repair, Homologous Recombination, Rad51					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT	b. ABSTRACT	c. THIS PAGE			USAMRMC
U	U	U	UU	9	19b. TELEPHONE NUMBER (include area code)

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Introduction

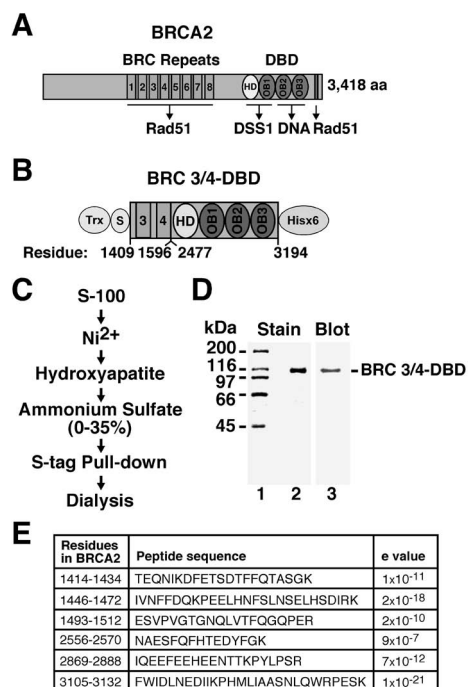
The repair of DNA double-strand breaks (DSB) induced by exogenous agents (e.g. ionizing radiation) or that arise endogenously (e.g. replication of damaged DNA template) is critical for the maintenance of genome stability. In eukaryotes, homologous recombination (HR) represents an important mechanism for the repair of DSBs and is mediated by genes of the *RAD52* epistasis group, whose structure and function are highly conserved (Symington, 2002; Sung et al., 2003). In HR-mediated DSB repair, the ends of the DNA break are processed nucleolytically to yield a pair of ssDNA tails, which serve as the nucleation site for the Rad51 recombinase. Polymerization of Rad51 onto the ssDNA results in the formation of a right-handed nucleoprotein filament, also commonly referred to as the presynaptic filament (Symington, 2002; Sung et al., 2003). After locating a DNA homologue, the presynaptic filament catalyzes pairing with the homologue to form a nascent DNA joint called a displacement loop (D-loop), the length of which is extended by DNA strand exchange. Subsequent steps include resolution of recombination intermediates and ligation to complete the recombination/repair reaction.

Mutations in the *BRCA2* (breast cancer susceptibility 2) gene are found in a significant portion of familial breast cancer cases and confer an increased risk of ovarian, pancreatic, and prostate cancer (Ford et al., 1998; Wooster et al., 1995). Mutations in *BRCA2* can also lead to the cancer prone-syndrome Fanconi anemia (D'Andrea, 2003). Significantly, cell lines deficient in *BRCA2* function exhibit genome instability, hypersensitivity to DNA damaging agents, and a pronounced deficiency in HR (Jasin, 2002). Cytological results indicate that *BRCA2* is important for the assembly of DNA damage-induced Rad51 nuclear foci (Yang et al., 2002; Tarsounas et al., 2003). *BRCA2* physically interacts with Rad51 through a series of eight copies of a reiterated motif called the BRC repeat. Furthermore, *BRCA2* possesses a single-stranded DNA (ssDNA) binding function (Yang et al., 2002) and tightly associates with a small partner protein called DSS1 (Marston et al., 1999), which is also needed for DSB repair by HR (Gudmundsdottir et al., 2004). These features of *BRCA2* are consistent with the possibility that it acts as a recombination mediator by promoting the assembly of the Rad51 presynaptic filament. This report details the progress made towards elucidating the molecular function of *BRCA2* in HR.

