



U.S. Army Medical Research
Institute of Chemical Defense

USAMRICD-TR-06-05

A Method for the Analysis of
Tabun in Multisol Using Gas
Chromatographic Flame
Photometric Detection

Thomas P. Logan
Edward D. Allen
Mark R. Way
Austin T. Swift
Sunil-Datta Soni

May 2006

Approved for public release; distribution unlimited

U.S. Army Medical Research
Institute of Chemical Defense
Aberdeen Proving Ground, MD 21010-5400

DISPOSITION INSTRUCTIONS:

Destroy this report when no longer needed. Do not return to the originator.

DISCLAIMERS:

The opinions or assertions contained herein are the private views of the author(s) and are not to be construed as official or as reflecting the views of the Army or the Department of Defense.

The use of trade names does not constitute an official endorsement or approval of the use of such commercial hardware or software. This document may not be cited for purposes of advertisement.

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE (DD-MM-YYYY) May 2006		2. REPORT TYPE Technical Report		3. DATES COVERED (From - To)	
4. TITLE AND SUBTITLE A Method for the Analysis of Tabun in Multisol Using Gas Chromatographic Flame Photometric Detection				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Logan, TP, Allen, ED, Way, MR, Swift, AT, Soni, S-D				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) US Army Medical Research Institute of Chemical Defense ATTN: MCMR-CDT-D 3100 Ricketts Point Road Aberdeen Proving Ground, MD 21010-5400				8. PERFORMING ORGANIZATION REPORT NUMBER USAMRICD-TR-06-05	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) US Army Medical Research Institute of Chemical Defense ATTN: MCMR-CDA-T 3100 Ricketts Point Road Aberdeen Proving Ground, MD 21010-5400				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for public release; distribution unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT In the past we have found tabun (GA) in saline at 2 mg/ml to be stable for only a month or less at -70 °C. Previous studies have shown that Multisol provides stable solutions of GA. We confirmed the stability of GA in Multisol with phosphorus nuclear magnetic resonance and developed a method for the analysis of GA in Multisol using gas chromatographic flame photometric detection (GCFPD) in the phosphorus mode. The GC method used acetonitrile (CH ₃ CN) for a dilution solvent because of its miscibility with GA in chloroform (CHCl ₃) standards and GA in Multisol samples at 1% (v/v). Furthermore, the dilutions with CH ₃ CN made the phosphorus mode interference peak present in CHCl ₃ analytically manageable, reduced the interferences of Multisol in the GC separation, and contributed to a safe and reliable analysis of GA at 20 µg/ml. We demonstrated the stability of GA in Multisol stored for more than a year at -70 °C. This method contributes a suitable technique for the preparation and analysis of reliable solutions of GA in nerve agent medical research and demonstrates the extended stability of GA in Multisol.					
15. SUBJECT TERMS GA, Tabun, Multisol, Gas Chromatography, Flame Photometric Detector, Nuclear Magnetic Resonance					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT UNLIMITED	18. NUMBER OF PAGES 13	19a. NAME OF RESPONSIBLE PERSON Thomas P. Logan
a. REPORT UNCLASSIFIED	b. ABSTRACT UNCLASSIFIED	c. THIS PAGE UNCLASSIFIED			19b. TELEPHONE NUMBER (include area code) 410-436-1484

