

AD_____

Award Number: W81XWH-06-1-0740

TITLE: ERR Gamma: Does an Orphan Nuclear Receptor Link Steroid Hormone Biogenesis to Endocrine Resistance?

PRINCIPAL INVESTIGATOR: Rebecca B. Riggins, Ph.D.

CONTRACTING ORGANIZATION: Georgetown University
Washington, DC 20057

REPORT DATE: September 2007

TYPE OF REPORT: Final

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE				<i>Form Approved</i> OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE (DD-MM-YYYY) 01-09-2007		2. REPORT TYPE Final		3. DATES COVERED (From - To) 1 SEP 2006 - 31 AUG 2007	
4. TITLE AND SUBTITLE ERR Gamma: Does an Orphan Nuclear Receptor Link Steroid Hormone Biogenesis to Endocrine Resistance?				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-06-1-0740	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Rebecca B. Riggins, Ph.D. E-Mail: rbr7@georgetown.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Georgetown University Washington, DC 20057				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT Estrogen-related receptor gamma (ERRγ) is an orphan nuclear receptor with structural similarities to ERα and ERβ (1). In addition to its ability to transactivate classical and imperfect estrogen response elements (EREs), ERRγ is a potent activator of transcription from steroidogenic factor-1 (SF-1) response elements (SF-1REs). Many genes regulated by SF-1REs control key aspects of cholesterol and fatty acid synthesis, important not only for generation of the plasma membrane but also for the synthesis of steroid hormones (2). In this study, we have investigated whether ERRγ expression and/or activity regulates the level of cholesterol in a pair of breast cancer cell lines – one sensitive to endocrine therapy (SUM44) and the other resistant to endocrine therapy (LCCTam, TAM). We found that endocrine-resistant LCCTam (TAM) cells, which overexpress the orphan nuclear receptor ERRγ, contain significantly greater levels of cholesterol than endocrine-sensitive parental SUM44 breast cancer cells, and that siRNA-mediated knockdown of ERRγ in the resistant TAM cell line significantly reduces cholesterol content. SF-1RE activity is 3-fold higher in TAM cells as compared to SUM44 cells, while expression of an endogenous gene (HMGCS2) that contains a consensus SF-1RE is also significantly overexpressed in TAM cells relative to SUM44 cells. Together, these findings represent an important enhancement in our knowledge of breast cancer biology and potential therapeutic response.					
15. SUBJECT TERMS Estrogen Receptor Alpha, Estrogen Related Receptor Gamma, SF-1RE, Endocrine Resistance, Lobular Breast Cancer					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code)
			UU	39	

Table of Contents

	<u>Page</u>
Introduction.....	5
Body.....	5
Key Research Accomplishments.....	9
Reportable Outcomes.....	9
Conclusion.....	10
References.....	11
Appendices.....	13

Introduction

Estrogen receptors alpha and beta (ER α , ER β) play a central role in the etiology of breast cancer and drive our approach to therapeutic intervention. While these and many other nuclear receptors (NRs) are responsive to specific ligands, there are orphan NRs for which no ligand has yet been identified. The function of these receptors in breast cancer biology is often poorly understood. Estrogen-related receptor gamma (ERR γ) is one orphan NR with structural similarities to ER α and ER β (1). In addition to its ability to transactivate classical and imperfect estrogen response elements (EREs), ERR γ is a potent activator of transcription from steroidogenic factor-1 (SF-1) response elements (SF-1REs). Many genes regulated by SF-1REs control key aspects of cholesterol and fatty acid synthesis, important not only for generation of the plasma membrane but also for the synthesis of steroid hormones (2). Consequently, upregulation of ERR γ may increase the cholesterol production capacity of breast cancer cells poised to proliferate in response to estrogen. This has the potential to promote the development of hormone-refractory breast cancer, and is also likely to attenuate the therapeutic effectiveness of endocrine agents designed to inhibit ER function. In this study, we have investigated whether ERR γ expression and/or activity regulates the level of cholesterol in a pair of breast cancer cell lines – one sensitive to endocrine therapy (SUM44) and the other resistant (LCCTam, TAM) to endocrine therapy.

Body

The original text of the Statement of Work (SOW), all progress associated with this SOW, and any additional data contributing to our overall conclusions are described below.

Months 1-3:

A. Task I – optimize the detection of cholesterol production by SUM44 and TAM cell lines.

Findings: We used a commercially-available cholesterol detection assay that detects both cholesterol and cholesteryl esters. Briefly, SUM44 and TAM cells were seeded in 6-well plastic tissue culture dishes for two to three days before being collected. 5×10^5 to 1×10^6 cells were subjected to lipid extraction using a modified version of the Bligh-Dyer method (3). Dried lipids were resuspended in isopropanol containing 10% Triton-X100 detergent and stored at -20°C until use in the assay. The cholesterol detection assay was colorimetric, allowing comparison of our cell line samples to a standard curve containing known amounts (μg) of cholesterol. Blank (zero control), standard curve, and test reactions containing 1 μl of extracted lipids were performed in triplicate and incubated at 37°C for approximately 45 minutes prior to being read on a spectrophotometer at a wavelength of 550 nm. Results are expressed as mean μg cholesterol per 10^6 cells $\pm$ standard error of the mean (SEM) for a representative experiment taken from at least three independent assays (**Figure 1**). We found that as hypothesized, endocrine-resistant TAM cells do contain modestly, but statistically significantly, more cholesterol than endocrine-sensitive SUM44 cells (2.0 $\mu\text{g}/10^6$ cells vs. 1.5 $\mu\text{g}/10^6$ cells, $p=0.003$).

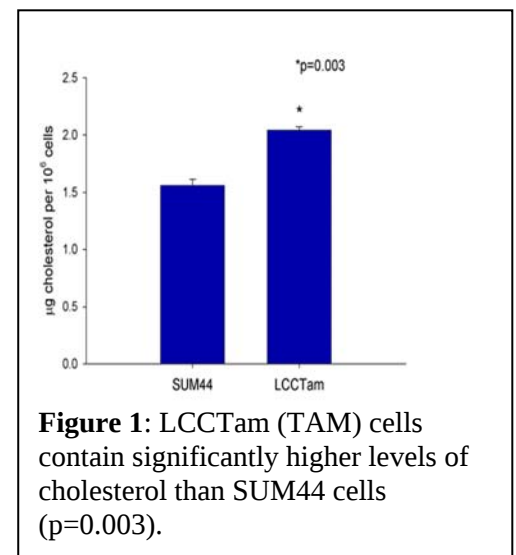


Figure 1: LCCTam (TAM) cells contain significantly higher levels of cholesterol than SUM44 cells ($p=0.003$).

B. Task II – Transfect SUM44 cells with a plasmid encoding the ERR gamma cDNA or the empty vector control, and select both individual clones and pooled populations.

Findings: SUM44 cells were seeded in T-25 plastic tissue culture flasks and co-transfected with 2 μg of a plasmid encoding the murine ERR gamma homologue (ERR3) cDNA tagged at the amino terminus with hemagglutinin antigen (generously provided by Dr. Michael Stallcup, USC, and referred to hereafter as HA-ERR γ) (4) or an empty pSG5 vector, and 0.5 μg of a plasmid encoding the puromycin resistance cassette

(pBABEpuro). ERR3 shares 100% identity with human ERR γ at the amino acid level (4). Forty eight hours after transfection, puromycin antibiotic was added to the culture media as a selection agent and maintained at a concentration of 0.75 μ g/ml. Individual clones and pooled populations were isolated, expanded, and stored in liquid nitrogen until analysis.