Body

(i) Expression and purification of hBRCA2-derived polypeptides - Human *BRCA2* contains eight BRC repeats in its middle portion and a DNA binding domain (DBD) toward the carboxyl terminus as deduced from biochemical and crystallographic studies done with the mouse *BRCA2* orthologue (Yang et al., 2002) (Figure 1A). Due to its enormous size (3,418 amino acid residues), we have not yet been able to express sufficient amounts of the full-length human *BRCA2* protein for purification. However, since several *BRCA2*-like molecules from organisms such as *Ustilago maydis* and *C. elegans* are of much smaller size and contain only a single BRC, we wished to test the premise that by fusing selected BRC repeats and the DBD, we would be able to synthesize a form of human *BRCA2* protein that is capable of physical and functional interactions with human Rad51

(hRad51) protein. Accordingly, we constructed a polypeptide that consists of the BRC3 and BRC4 repeats and the DBD derived from human BRCA2 (Figure 1B). To facilitate



the purification of the BRC3/4-DBD polypeptide, a Thioredoxin/S-protein-tag and hexahistidine-tag were attached to its amine and carboxy-termini, respectively. The BRC3/4-DBD fusion protein expressed in *E. coli* is soluble, and a simple four-step purification procedure encompassing: nickel-NTA beads, hydroxyapatite column chromatography, ammonium sulfate precipitation and S-tag affinity pulldown, was devised to purify it to near homogeneity (Figure 1C,D). The identity of the purified protein was established by immunoblot analysis and MALDI-TOF (Figure 1E). Up to 200 μ g of the purified BRC3/4-DBD can be obtained from 400 g of *E. coli* cell paste. We also expressed and purified (1) a GST fusion protein that contains the BRC3 and BRC4 repeats (GST-BRC3/4) and (2) a (His)₆-tagged form of the DBD (data not shown).

Figure 1 Purification of BRC3/4-DBD. **(A)** Schematic outlining the functional domains in BRCA2. The DNA binding domain (DBD) consists of a helical domain (HD) and three oligo-nucleotide binding (OB1, OB2, and OB3) folds that confer the ability to interact with DSS1 and ssDNA. **(B)** The human BRCA2-derived polypeptide BRC3/4-DBD harbors two of the BRC repeats (BRC3 and BRC4 within residues 1409-1596) linked to the DBD (residues 2477 to 3194). BRC3/4-DBD contains thioredoxin (Trx) and S tags at its amino-terminus and a (His)₆ tag at its carboxyl-terminus. **(C)** Schematic of the chromatographic procedure devised for BRC3/4-DBD purification. **(D)** Purified BRC3/4-DBD was analyzed by SDS-PAGE and Coomassie Blue staining (2 μ g in lane 2) or immunoblotting with anti-histidine antibodies (200 ng in lane 3). **(E)** Results from MALDI-TOF analysis of purified BRC3/4-DBD. Six representative fragments and their corresponding e values are shown.

(ii) Interaction of BRC3/4 and BRC3/4-DBD with hRad51 - Taking advantage of the affinity tags on BRC3/4-DBD, GST-BRC3/4, and DBD, we used several *in vitro* pulldown assays to investigate whether BRC3/4-DBD binds hRad51. First, purified hRad51 was incubated with BRC3/4-DBD, and the resulting protein complexes were captured on anti-S antibody agarose beads through the S-tag on BRC3/4-DBD. As shown in Figure 2A, BRC3/4-DBD bound hRad51. Affi-gel beads containing crosslinked hRad51 were used to verify the role of the BRC repeats in hRad51 binding. As anticipated, BRC3/4-DBD and GST-BRC3/4 bound the hRad51 beads with considerable avidity, while DBD did not (Figure 2B). We also examined whether the *E. coli* RecA protein and human Dmc1 (hDmc1) protein interact with the two BRCA2-derived polypeptides. No association of either RecA or hDmc1 with either BRCA2-derived polypeptide was seen (Figure 2A,C and data not shown). We conclude that BRC3/4-DBD associates with hRad51 in a specific manner, through the BRC repeats.