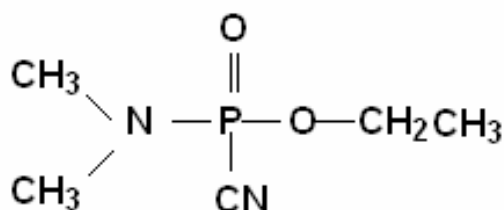
Abstract

Preparation and analysis of tabun (GA) solutions are necessary for the continued development of countermeasures to this nerve agent. GA solutions must be stable and compatible for use in the test systems chosen for study; however, GA is very unstable in saline solutions. In the past we have found GA in saline at 2 mg/ml to be stable for a month or less at -70 °C, whereas saline solutions of sarin (GB), soman (GD), and cyclosarin (GF) were stable for many months. Previous studies have shown that Multisol (48.5% H₂O, 40% propylene glycol, 10% ethanol, and 1.5% benzyl alcohol) provides stable solutions of GA. We confirmed the stability of GA in Multisol with phosphorus nuclear magnetic resonance (P-NMR) and developed a method for the analysis of GA in Multisol using gas chromatographic flame photometric detection (GCFPD) in the phosphorus mode. The GC method used acetonitrile (CH₃CN) for a dilution solvent because of its miscibility with GA in chloroform (CHCl₃) standards and GA in Multisol samples at 1% (v/v). Furthermore, the dilutions with CH₃CN made the phosphorus mode interference peak present in CHCl₃ analytically manageable, reduced the interferences of Multisol in the GC separation, and contributed to a safe and reliable analysis of GA at 20 µg/ml. We demonstrated the stability of GA in Multisol stored for more than a year at -70 °C. This method contributes a suitable technique for the preparation and analysis of reliable solutions of GA in nerve agent medical research and demonstrates the extended stability of GA in Multisol.

Introduction

Tabun (GA) is a chemical warfare agent against which the United States is actively developing countermeasures. GA was developed by Schrader in 1936 as a pesticide and became one of the German (G) nerve agents.

GA continues to be of interest¹⁻⁴ as a chemical warfare agent due to its suspected use by Iraq, its potential use as a terrorist threat agent, and its historical reference as one of the original G nerve agents. Furthermore, GA-induced toxic effects are difficult to counteract due to the very low reactivating efficacy of pralidoxime chloride (2-PAMCl), the oxime currently used by the United States as a nerve agent antidote.^{5,6} Security⁷ and safety⁸ regulations of the United States Army constrain dilute nerve agent research to solutions of 2 mg/ml or less. The stability of these solutions is a concern for researchers. The structure and chemical name of GA are shown below:



Dimethylphosphoramidocyanidic acid, ethyl ester

This structure is highly susceptible to hydrolysis at the -CN , $\text{-OC}_2\text{H}_5$, and $\text{-N(CH}_3)_2$ bonds to phosphorus. In neutral or near neutral aqueous solutions, GA hydrolyzes rapidly at the P-CN bond giving ethyl hydrogen N,N -dimethylphosphoroamidate as the major product.⁹ GA is unstable in aqueous solutions¹⁰⁻¹⁴ even when prepared and immediately stored at -70°C . We developed a program to monitor the stability of dilute solutions of nerve agent. Our guideline for solution stability is an analysis result that is within $\pm 10\%$ of the original gravimetric preparation concentration for that solution. It is our experience that saline solutions of sarin (GB), soman (GD), and cyclosarin (GF) remain stable for six months to a year at -70°C , while saline solutions of GA may be stable for one month at -70°C . Obviously, GA's instability is problematic when providing this agent in saline for the researcher. Therefore an alternative solvent for GA was sought to replace saline in research investigating medical countermeasures to this toxic compound. A vehicle that we called Multisol has been used at this institute for years as a solvent for compounds that are insoluble in water. The composition of Multisol (48.5% H_2O , 40% propylene glycol, 10% ethanol, and 1.5% benzyl alcohol) is similar to the vehicle used in the commercial formulation of Valium® for injection.¹⁵ Multisol has been used to provide stable GA solutions,¹⁶ and this four-component mixture was found to be an acceptable substitute for saline in subcutaneous injections of GA solutions.

Two instrumental methodologies, ^{31}P nuclear magnetic resonance (NMR) and gas chromatographic flame photometric detection (GCFPD), were used to demonstrate the stability of GA in Multisol. ^{31}P nuclear magnetic resonance (NMR) was used to compare the stability of GA in Multisol with the instability of GA in saline by monitoring the ^{31}P resonance of GA. The phosphorus mode GCFPD method was used to remove interferences from nonphosphorus components. The sensitivity of the phosphorus mode permitted the use of acetonitrile (CH_3CN)

as a dilution solvent for GA in chloroform (CHCl_3) standards and GA in Multisol samples at 1% (v/v).