Months 4-7:

A. Task II – Analyze the SUM44 clones and pooled populations to determine the level and stability of overexpressed ERR gamma.

Findings: Pooled populations and individual clones of SUM44 cells that had been transfected with HA-ERR γ or pSG5 and stored in liquid nitrogen were cultured, and whole cell lysates were prepared using modified radioimmunoprecipitation assay (RIPA) buffer supplemented with phosphatase and protease inhibitors. Lysates were fractionated by size using SDS-PAGE, transferred to nitrocellulose membranes, and immunoblotted for expression using an antibody specific to the HA tag (HA.11, purchased from Covance). Unfortunately, enhanced chemiluminescent detection revealed no overexpression of HA-ERR γ in any pooled population or individual clone (data not shown).

Faced with this setback, we took the alternative approach of transiently transfecting HA-ERR γ or the empty pSG5 vector into SUM44 and other breast cancer cell lines to determine if we could express an adequate amount of the construct using this technique. In **Figure 2**, we show that indeed, HA-ERR γ is detectable between 24 and 96 hours post-transfection, with a peak in expression at 48 hours. This allowed us to modify our approach to later tasks in this SOW, by using transient rather than stable transfection.

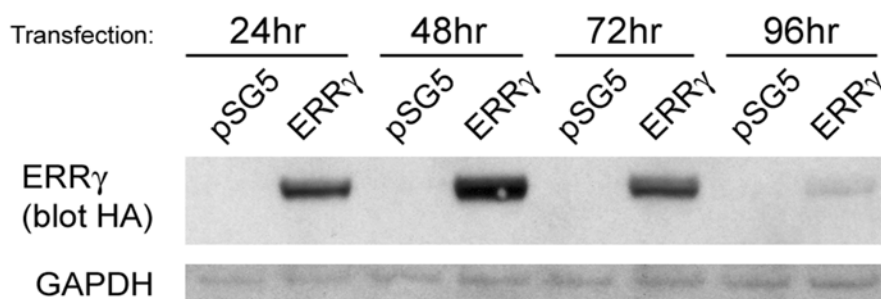


Figure 2: HA-ERR γ expression is detectable by immunoblot following transient transfection. GAPDH was used as a loading control.

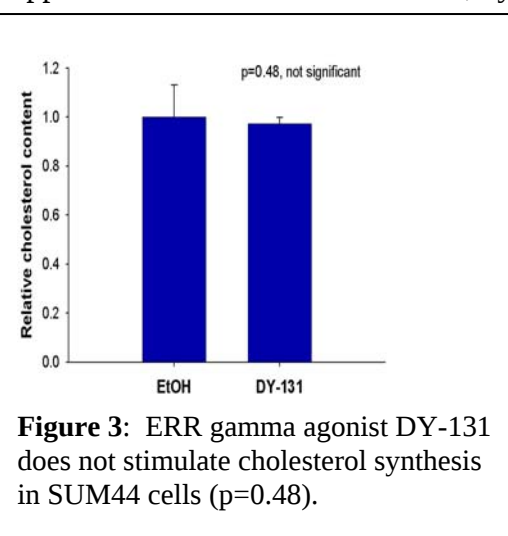


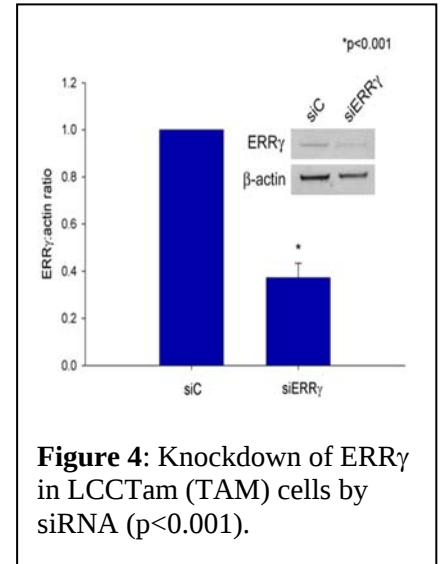
Figure 3: ERR gamma agonist DY-131 does not stimulate cholesterol synthesis in SUM44 cells (p=0.48).

B. Task II – stimulate parental SUM44 cells with ERR gamma agonist DY-131 and perform cholesterol synthesis assays.

Findings: SUM44 cells were seeded in 6-well plastic tissue culture dishes, and stimulated with 1 μ M DY-131 or vehicle control (EtOH) for 72 hours prior to harvesting, lipid extraction, and cholesterol assays as described above. Results are expressed as relative cholesterol content (normalized to EtOH control) $\pm$ SEM for a representative experiment (**Figure 3**). Although 1 μ M DY-131 has been shown to be sufficient to activate ERR γ transcriptional activity in reporter assays (IC₅₀ = 300-500 nM) (5), DY-131 is not able to induce cholesterol synthesis in SUM44 cells.

C. Task III – design and test the efficacy of siRNA against ERR gamma in the TAM cell line.

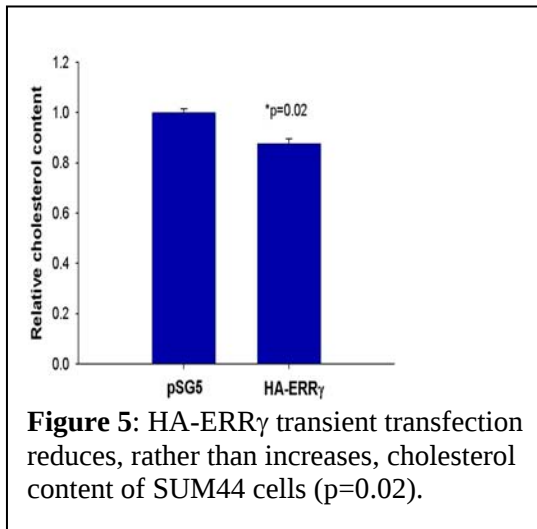
Findings: LCCTam (TAM) cells were seeded into 6-well plastic tissue culture dishes one day prior to transfection with a pre-validated siRNA specific to ERR γ (siERR γ), or a scrambled control siRNA (siC) (oligos and DharmaFECT1 reagent purchased from Dharmacon). Cells were incubated for a further 72 hours prior to lysis and immunoblot analysis as described above using an antibody directed against ERR γ (purchased from GenWay). Densitometric analysis of ERR γ expression relative to expression of the β -actin loading control for four independent experiments showed that ERR γ siRNA is successfully able to knock down ERR γ protein expression by approximately 60% (**Figure 4**, $p<0.001$).