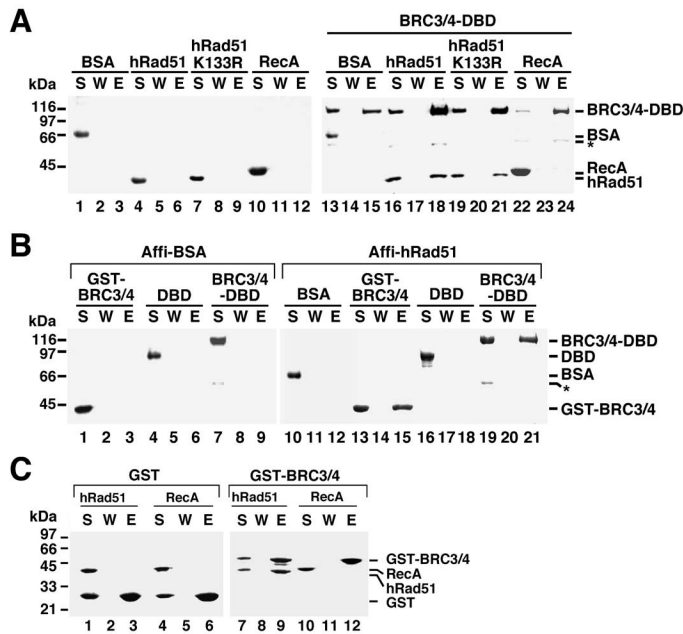


Figure 2 BRC3/4-DBD binds hRad51. (A) BSA, hRad51, hRad51K133R, and RecA were mixed with anti-S protein agarose beads in the absence (lanes 1-12) or presence of BRC3/4-DBD (lanes 13-24) and then subject to affinity pulldown with anti-S agarose beads. (B) GST-BRC3/4, DBD, and BRC3/4-DBD were subject to affinity pulldown with Affi-beads conjugated to BSA (Affi-BSA; lanes 1-9) or hRad51 (Affi-Rad51; lanes 10-21). (C) Pulldown assays using GST or GST-BRC3/4 on glutathione Sepharose confirmed that BRC3/4 binds Rad51 but not RecA. The supernatant (S), wash (W), and SDS eluate (E) from the above reactions were analyzed by SDS-PAGE and Coomassie Blue staining. A proteolytic product of BRC3/4-DBD is marked by the asterisk.

(iii) **BRC3/4-DBD binds ssDNA preferentially** - The mouse Brca2 protein possesses three OB (oligonucleotide-oligosaccharides binding) folds that endow it with DNA binding ability. We wished to verify that the BRC3/4-DBD species that we constructed from the human BRCA2 sequence also has DNA binding capability. To do this, increasing amounts of the purified BRC3/4-DBD was incubated with either 32 P-labeled 80mer ssDNA (Figure 3A) or 32 P-labeled 80bp duplex (Figure 3B) obtained by hybridizing the former oligonucleotide to its complement. The reaction mixtures were resolved in non-denaturing polyacrylamide gels, which were dried and then analyzed in the phosphorimager to reveal shifting of the DNA species by BRC3/4-DBD. Consistent with published data obtained with the mouse Brca2 DBD, BRC3/4-DBD binds DNA and with a distinct preference for the ssDNA species (Yang et al., 2002). As expected, when we incubated BRC3/4-DBD with the mixture of ssDNA and dsDNA, it first shifted all of the ssDNA substrate before binding the dsDNA (Figure 3C,D). The DBD alone also bound DNA with a similar preference for the ssDNA species and we could verify GST-BRC3/4 without the DBD does not possess any DNA binding activity (data not shown). Taken together, the