Materials and Methods

Materials. Tabun (dimethylphosphoramidocyanidic acid, ethyl ester, GA) was obtained from the US Army Edgewood Chemical Biological Center (ECBC), Aberdeen Proving Ground, MD; GA purity was 98.5% as determined by NMR spectroscopy. Deuterium oxide (D_2O) 99.9 atom % D and acetonitrile (CH_3CN), HPLC grade, were obtained from Aldrich Chemical Company, Milwaukee, WI. Absolute ethyl alcohol was obtained from Pharmco, Brookfield, CT. Propylene glycol was obtained from Phoenix Pharmaceutical Inc., St. Joseph, MO. Benzyl alcohol was obtained from Eastman Kodak, Rochester, NY. CHCl_3 , pentene-stabilized HPLC grade, was obtained from Fisher Scientific Company, Pittsburgh, PA. Diisopropyl methanephosphonate (DIMP) was obtained from Lancaster Synthesis, Pelham, NH.

NMR measurements were made on GA in modified Multisol (48.5% D_2O , 40% propylene glycol, 10% ethanol, and 1.5% benzyl alcohol) and on GA in modified saline (D_2O containing 0.9 mg/ml NaCl). Solutions of GA were prepared gravimetrically at 2 mg/ml and stored at -70°C . Immediately upon thawing and mixing at room temperature, approximately 500 μL of the solution was transferred into a 5 mm o.d. NMR tube. All NMR data was collected on a Varian Unity Inova spectrometer equipped with 5 mm gradient Penta^R probe. Probe temperature was maintained at $25^\circ\text{C} \pm 0.1^\circ\text{C}$ and the VT air flow was maintained at 10 liters per minute (LPM). Standard s2pul sequence with continuous proton decoupling [$^{31}\text{P}\{^1\text{H}\}$] was used and all the calibrations were done as per the installation procedure. Chemical shifts were reported in parts per million (ppm) using 85% phosphoric acid (H_3PO_4) as the external reference at -0.9 ppm.

Gas chromatographic analyses of GA in Multisol samples and GA in CHCl_3 standards were made after 1:100 dilution with CH_3CN containing 0.0056 mM DIMP as the internal standard. An HP 5890 series II Plus gas chromatograph/Flame photometric detector was used for detection of GA. The gas chromatograph was equipped with a 30 m, 0.25 mm i.d., 0.25 mm DB-5MS column (J&W Scientific, Folsom, USA). Initial temperature was 70°C for 2 min, and the temperature ramp rate was $30^\circ\text{C min}^{-1}$ until the final temperature 250°C was reached and held for 3 min. Samples were run in the split mode 20:1 with a helium carrier gas flow rate of 1 ml min^{-1} measured at 70°C .

Results

^{31}P -NMR demonstrated the stability of GA in modified Multisol for 14 h compared with the instability of GA in modified saline for 5 h. GCFPD was used to quantify GA in Multisol samples against stable GA in chloroform standards.

Figure 1 shows the stability of GA in modified Multisol for 14 h at 25°C using ^{31}P NMR to monitor the GA and potential decomposition product resonances in a sample at 2 mg/ml. The (-10) ppm peak is the GA resonance that remains the major component of the 28 spectra.⁹ In addition, a small resonance at (-0.9) ppm can be seen early but remains a minor component. This result was a clear indication that GA in Multisol could be used for several hours at 25°C before the onset of decomposition.

Figure 2 demonstrates the instability of GA in saline, modified with D_2O , over a 5-h period at 25°C using ^{31}P -NMR to monitor the GA and decomposition product resonances. The

(-10) ppm peak is the GA resonance that decreases through the 11 spectra. When figures 1 and 2 are compared the instability of GA is readily apparent.