Months 8-11:

A. Task II – perform cholesterol synthesis assays on ERR gamma overexpressing SUM44 clones and pooled populations.

Findings: Because we were unable to isolate clones or pooled

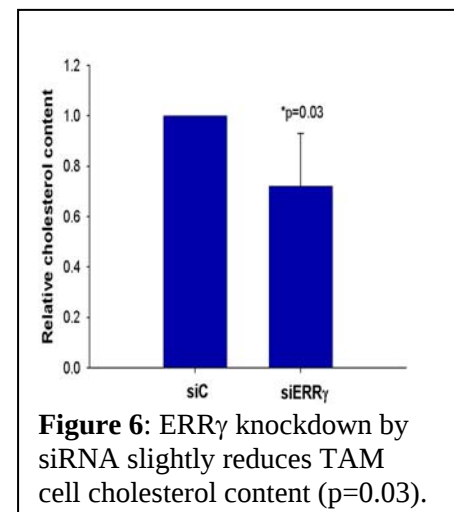


populations of SUM44 cells that stably overexpressed ERR γ , but were able to observe robust expression by transient transfection techniques (**Figure 3**), we chose to do these experiments in a transient manner. SUM44 cells were seeded into 6-well dishes for one day prior to transfection with 1 μ g of either the plasmid encoding the HA-ERR γ cDNA, or the empty pSG5 vector control. 72 hours post-transfection, cells were harvested, lipids extracted, and cholesterol assays performed as described above. Despite good expression of the HA-ERR γ construct after 72 hours of transfection (see **Figure 3**), we did not observe an increase in cholesterol synthesis in HA-ERR γ expressing cells relative to vector control (**Figure 5**). In fact, we saw a slight but significant *reduction* in cholesterol content in cells that were transfected with HA-ERR γ ($p=0.02$). While the meaning of this result is not yet clear, it is important to point out that total levels of cholesterol were, overall,

noticeably lower in cells transfected with HA-ERR γ or pSG5 as compared to mock transfected cells which did not receive the cationic lipid transfection reagent. This may suggest that transfection with this particular reagent (FuGENE6) had some effect on cholesterol production or detection in this context. We did not observe this effect with the siRNA transfection reagent (DharmaFECT1, see below), further suggesting that transfection using FuGENE6 had an unintended impact on cholesterol content or detection.

B. Task III – perform cholesterol synthesis assays in TAM cells transfected with ERR gamma siRNA

Findings: Having established that ERR γ knockdown by the Dharmacon siRNA reagent was successful, we performed the same siRNA experiment outlined above but instead harvested cells to extract lipids and perform cholesterol assays 72 hours post-transfection. Data are presented as mean relative cholesterol content $\pm$ SEM (**Figure 6**). We observed that ERR γ knockdown



slightly, but significantly, reduced cholesterol content in TAM cells ($p=0.03$).

C. Task III – stimulate TAM cells with the ERR gamma antagonist diethylstilbestrol and perform cholesterol synthesis assays.

Findings: LCCTam (TAM) cells were seeded in 6-well plastic tissue culture dishes one day prior to treating with 1 μ M diethylstilbestrol (DES) or vehicle control (DES). 72 hours post-treatment, cells were harvested for cholesterol assays as above (**Figure 7**). We observed no statistically significant increase or decrease in LCCTam cell cholesterol content in the presence of DES ($p=0.2$).

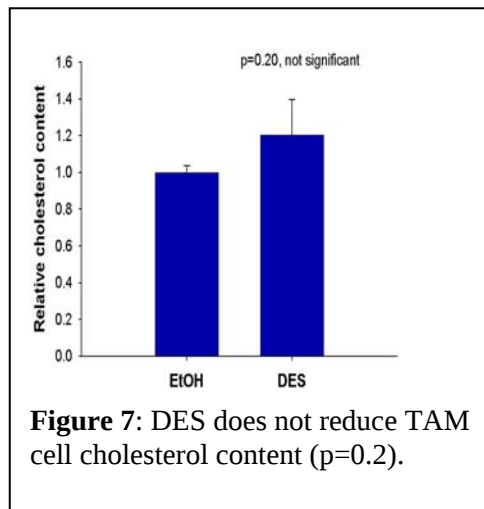


Figure 7: DES does not reduce TAM cell cholesterol content ($p=0.2$).

Month 12:

- A. Complete any unfinished experiments
- B. Begin work on a manuscript for publication

Additional Studies:

- A. Activity of an SF-1RE-driven luciferase reporter in SUM44 and TAM cells.

Rationale: As stated in the original grant application and in the above Introduction section, $ERR\gamma$ is a potent activator of transcription from steroidogenic factor-1 (SF-1) response elements (SF-1REs).

Therefore, we asked whether SF-1RE transcriptional activity is

higher in TAM cells than in SUM44 cells, as would be expected given their overexpression of $ERR\gamma$.

Findings: SUM44 and LCCTam (TAM) cells were seeded into 12-well plastic tissue culture plates one day prior to transfection. Cells were co-transfected with a luciferase reporter construct bearing three tandem copies of the consensus sequence for SF-1RE (TCAAGGTCA; generously provided by Dr. Jean-Marc Vanacker, INSERM, Montpellier, France) (6) and a control plasmid (pRL-SV40-Renilla, purchased from Promega). 48 hours post-transfection, cells were lysed in 1X passive lysis buffer and subjected to dual-luciferase assays performed in triplicate (Promega). SF-1RE-luciferase signal intensity was divided by that of the control Renilla, and data are presented as the mean relative luciferase activity $\pm$ SEM (**Figure 8**). As we hypothesized, LCCTam cells do exhibit approximately 3-fold greater SF-1RE activity as compared to SUM44 cells ($p<0.001$).

- B. Expression of SF-1RE-regulated endogenous genes in SUM44 and TAM cells.

Rationale: Heterologous transcriptional reporters, such as the SF-1RE-luciferase construct used above, are not always representative of transcription at native or endogenous SF-1RE sites. Therefore we re-analyzed our gene expression microarray data showing $ERR\gamma$ overexpression in the resistant TAM cells to look for genes that a.) contain consensus SF-1RE and b.) have been shown to be connected to cholesterol, bile acid, or fatty acid synthesis in the literature.

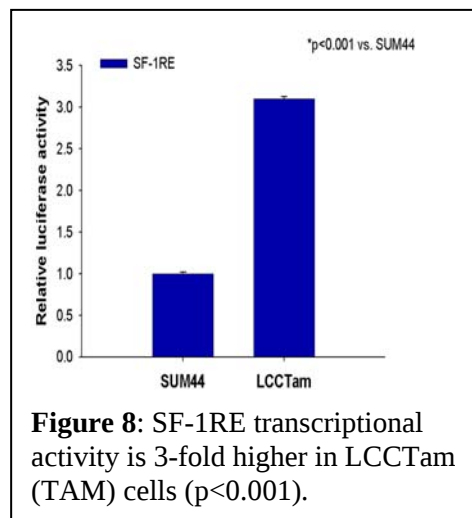


Figure 8: SF-1RE transcriptional activity is 3-fold higher in LCCTam (TAM) cells ($p<0.001$).