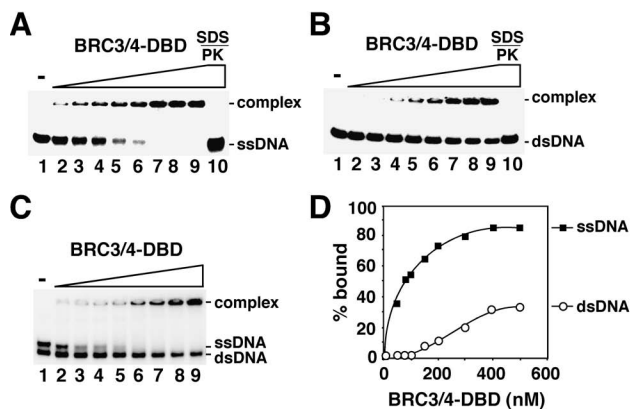


Figure 3 BRC3/4-DBD has high affinity for ssDNA. (A) & (B) Increasing amounts of purified BRC3/4-DBD (20-500 nM, lanes 2-9) was incubated with 30 nM of 32 P-labeled ssDNA or dsDNA and then analyzed. Treatment of the nucleoprotein with SDS and proteinase K (SDS/PK) released the DNA substrate (lane 10). (C) BRC3/4-DBD (50 to 500 nM) was incubated with the mixture of 32 P-labeled ssDNA and dsDNA and then analyzed. (D) The results from the experiment in C are plotted.

data clearly show that the human BRCA2 protein harbors a DNA binding function that is highly specific for ssDNA.

(iv) *BRC3/4-DBD targets Rad51 to ssDNA* - In mediating HR, Rad51 protein must first assemble onto ssDNA to form a helical nucleoprotein filament, which provides the catalytic center for the pairing of recombining DNA molecules (Sung et al., 2003). The assembly of the Rad51-ssDNA nucleoprotein filament is hampered by (1) slow nucleation of Rad51 onto ssDNA, (2) competition for binding site on the DNA by RPA, and (3) sequestration of Rad51 on dsDNA. Given that BRC3/4-DBD physically interacts with hRad51 and binds ssDNA with avidity, we tested to see if BRC3/4-DBD could target hRad51 to ssDNA. To address this question, we incubated hRad51 with a mixture of dsDNA and biotinylated ssDNA with or without BRC3/4-

DBD, followed by capture of the biotinylated ssDNA on magnetic beads that contains streptavidin, to which the biotin moiety on the ssDNA binds with high affinity. The magnetic beads and the supernatant fractions were treated with SDS and then subject to SDS-PAGE to determine their content of Rad51 and BRC3/4-DBD (Figure 4A). The results showed that inclusion of BRC3/4-DBD increased the percentage of Rad51 associated with the magnetic bead-bound ssDNA (Figure 4B, lanes 4-5). Importantly, neither GST-BRC3/4 nor the DBD alone was effective in enhancing the association of Rad51 with ssDNA (Figure 4B, lanes 6-7).

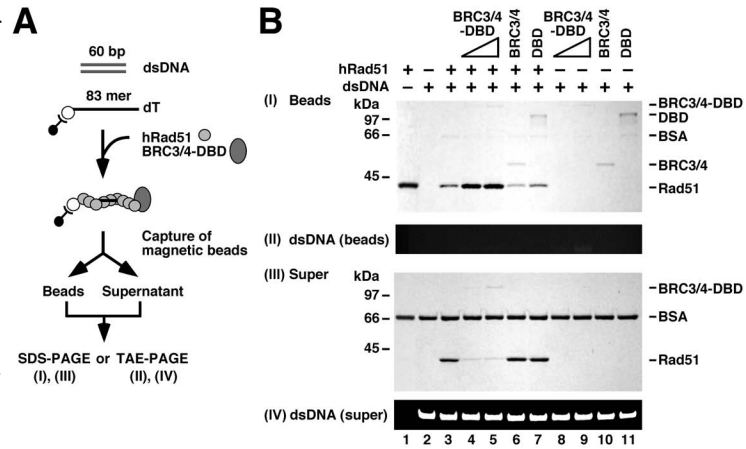


Figure 4 BRC3/4-DBD targets hRad51 to ssDNA. **(A)** Schematic of the assay. Magnetic bead-bound oligo dT was incubated with hRad51, BSA and BRCA2-derived polypeptides, without or with an excess of dsDNA, as indicated. Proteins bound to the oligo dT were captured with a magnet and then eluted with SDS. **(B)** The supernatants (super) and SDS eluates (beads) were analyzed for their protein and DNA contents. While, as expected, the majority of hRad51 was trapped on the dsDNA (lane 3), BRC3/4-DBD efficiently targeted hRad51 to the ssDNA (lanes 4 and 5). GST-BRC3/4 (BRC3/4) or DBD was ineffective in this regard (lanes 6 and 7, respectively).