Figure 3 shows the chromatogram obtained for the analysis of a 20 ug/ml solution (1 ul injection slit 20/1) obtained from the dilution of GA in CHCl_3 with CH_3CN containing DIMP. Peak #1 is a chloroform impurity, peak #2 is DIMP, and peak #3 is GA.

Figure 4 shows the chromatogram obtained for the analysis of a 20 ug/ml solution (1 ul injection slit 20/1) obtained from the dilution of GA in Multisol with CH_3CN containing DIMP. Peak #1 is DIMP and peak #2 is GA.

Table 1 contains the GC/MS analysis results for a GA in Multisol (GA-M) sample gravimetrically prepared at 1.89 mg/ml. The first analysis result, 1.82 mg/ml, was determined one day after preparation and storage at -70°C from three analyses of a single dilution of a GA-M sample. The second analysis result, 1.76 mg/ml, was obtained following five months of storage at -70°C using three analyses of three separate dilutions of a GA-M sample. The third analysis result, 1.75 mg/ml was obtained 14 months after preparation and storage at -70°C using three analyses of three separate dilutions of a GA-M sample.

Time after preparation	Concentration mg/ml (deviation from gravimetric concentration)
1 day	1.82 (-4 %)
5 months	1.76 (-7 %)
14 months	1.75 (-7 %)

Table 1. GC/MS analysis results at selected times after preparation demonstrate GA in Multisol within $\pm 10\%$ of the gravimetric concentration at 14 months.