Findings: We identified mitochondrial 3-hydroxy-3-methylglutaryl-Coenzyme A synthase 2 (HMGCS2) as

HUGO symbol	Fold-change, Affymetrix ^a	p value	Fold-change, qPCR ^a	p value
HMGCS2	9.58	<0.001	10.32	0.04

Table 1: mRNA expression of HMGCS2 in TAM vs. SUM44 cells.

a – fold change for TAM vs. SUM44.

being significantly upregulated in TAM cells vs. SUM44 cells. **Table 1** shows the fold-change in expression as measured both by microarray and confirmatory quantitative PCR. As determined by promoter analysis using Genomatix MatInspector software (7), HMGCS2 contains a putative SF-1RE sequence in its upstream regulatory regions (sequence atccCAAGgtaac at position -314 to -302

bp). Perhaps more importantly, this enzyme produces HMG-CoA. HMG-CoA is an essential precursor of the cholesterol synthetic pathway, which is synthesized from acetyl-CoA and subsequently reduced to mevalonate. The cytoplasmic form of HMG-CoA synthase (HMGCS1) is thought to be primarily responsible for cholesterol synthesis, and the hamster HMGCS1 gene is known to be positively regulated by SF-1 (8). However, stable transfection of HMGCS2 (the mitochondrial form) in cells lacking HMGCS1 rescues production of cholesterol and promotes mevalonate independence, suggesting that this enzyme can produce cholesterol precursors directly in the mitochondria (9).

Key Research Accomplishments

- Endocrine-resistant LCCTam (TAM) cells, which overexpress the orphan nuclear receptor ERR γ , contain significantly greater levels of cholesterol than endocrine-sensitive parental SUM44 breast cancer cells.
- Neither treatment of SUM44 cells with the ERR γ agonist DY-131, nor of TAM cells with the ERR γ antagonist DES has any effect on cholesterol content of these cell lines.
- While transient overexpression of wild-type ERR γ is not sufficient to stimulate cholesterol synthesis in SUM44 cells, siRNA-mediated knockdown of ERR γ in the resistant TAM cell line does significantly reduce cholesterol content.
- ERR γ is known to activate transcription from SF-1REs, SF-1RE activity is 3-fold higher in TAM cells as compared to SUM44 cells, and expression of an endogenous gene (HMGCS2) that contains a consensus SF-1RE and has been shown to be connected to cholesterol, bile acid, or fatty acid synthesis is also significantly overexpressed in TAM cells relative to SUM44 cells.

Reportable Outcomes

Abstracts:

1.) “Does the orphan nuclear receptor ERR γ link cholesterol synthesis to endocrine resistance?”
Rebecca B. Riggins, Jennifer P-J. Lan, Alan Zwart, and Robert Clarke, Georgetown University
Poster presentation at the 2007 Annual Meeting of the AACR, April 14-18, in Los Angeles, CA

Manuscripts:

1.) “Pathways to Tamoxifen resistance”
Rebecca B. Riggins, Randy S. Schrecengost, Michael S. Guerrero, and Amy H. Bouton
Cancer Letters 256 (2007), pp.1-24.

*This manuscript is a mini-review that summarizes published work on the mechanisms of Tamoxifen resistance independent from the studies funded here, but was written during the granting period and the DoD is acknowledged as a source of funding.

Funding applied for based on this award:

1.) “Estrogen related receptor gamma: a novel biomarker of endocrine resistance in invasive lobular breast carcinoma.”
Submitted to the FY07 DoD Breast Cancer Research Program Idea Grant mechanism.

2.) “Regulation of ERR γ action in endocrine resistant breast cancer by activation of the Erk/MAPK signaling pathway.”

Will be submitted to the Susan G. Komen for the Cure Foundation’s 2007 Career Catalyst Research Grant mechanism, November 2007.

Employment:

Following my receipt of this award from the DoD BCRP, I applied for and was granted a Research-track faculty position in the Department of Oncology and Lombardi Comprehensive Cancer Center at Georgetown University. I currently hold the position of Research Assistant Professor.

Conclusion

There are three major, positive findings from this one-year DoD BCRP Concept award. First, endocrine-resistant LCCTam (TAM) cells, which overexpress the orphan nuclear receptor ERR γ , contain significantly greater levels of cholesterol than endocrine-sensitive parental SUM44 breast cancer cells. Two, siRNA-mediated knockdown of ERR γ in the resistant TAM cell line does significantly reduce cholesterol content. And three, SF-1RE activity is 3-fold higher in TAM cells as compared to SUM44 cells, while expression of an endogenous gene (HMGCS2) that contains a consensus SF-1RE and has been shown to be connected to cholesterol, bile acid, or fatty acid synthesis is also significantly overexpressed in TAM cells relative to SUM44 cells.

Despite the fact that funding for this award is complete, we will continue to actively pursue this research and make several modifications or additions that will complement our current findings. We experienced difficulty producing stable transfectants of HA-ERR γ in SUM44 breast cancer cells. Therefore our next step will be to attempt to produce this cell line again, but instead we will use an adenoviral or retroviral expression vector that expresses this cDNA. Infection rather than transfection is a more rapid and efficient method of gene expression, so we are hopeful that this will be more successful.

The SUM44/LCCTam cell culture model was derived from an invasive lobular breast carcinoma, a distinct histologic subtype of breast cancer that is under-studied despite its rising incidence in Western Europe and the United States (10). Therefore, our data showing that cholesterol content is elevated in an endocrine-resistant variant of this type of breast cancer provides a solid foundation on which to build future studies of resistance that will specifically address risk factors in lobular breast cancer. The function of the orphan NR ERR γ is also poorly understood in breast cancer, although it has been shown to be overexpressed in this context (11), and its family member ERR α 1 has received greater attention (12). Therefore, future studies on the role of ERR γ in breast cancer initiation, progression, and therapeutic response are clearly warranted. Finally, the relevant targets of SF-1RE-mediated gene transcription in breast cancer, and the importance of HMGCS2 in this context, are also not known. However it is interesting to note that in a search of the ONCOMINE database of 42 published gene expression microarray datasets of breast cancer tissue (13), SF-1 (HUGO symbol NR5A1) and HMGCS2 expression are both significantly associated with estrogen receptor alpha-positive status (original microarray data from 14-17), and HMGCS2 is also significantly overexpressed in lobular vs. ductal breast carcinomas (<http://www.intgen.org/expo.cfm>). Together, the knowledge gained during this research represents an important enhancement in our knowledge of breast cancer biology and potential therapeutic response.