(v) *BRC3/4-DBD stimulates Rad51-mediated homologous DNA pairing* - Given that BRC3/4-DBD binds both Rad51 and ssDNA and is capable of targeting Rad51 to ssDNA, we used an oligonucleotide-based assay to enquire whether BRC3/4-DBD enhances the homologous DNA pairing activity of Rad51. Indeed, the results clearly showed a stimulation of homologous DNA pairing when BRC3/4-DBD was added with Rad51 (Figure 5).

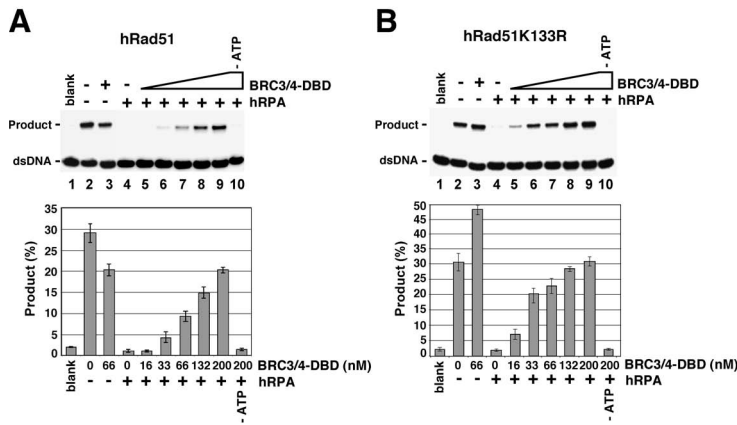


Figure 5 Recombination mediator activity of BRC3/4-DBD. (A) & (B) Homologous DNA pairing reactions containing hRad51 or hRad51 K133R and varying amounts of BRC3/4-DBD were carried out with ssDNA or hRPA-coated ssDNA, as indicated. The averaged values of results from three independent experiments are presented in the histograms. The no protein control (*lane 1*) is marked as “blank”, and ATP was omitted from the reaction in *lane 10*.

Key Research Accomplishments

- Expressed and purified a polypeptide that harbors the BRCA2 DNA binding domain and two of the BRC repeats, BRC3 and BRC4
- Demonstrated that the BRCA2-derived polypeptide, BRC3/4-DBD interacts with hRad51 through the BRC motifs, and binds DNA with a distinct preference for ssDNA
- Demonstrated by biochemical means that BRC3/4-DBD is capable of nucleating hRad51 on ssDNA, and can act as a recombination mediator by enabling hRad51 to utilize hRPA-coated ssDNA as a recombination substrate

Reportable Outcomes

Papouli, E., Chen, S., Davies, A. A., Huttner, D., **Krejci, L.**, Sung, P. & Ulrich, H. D. (2005) Crosstalk between SUMO and ubiquitin on PCNA is mediated by recruitment of the helicase Srs2p. **Mol. Cell** 19:123-33.

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Chen, L., Trujillo, K. M., Van Komen, S., Roh, D. H., **Krejci, L.**, Lewis, L. K., Resnick, M. A., Sung, P. & Tomkinson, A. E. (2005) Effect of amino acid substitutions in the rad50 ATP binding domain on DNA double strand break repair in yeast. **J. Biol. Chem.** 280:2620-7.

Conclusions

A role for BRCA2 in HR and DNA repair is now well established, however, relatively little is known about the protein's function at a mechanistic level. The studies summarized in this report support a role for BRCA2 as a recombination mediator, specifically functioning to promote the assembly of the Rad51 presynaptic filament. This biochemical system should be valuable for systematically dissecting the HR functions of BRCA2 in the context of the Rad51-mediated homologous DNA pairing reaction. Comprehending the manner in which BRCA2 modulates Rad51 activity and the functional integrity of the HR machinery could very well pave the way for devising new strategies in breast cancer diagnosis, prevention, and treatment.

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