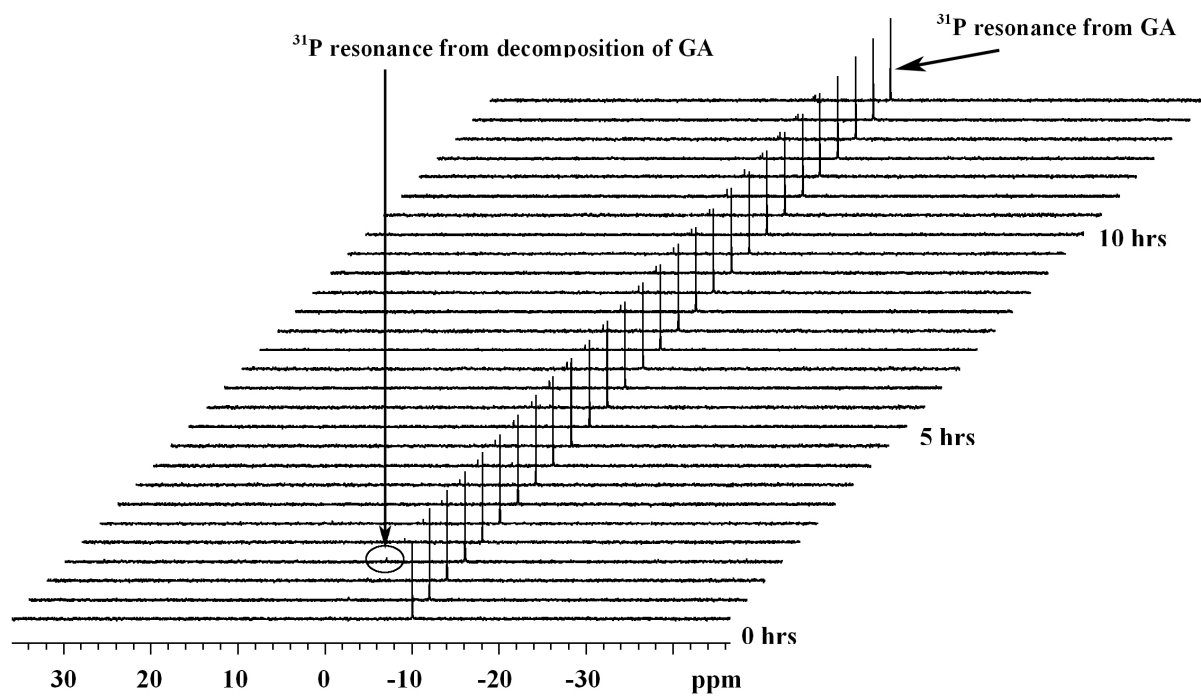


Figure 1. Stability of GA in Multisol Demonstrated with ^{31}P -NMR

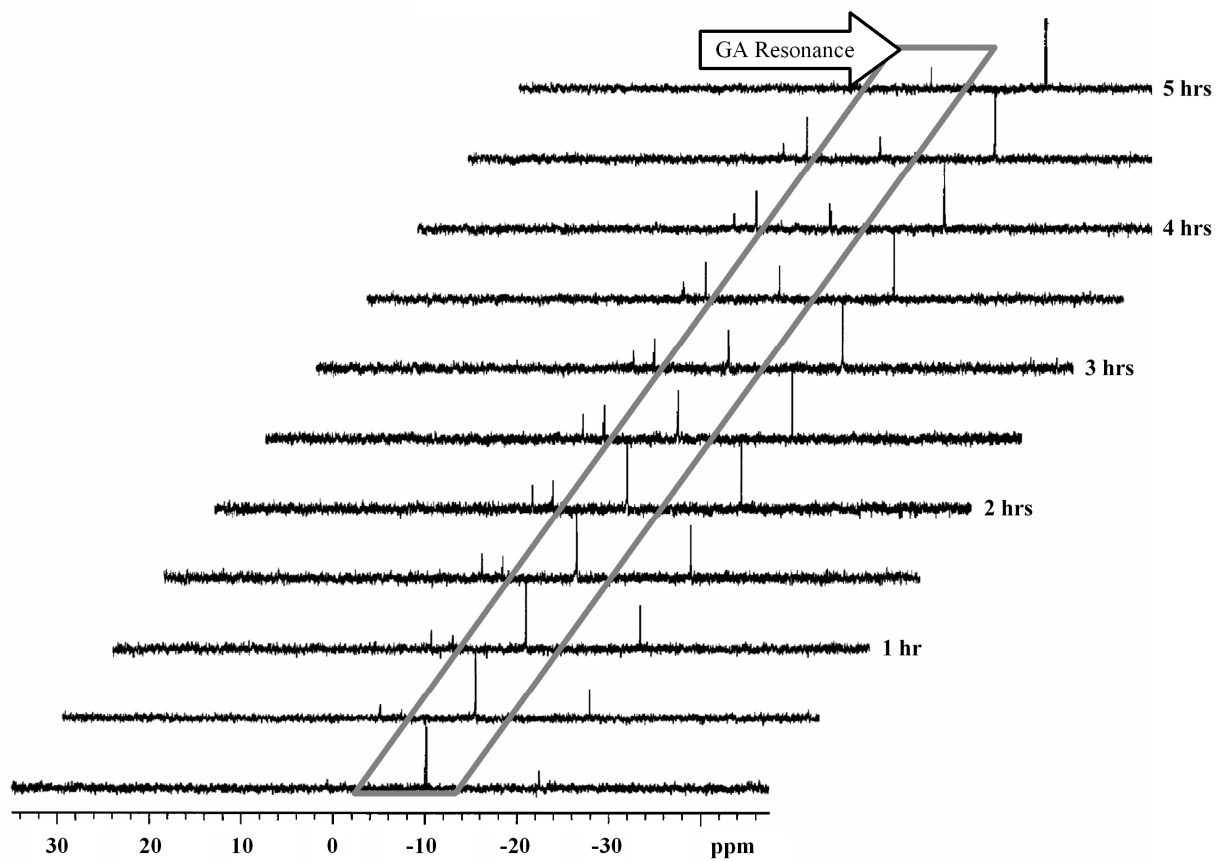


Figure 2. Instability of GA in Saline Demonstrated with ³¹P NMR