References

- (1) Ariazi EA, Jordan VC. Estrogen-related receptors as emerging targets in cancer and metabolic disorders. *Curr Top Med Chem* 2006;6(3):203-15.
- (2) Val P, Lefrancois-Martinez AM, Veyssiere G, Martinez A. SF-1 a key player in the development and differentiation of steroidogenic tissues. *Nucl Recept* 2003 Sep 18;1(1):8.
- (3) BLIGH EG, DYER WJ. A rapid method of total lipid extraction and purification. *Can J Biochem Physiol* 1959 Aug;37(8):911-7.
- (4) Hong H, Yang L, Stallcup MR. Hormone-independent transcriptional activation and coactivator binding by novel orphan nuclear receptor ERR3. *J Biol Chem* 1999 Aug 6;274(32):22618-26.
- (5) Yu DD, Forman BM. Identification of an agonist ligand for estrogen-related receptors ERRbeta/gamma. *Bioorg Med Chem Lett* 2005 Mar 1;15(5):1311-3.
- (6) Vanacker JM, Bonnelye E, Chopin-Delannoy S, Delmarre C, Cavailles V, Laudet V. Transcriptional activities of the orphan nuclear receptor ERR alpha (estrogen receptor-related receptor-alpha). *Mol Endocrinol* 1999 May;13(5):764-73.
- (7) Cartharius K, Frech K, Grote K, Klocke B, Haltmeier M, Klingenhoff A, et al. MatInspector and beyond: promoter analysis based on transcription factor binding sites. *Bioinformatics* 2005 Jul 1;21(13):2933-42.
- (8) Mascaro C, Nadal A, Hegardt FG, Marrero PF, Haro D. Contribution of steroidogenic factor 1 to the regulation of cholesterol synthesis. *Biochem J* 2000 Sep 15;350 Pt 3:785-90.
- (9) Ortiz JA, Gil-Gomez G, Casaroli-Marano RP, Vilaro S, Hegardt FG, Haro D. Transfection of the ketogenic mitochondrial 3-hydroxy-3-methylglutaryl-coenzyme A synthase cDNA into Mev-1 cells corrects their auxotrophy for mevalonate. *J Biol Chem* 1994 Nov 18;269(46):28523-6.
- (10) Li CI, Anderson BO, Daling JR, Moe RE. Trends in incidence rates of invasive lobular and ductal breast carcinoma. *J Am Med Assoc* 2003 Mar 19;289(11):1421-4.
- (11) Ariazi EA, Clark GM, Mertz JE. Estrogen-related receptor alpha and estrogen-related receptor gamma associate with unfavorable and favorable biomarkers, respectively, in human breast cancer. *Cancer Res* 2002 Nov 15;62(22):6510-8.
- (12) Ariazi EA, Kraus RJ, Farrell ML, Jordan VC, Mertz JE. Estrogen-Related Receptor {alpha}1 Transcriptional Activities Are Regulated in Part via the ErbB2/HER2 Signaling Pathway. *Mol Cancer Res* 2007 Jan;5(1):71-85.
- (13) Rhodes DR, Yu J, Shanker K, Deshpande N, Varambally R, Ghosh D, et al. ONCOMINE: a cancer microarray database and integrated data-mining platform. *Neoplasia* 2004 Jan;6(1):1-6.
- (14) Van't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M, et al. Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 2002 Jan 31;415(6871):530-6.

- (15) van de Vijver MJ, He YD, Van't Veer LJ, Dai H, Hart AA, Voskuil DW, et al. A gene-expression signature as a predictor of survival in breast cancer. *N Engl J Med* 2002 Dec 19;347(25):1999-2009.
- (16) Chin K, DeVries S, Fridlyand J, Spellman PT, Roydasgupta R, Kuo WL, et al. Genomic and transcriptional aberrations linked to breast cancer pathophysiologies. *Cancer Cell* 2006 Dec;10(6):529-41.
- (17) Wang Y, Klijn JGM, Zhang Y, Sieuwerts AM, Look MP, Yang F, et al. Gene-expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. *Lancet* 2005 Feb 19;365(9460):671-9.

Appendices

- I. Personnel receiving pay from BC051851 in connection with this research: Rebecca B. Riggins
- II. Abstract presented at the 2007 Annual Meeting of the AACR (1 page).
- III. Cancer Letters mini-review, written during the granting period and in which DoD is acknowledged as a source of funding (24 pages).
- IV. Current *curriculum vitae* (2 pages).

2007 AACR Annual Meeting**April 14-18, 2007****Los Angeles, CA**[Print this Page for Your Records](#)[Close Window](#)

Abstract Number: 990

Presentation Title: **Does the orphan nuclear receptor ERR γ link cholesterol synthesis to endocrine therapy resistance?**

Presentation Start/End Time: Sunday, Apr 15, 2007, 1:00 PM - 5:00 PM

Location: Exhibit Hall, Los Angeles Convention Center

Poster Section: 9

Poster Board Number: 11

Author Block: *Rebecca B. Riggins, Jennifer P-J Lan, Alan Zwart, Robert Clarke. Georgetown Univ., Lombardi Comp. Cancer Ctr., Washington, DC*

Estrogen receptors alpha and beta (ER α , ER β) play a central role in the etiology of breast cancer and drive our approach to therapeutic intervention. While these and many other nuclear receptors (NRs) are responsive to specific ligands, there are orphan NRs for which no ligand has yet been identified. The function of these receptors in breast cancer biology is often poorly understood. Estrogen-related receptor gamma (ERR γ) is one orphan NR with structural similarities to ER α and ER β . In addition to its ability to transactivate classical and imperfect estrogen response elements (EREs), ERR γ is a potent activator of transcription from steroidogenic factor-1 (SF-1) response elements (SF-1REs). Many genes regulated by SF-1REs control key aspects of cholesterol and fatty acid synthesis, important not only for generation of the plasma membrane but also for the synthesis of steroid hormones. Consequently, upregulation of ERR γ may increase the cholesterol production capacity of breast cancer cells poised to proliferate in response to estrogen. This has the potential to promote the development of hormone-refractory breast cancer, and is also likely to attenuate the therapeutic effectiveness of antiestrogens designed to inhibit ER function. We have found that ERR γ is upregulated 4-fold ($p=0.015$) in LCCTam cells, a multi-antiestrogen resistant variant of the ER-positive SUM44 breast cancer cell line, a model of invasive lobular carcinoma. ERR γ overexpression is accompanied by a significant increase in basal and estrogen-stimulated ERE transcriptional activity, and siRNA-mediated knockdown of ERR γ in LCCTam cells completely restores sensitivity to both 4-hydroxytamoxifen and Fulvestrant ($p=0.03$). Furthermore, LCCTam cells produce greater levels of cholesterol than SUM44, and we have found that the expression of three genes with established connections to cholesterol and steroid hormone synthesis is significantly increased in LCCTam: HMG-CoA synthase 2 (HMGCS2, 9.6-fold), GATA binding protein 4 (GATA4, 4.4-fold), and sterol regulatory element binding transcription factor 1 (SREBF1, 1.8-fold). Together these data suggest that ERR γ may affect endocrine resistance in the LCCTam breast cancer model by regulating cholesterol production.

Mini-review

Pathways to tamoxifen resistance

Rebecca B. Riggins^a, Randy S. Schrecengost^b, Michael S. Guerrero^b,
Amy H. Bouton^{b,*}

^a Department of Oncology, Lombardi Comprehensive Cancer Center, Georgetown University, Washington, DC 20057, USA

^b Department of Microbiology and Cancer Center, University of Virginia Health System, Box 800734, Charlottesville, VA 22908-0734, USA

Received 16 January 2007; received in revised form 15 March 2007; accepted 15 March 2007

Abstract

Therapies that target the synthesis of estrogen or the function of estrogen receptor(s) have been developed to treat breast cancer. While these approaches have proven to be beneficial to a large number of patients, both *de novo* and acquired resistance to these drugs is a significant problem. Recent advances in our understanding of the molecular mechanisms that contribute to resistance have provided a means to begin to predict patient responses to these drugs and develop rational approaches for combining therapeutic agents to circumvent or desensitize the resistant phenotype. Here, we review common mechanisms of antiestrogen resistance and discuss the implications for prediction of response and design of effective combinatorial treatments.

© 2007 Elsevier Ireland Ltd. All rights reserved.