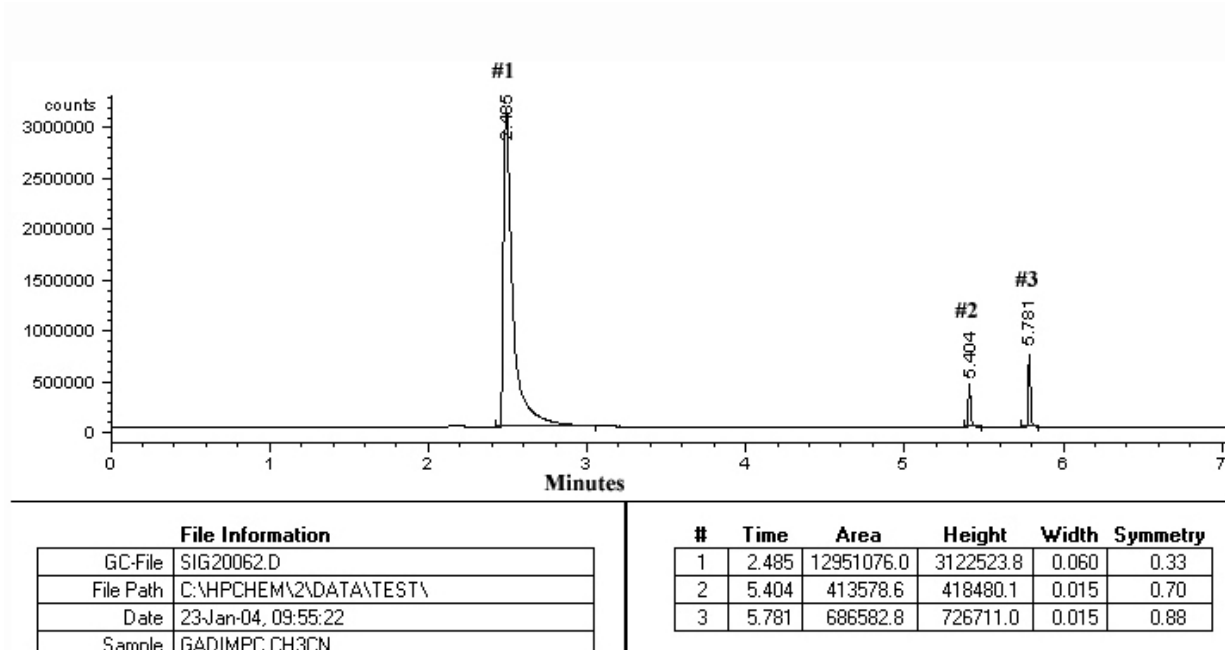


Figure 3. Chromatogram of GA in CHCl_3 after Dilution with CH_3CN containing DIMP as the internal standard. Peak #1 is CHCl_3 impurity. Peak #2 is DIMP. Peak #3 is GA.

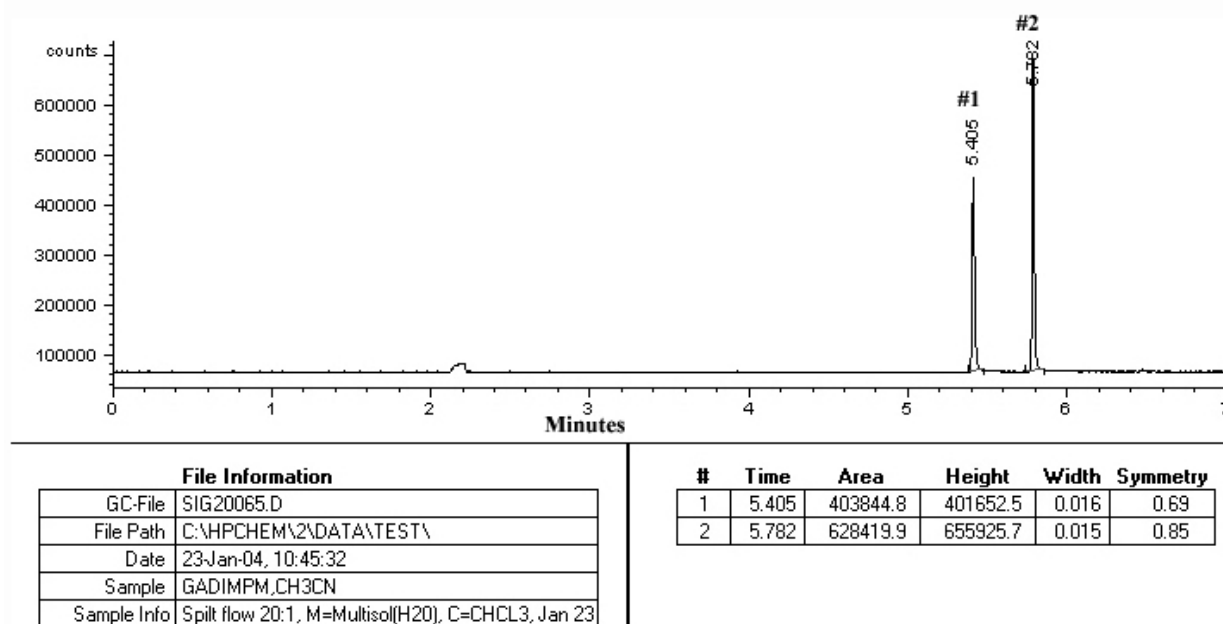


Figure 4. Chromatogram of GA in Multisol after Dilution with CH_3CN containing DIMP as the internal standard. Peak #1 is DIMP. Peak #2 is GA.

Discussion

GA is unstable in aqueous solutions. Our analysis results routinely show the concentration of GA in saline to be less than 90% of the gravimetric concentration within a month of preparation while kept at -70°C . Because the toxicity of GA is a frequently used reference in research to develop medical countermeasures to chemical warfare nerve agents, we sought a suitable solvent substitute for saline in the preparation and analysis of stable GA solutions. We chose Multisol as a substitute for saline because it was used previously to prepare anticholinergic,^{17, 18} anticonvulsant,^{15,18,19} and GA solutions¹⁶ that were stable and suitable for research. We confirmed the stability results¹⁶ and extended the shelf life of GA in Multisol to fourteen months.

As part of our investigation we used 31-P NMR as a methodology separate from gas chromatography to further investigate and confirm GA's extended stability in Multisol, Figure 1, versus GA's instability in saline, Figure 2. In Figure 1 the Multisol stabilizes the GA resonance at (-10) ppm against rapid hydrolysis as shown by the slow rise over 14 hrs of the resonance at (-0.9) ppm, which we attribute to the first hydrolysis product, ethyl hydrogen N,N-dimethylphosphoramidate.⁹ In Figure 2 we see the rapid onset of multiple hydrolysis product resonances as the GA resonance (-10) ppm decreases over 5 hrs.

We developed a relatively simple GCFPD method in the phosphorus mode for a routine and sensitive GC method of analysis for GA in Multisol. The phosphorus mode removed potential interferences from components in Multisol that do not contain phosphorus. This GC method by virtue of dilution in CH_3CN reduced and made manageable a peak present in CHCl_3 and detected in the phosphorus mode, Figure 3. The dilution with CH_3CN also removed the problem of sample immiscibility that arose with other solvents and reduced the GC column interferences due to the components of Multisol. Using this method we demonstrated that GA in Multisol solutions is stable for fourteen months when stored at -70°C . The extended shelf life at -70°C of GA in Multisol versus GA in saline provides more reliable solutions of this nerve agent for our research.

Finally, we contributed a suitable technique for the preparation and analysis of reliable solutions of GA. We offer Multisol for consideration as a possible solvent system for researchers working with analytes that are unstable or insoluble in completely aqueous based solvents.