Keywords: Tamoxifen; Antiestrogen; Aromatase inhibitor; Fulvestrant; Estrogen receptor

1. Endocrine therapies for breast cancer

Estrogen and the steroid estrogen receptors (ERs) are critical regulators of breast epithelial cell proliferation, differentiation, and apoptosis. Mammals express two ERs, ER α and ER β , which show distinct tissue distributions and functions (for review, see [1]). Mice with targeted deletions of one or both ER genes have established that ER α is the key regulator of mammary gland development. ER α is expressed in 15–30% of the luminal epithelial cells present in normal breast tissue; estrogen stimulation of these cell results in transcription of various genes, including

many that are involved in cell cycle regulation (for review, see [2]). However, estrogen-dependent proliferation of breast epithelial cells is thought to occur in a paracrine fashion, such that ER α -containing cells produce growth factors that induce proliferation in adjacent ER-negative cells. In malignant breast tissue, the action of estrogen is deregulated, resulting in a shift to proliferation without differentiation or apoptosis. The percentage of epithelial cells expressing ER α is significantly increased under these conditions. Moreover, these cells now proliferate, marking a shift from paracrine to autocrine growth.

ER β is more ubiquitously expressed throughout mammary tissue than is ER α [3]. ER β is often found co-expressed with ER α in the luminal epithelial cells but has also been detected in myoepithelial cell and surrounding stromal cell nuclei. While its specific

* Corresponding author. Tel.: +1 434 924 2513; fax: +1 434 982 1071.

E-mail address: ahb8y@virginia.edu (A.H. Bouton).

function(s) in normal and neoplastic tissues remain unknown, ER β is essential for the fully differentiated phenotype of the normal mammary gland in mice [4]. In ER $\beta^{-/-}$ mice, the mammary gland develops normally until puberty. However, several abnormalities are apparent during pregnancy and lactation, including incomplete penetration of the fat pad by glandular tissue, an abnormally large increase in the size of the lumen of ducts and alveoli, and a decrease in the total number of alveoli. Also, in luminal epithelial cells of these mice, the loss of ER β disrupts the formation of tight junctions and alters the expression of β -catenin, features often associated with a malignant phenotype. It has been suggested that the function of ER β in these processes may contribute to the protection conferred by pregnancy and lactation against breast cancer.

Several reports describe a loss of ER β during carcinogenesis (for review see [5,6]), providing support for its possible role as a tumor suppressor. Indeed, ER β has been reported to suppress the function of ER α as a transcriptional activator when both receptors are co-expressed [5,7] and to promote anti-proliferative and pro-apoptotic effects [8]. Several genes have been identified that are regulated by ER β but not ER α ; studies are underway to determine whether these genes may have anti-proliferative or pro-apoptotic properties that could account for the putative tumor suppressor function of ER β [5,9,10].

A majority of invasive breast tumors express ER α , and as discussed above, ER β is often downregulated. These findings provide a strong rationale for therapies that disrupt the actions of estrogen and ER α . Current approaches include inhibiting the function of ERs with the use of antiestrogens such as tamoxifen, raloxifene, and ICI 182,780 (faslodex, fulvestrant), or blocking the conversion of androgens into biologically active estrogens through the action of aromatase inhibitors such as letrozole, anastrozole, and exemestane (see Section 1.1). Recent studies on ER β have also highlighted its growing value as a diagnostic and prognostic marker in breast cancer and its potential for use as a new strategy for delaying and retarding breast tumorigenesis (see Section 1.1.1).

1.1. Classes of therapeutic agents

1.1.1. Selective estrogen receptor modulators (SERMs)

SERMs have been developed as agents that function as ER antagonists in some tissues, including

breast, and agonists in other tissues such as the heart and bone (for review, see [11]). The antagonist effects of tamoxifen in breast tissue are thought to result from its ability to bind to the ligand-binding domain (LBD) of the ER, effectively blocking the potential for estrogen stimulation. Tamoxifen binding further prevents critical ER conformational changes that are required for the association of coactivators. However, tamoxifen treatment is associated with an increased risk of endometrial cancer [12]. Other SERMs, such as raloxifene, have been developed with the goal of reducing some of the deleterious effects of tamoxifen.

New classes of SERMs are being developed that exhibit mixed agonist/antagonist activities that are subtype specific (“dual SERMs”) or preferentially affect one ER over the other (“subtype-selective SERMs”) (reviewed in [13]). However, it is important to note that selectivity is often assessed in the context of binding affinity relative to 17 β -estradiol. Therefore, a “selective” compound may actually interact with both receptors but have a higher affinity for one versus the other. The relative binding affinities of several agents have been determined in HeLa cells engineered to express ER α or ER β [14]. Propyl pyrazole triol (PPT) was shown to be a selective agonist for ER α in this system, while the synthetic ligands diarylpropionitrile (DPN) and 8 β -VE2, and the natural products genistein and biochanin A, were preferential agonists of ER β . In contrast, the antiprogesterin RU486 was shown to be a strong ER β antagonist with only modest agonist activity toward ER α . DPN has been shown to inhibit proliferation and induce apoptosis in the HC11 mouse mammary cell line but this compound has not been widely studied in human breast cancer cell lines [8]. It has also been reported that a modified androstenediol is a potent activator of ER β by ligand-binding assay, but its biological activity in breast cancer cells requires further investigation [15].

Dual SERM compounds that inhibit ER α while activating ER β are particularly attractive, as ER β often performs growth-inhibitory functions in breast tissues and cell lines (see above, Section 1). An example of this type of SERM is TAS-108, a novel, steroidal antiestrogen that inhibits ER α while acting as a partial agonist of ER β ([16] and references therein). *In vitro*, TAS-108 effectively inhibits the growth of tamoxifen-resistant breast cancer cells, and Phase I studies have established that this agent is not only well-tolerated but appears to induce stable disease in women with advanced

breast cancer. Where and how this particular SERM will be used in the clinic, particularly in sequence with the other classes of therapeutic agents described below, has yet to be determined. However, this and future compounds that have similar profiles will be important additions to the current repertoire of endocrine therapies.

1.1.2. Selective estrogen receptor downregulators (SERDs)

The pure ER antagonist ICI 182,780 (hereafter called fulvestrant) binds to ER α with 100-fold greater affinity than does tamoxifen and in so doing, inhibits receptor dimerization and abrogates estrogen signaling [17]. Both laboratory and clinical studies have shown a decrease in overall ER protein levels in response to fulvestrant treatment (discussed in [18]). Importantly, a majority of ER α -expressing tamoxifen-resistant breast cancers remain sensitive to fulvestrant treatment. This has led to its approval in the United States for use in treating ER-positive patients following failure to respond to first-line antiestrogen therapy.

1.1.3. Aromatase inhibitors (AIs)

Aromatase inhibitors (AIs) represent a new class of endocrine therapy drugs that inhibit the action of aromatase, an enzyme necessary for the conversion of androgens to estrogens (for review, see [19]). Clinically, AIs have had success as a second line of therapy for post-menopausal patients who have progressed after tamoxifen treatment. They also show promise as first-line adjuvant treatment for ER α -positive breast cancers.

1.2. Resistance to endocrine therapies

While considerable progress has been made in developing strategies to target estrogen and ERs in breast cancer, approximately 30% of ER α -positive breast cancers do not respond to tamoxifen treatment (*de novo* resistance). In addition, the majority of tumors that initially respond to treatment develop resistance over time (acquired resistance), despite continued expression of ER α (reviewed in [11,20]). Interestingly, many of these resistant tumors still respond to fulvestrant and AIs, indicating that estrogen remains an important regulator of tumor growth under these circumstances [11]. These data provide support for the idea that endocrine-targeted therapies can lead to the activation of novel signaling pathways that circumvent the effects of

antiestrogens [21]. *De novo* and acquired resistance to AIs have also been reported. The appearance and development of resistant tumors are therefore major obstacles in the struggle to control the growth and metastasis of breast carcinomas.

1.3. Biological effects of endocrine therapies

Homeostasis of non-cancerous cells requires a balance between proliferation, cell cycle arrest, and apoptosis; this equilibrium is disrupted during cancer progression. Endocrine therapies induce cell cycle arrest and apoptosis through a reduction in expression of cell cycle regulators such as c-Myc and cyclin D1, accumulation of hypophosphorylated Rb, induction of the cell cycle inhibitors p21^{WAF1/CIP1} and p27^{Kip1}, induction of the putative tumor suppressor interferon regulatory factor-1 (IRF-1), and inhibition of pro-survival pathways through decreased Bcl-2 expression and increased Bax expression ([22–25]; for review, see [20,26]). Breast tumor cells that are resistant to the growth-inhibitory and apoptotic effects of antiestrogens such as tamoxifen overcome these influences through a variety of mechanisms that are the focus of this review.

2. Molecular mechanisms of antiestrogen resistance

Emerging data from breast tumor biopsies indicate that altered expression and/or modification of several growth factor receptors and downstream signaling molecules correlate with tamoxifen resistance (Fig. 1). Epidermal growth factor receptor (EGFR), human epidermal growth factor receptor type 2 (HER2), and insulin-like growth factor-1 receptor (IGF-1R) signaling pathways are often elevated in non-responsive tumors that exhibit either *de novo* or acquired resistance [27–30], as is the activity of kinases that function downstream of these receptors such as extracellular-regulated kinase (ERK) 1/2, p38, AKT, and p21-activated kinase-1 (Pak1) [30–32]. Expression and/or phosphorylation of substrates of these receptor and non-receptor kinases are also elevated in tumors resistant to tamoxifen; such molecules include the cytoplasmic adapter molecule “breast cancer antiestrogen resistance 1” (BCAR1; also known as p130^{Cas}; hereafter called Cas), the coactivator “amplified in breast cancer 1” (AIB1), and ER α itself [28,33–35]. Finally, dysregulated expression of molecules involved in cell survival, such as c-Myc, Bad, and Bcl-2, can also be predictive of a

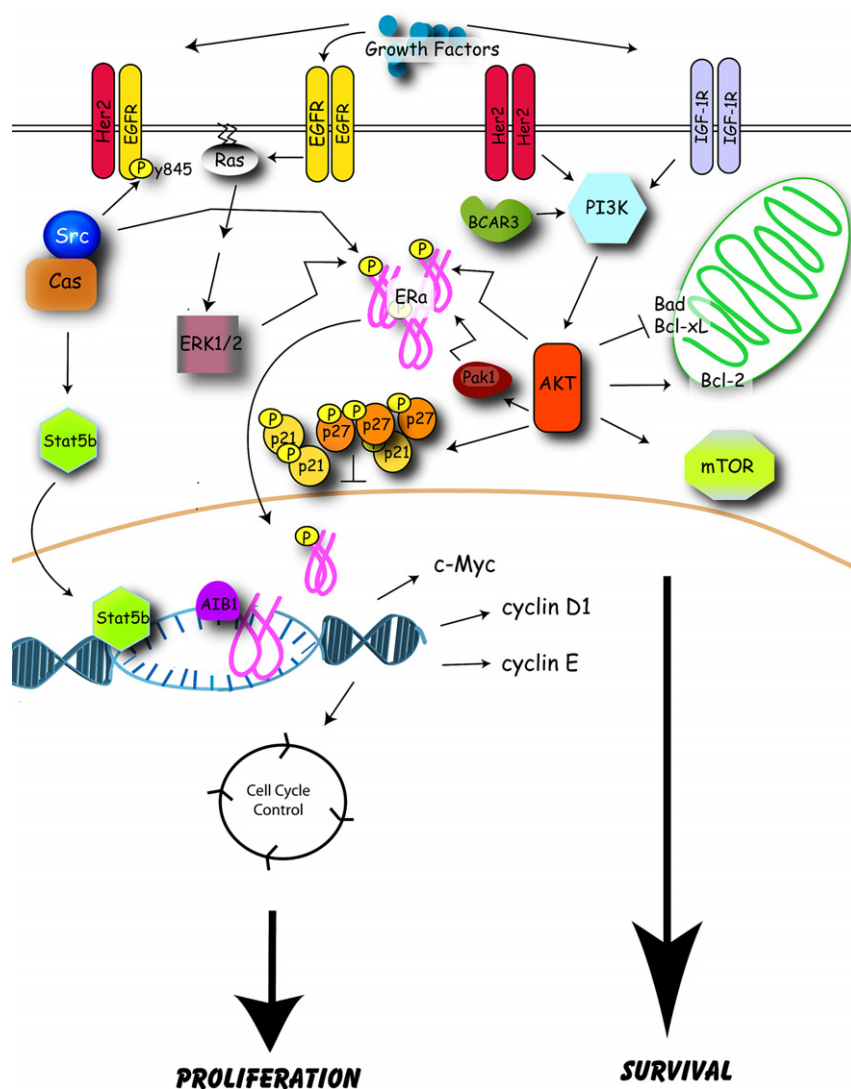


Fig. 1. Molecular mechanisms of tamoxifen resistance. Model depicting molecules implicated in antiestrogen resistance and discussed in this review.

poor response to endocrine therapies such as tamoxifen [36–39]. Clinical data addressing the correlation between some of these markers and antiestrogen resistance are presented in greater detail in Section 4.1. While these studies are limited in both scope and number, however, the association between expression/activation of these molecules and resistance to endocrine therapies has been corroborated (and in fact extended) in tissue culture and animal models. These data are reviewed below.

2.1. *ERα*, *ERβ*, and receptor co-regulatory proteins

Expression of *ERα* has long been considered the primary determinant of a clinical response to endo-

crine or antiestrogen therapy. However in 1996, the concept of estrogen signaling and ER function in breast cancer was significantly altered by the discovery of *ERβ* [40]. The DNA binding domain of full-length *ERβ*1 shares 96% identity with that of *ERα*, and although the affinity of this receptor for estrogen is similar to that of *ERα*, the response to antagonists like tamoxifen and fulvestrant is often different. Both receptors can modulate transcription, either directly from estrogen response elements (EREs), or indirectly through tethering to AP-1 and Sp1 binding sites (reviewed in [41]). However, although *ERα* perceives tamoxifen and fulvestrant as antagonists, it has been shown that *ERβ* can utilize these compounds as agonists when associated

with AP-1 and Sp1 sites [42,43]. Additionally, two naturally occurring variants, ER β 2/ER β cx and ER β 5, contain carboxy-terminal deletions that prevent their binding to ligand, and it has been suggested that these receptors may function as dominant-inhibitory mutants of full-length ER β 1. Murphy et al. have recently reviewed the evidence for a role for ER β 1 and its variants in breast cancer diagnosis and prediction of endocrine therapy response [6].