References

1. Cabal J, Kuca K, Kassa J. Specification of the structure of oximes able to reactivate tabun-inhibited acetylcholinesterase. *Basic Clin Pharmacol Toxicol.* 2004; 95:81-86.
2. Kassa J, Krejcova G. Neuroprotective effects of currently used antidotes in tabun-poisoned rats. *Pharmacol Toxicol.* 2003; 92:258-264.
3. Krejcova G, Kassa J. Neuroprotective efficacy of pharmacological pretreatment and antidotal treatment in tabun-poisoned rats. *Toxicology* 2003; 185:129-139.
4. Kassa J, Vachek J. A comparison of the efficacy of pyridostigmine alone and the combination of pyridostigmine with anticholinergic drugs as pharmacological pretreatment of tabun-induced rats and mice. *Toxicology* 2002; 177:179-185.
5. Koplovitz I, Menton R, Matthews C, Shutz M, Nalls C, Kelly S. Dose-response effects of atropine and HI-6 treatment of organophosphorus poisoning in guinea pigs. *Drug Chem Toxicol.* 1995; 18:119-36.
6. Koplovitz I, Stewart JR. A comparison of the efficacy of HI6 and 2-PAM against soman, tabun, sarin, and VX in the rabbit. *Toxicol Lett.* 1994; 70:269-279.
7. Army Regulation 50-6, Nuclear and Chemical Weapons and Materiel, CHEMICAL SURETY, Headquarters, Department of the Army, Washington, DC, 26 June 2001.
8. Army Regulation 385-61, Safety, THE ARMY CHEMICAL AGENT SAFETY PROGRAM, Headquarters, Department of the Army, Washington, DC, 12 October 2001.
9. Chemical and Instrumental Verification of Organophosphorus Warfare Agents, Prepared for The Ministry for Foreign Affairs of Finland by The Advisory Board for Disarmament, Helsinki, 1977.
10. Larson L. The alkaline hydrolysis of two sarin analogues and of Tabun. *Acta Chim. Scand.* 1958; 12: 783-785.
11. Larson L. The hydrolysis of dimethylamido-ethoxy-phosphoryl cyanide (Tabun), *Acta Chim. Scand.* 1953; 7: 306-314.
12. Holmstedt B. Synthesis and pharmacology of dimethylamido-ethoxy-phosphoryl cyanide (Tabun). *Acta Chim. Scand.* 1951, Suppl. No. 90, Stockholm, half-lives taken from figure 2, p 35.
13. Desire B, Saint-Andre, S. Interaction of soman with b-cyclodextrin. *Fundam. Appl. Toxicol.* 1986; 7: 646-657.

14. Epstein J, Rosenblatt, DH, Gallacio A, McTeague, WF. Summary report on predicting consequences of chemical disposal operations, Edgewood Arsenal Special Publication 1200-12, January 1973, AD-B955399.
15. Physicians' Desk Reference, 56th edition, p 3026, Medical Economics Company, Montvale, NJ, 2002.
16. Joiner RL, Dill GS, Hobson DW, Olson CT, Feder PI, Menton RG, Kiser, RC, Task 87-35 Evaluating the efficacy of antidote drug combinations against soman or tabun toxicity in the rabbit. Battelle Memorial Institute, Columbus, OH, Contract No. DAMD17-83-C3129. AD A216386, 1989.
17. Capacio BR, Shih T-M. Anticonvulsant action of anticholinergic drugs in soman poisoning. *Epilepsia* 1991; 32: 604-615.
18. McDonough JH, Zoeffel, LD, McMonagle J, Copeland TL, Smith CD, Shih, T-M. Anticonvulsant treatment of nerve agent seizures: anticholinergics vs diazepam in soman-intoxicated guinea pigs. *Epilepsy Research* 2000; 38: 1-14.
19. McDonough JH, McMonagle J, Copeland T, Zoeffel D, Shih T-M. A comparative evaluation of benzodiazepines for control of soman-induced seizures. *Archives of Toxicology* 1999; 73:473-478.