The transcriptional activities of ER α , ER β , and other steroid hormone receptors are modulated by coregulatory proteins in either a positive or negative fashion. The association of coactivators and corepressors with estrogen receptors is regulated by conformational changes induced by ligand binding, with estrogen inducing coactivator recruitment and antiestrogens leading to association with corepressors. Several excellent reviews have recently detailed the expression, function, and clinical relevance of these coregulators in breast cancer and endocrine resistance [44–47].

2.1.1. ER α expression and downregulation

In human breast tumors, the vast majority of ER α -negative tumors exhibit *de novo* or intrinsic resistance to tamoxifen and other antiestrogens; however, small numbers of ER α -negative but progesterone receptor-positive (PR-positive) tumors respond to antiestrogen treatment (reviewed in [48]). One possible explanation for this finding is that PR-positive tumors retain a functional ER α signaling pathway, particularly since expression of PR is an estrogen-regulated event, and that ER α is present in these tumors at levels that are below the limit of detection for ligand-binding or immunohistochemical assays. The importance of PR as a marker of antiestrogen response is substantiated by studies showing that 70% of ER α -positive/PR-positive tumors effectively respond to tamoxifen, while only 34% of ER α -positive/PR-negative patients respond to tamoxifen therapy [49].

In the context of *de novo* antiestrogen resistance displayed by ER α -negative breast cancer cells, several studies suggest that it may be possible to restore tamoxifen sensitivity through re-expression of ER α . Epigenetic silencing of the ER due to hypermethylation of the promoter region has been reported both *in vitro* and *in vivo* [50,51]. For example, ER α expression can be restored in the ER α -negative MDA-MB-231 breast cancer cell line by either treatment with the demethylating agent 5-aza-2'-deoxy-

cytidine or the histone deacetylase inhibitor trichostatin A [52]. Importantly, when endogenous ER α expression was restored in this way, MDA-MB-231 cells became sensitive to growth inhibition by tamoxifen. Based on data such as these, it has been suggested that restoration of ER α expression may be a viable option for the treatment of some ER-negative breast cancers (discussed in [53]). However, a separate study showed that ectopic expression of ER α in MDA-MB-231 cells was not sufficient to render the cells sensitive to tamoxifen [54]. This suggests that the level and cellular context of ER α expression will be important factors to consider when attempting to modulate ER α levels in “ER-negative” tumors.

Although downregulation of ER α expression can contribute to *de novo* antiestrogen resistance, loss of the receptor during the acquisition of tamoxifen resistance is not commonly observed *in vivo* ([55,56]; reviewed in [57]). However, loss of ER α has been reported in a few models of acquired antiestrogen resistance. For example, the ZR-75-derived 9a1 cell line becomes ER α -negative in the presence of tamoxifen but reverts to being ER α -positive when tamoxifen is removed and estrogen is reintroduced [58].

Like tamoxifen, fulvestrant inhibits ER α transcriptional activity. In addition, fulvestrant has also been shown to accelerate ER α protein degradation via the ubiquitin–proteasome pathway, leading to its reclassification as a SERD (see above). Loss or downregulation of ER α has the potential to be a viable mechanism for antiestrogen resistance in the context of acquired resistance to fulvestrant or other SERDs, but current evidence to this effect is contradictory. ER α protein is completely lacking and only minimal mRNA is detectable in several fulvestrant-resistant cell lines, including MCF-7-r, T47D-r, and ZR-75-r cells [59]. MCF-7-F resistant cells show a 90% decrease in both ER α mRNA and protein, accompanied by significant changes in estrogen-induced expression of ER target genes [60]. In contrast, the MCF-7/182^R-1, -6, and -7 models of fulvestrant resistance maintain ER α expression and function, although its expression at the protein level is reduced by one-third as compared to the parental cell line [61,62]. MCF-7/LCC9 breast cancer cells, which display acquired resistance to fulvestrant and cross-resistance to tamoxifen, show no downregulation of ER α [63]. This cell line was originally derived from the estrogen-independent MCF-7/LCC1 cell line [64]; interestingly both MCF-7/

LCC1 and MCF-7/LCC9 cells express higher levels of ER α mRNA and protein than the parental MCF-7 cell line [65]. MCF-7/LCC9 cells have also been shown to contain increased basal ER α transcriptional activity that is not inhibited by fulvestrant [66]. Factors that drive the differential expression and activity of ER α among these cell culture models of fulvestrant resistance are not known, but one likely explanation for these disparities may be the distinct schemes of drug selection that have been used to generate resistant cells *in vitro*. Clinical studies confirm that fulvestrant can reduce ER α levels following either short- (21 days) or long- (6 months) term treatments, but that expression is not completely eliminated (reviewed in [67]). Interestingly, patients who do not show a clinical response to fulvestrant treatment also do not show a decrease in ER α expression.

2.1.2. ER α mutation

Mutation of the ER may also impact endocrine therapy responsiveness. For example, one tamoxifen-resistant variant of the ER α -positive T47D cell line, T47D_{CO}, has acquired three ER α deletion mutations that affect the hormone- and DNA-binding domains of the receptor [68]. This cell line also expresses different levels of PR [69], suggesting that these variants are functionally distinct from the wild type receptor. These and many other deletions, exon duplications or insertions, point mutations, and alternative splicing events have the potential to regulate ER α function in breast and other tissues. Herynk and Fuqua have recently published a review of known ER mutations and their connection to human disease, several of which are linked to tamoxifen response and resistance [70]. For example, duplication of exons 6 and 7 in an estrogen-independent variant of the MCF-7 cell line leads to the production of an ER that can no longer bind to estrogen or tamoxifen [71]. Short insertions in exon 6 have also been identified in tamoxifen-resistant tumors [72]. A tyrosine-to-asparagine substitution at residue 537 (Y537N), originally identified in metastatic breast cancer, confers constitutive transcriptional activity *in vitro* and renders the ER insensitive to both estrogen and tamoxifen, while mutation of this tyrosine to either serine or alanine results in a constitutively active receptor whose activity is still inhibited by tamoxifen and fulvestrant [73]. In contrast, a lysine-to-arginine substitution at residue 303 (K303R) that

was first identified in breast hyperplasia and a number of breast tumors results in a receptor that is hypersensitive to substantially lower concentrations of estrogen than normal (10^{-12} vs. 10^{-9} M) [74]. The prevalence of this mutation in patient populations is unclear, as more recent studies have not found ER α K303R in other patient groups [75]. The difficulty in identifying functional ER α mutations *in vivo* that affect tamoxifen therapy response may stem from the fact that such mutations are relatively rare, estimated to be present in only 1% of breast tumors [51].

2.2. Growth factor receptors

As discussed above, tumors resistant to tamoxifen often show high expression and/or activation of EGFR, its close relative HER2, IGF-R1, and proteins that function downstream of these receptors such as ERK1/2, and AKT (for review, see [28,76–78]). *In vitro* data corroborate a role for these growth factor receptors in mediating antiestrogen resistance, both through crosstalk with the ER and through independent pathways.

2.2.1. EGFR and HER2

Several studies have established that overexpression of EGFR or HER2 in ER-positive, antiestrogen-sensitive, breast cancer cells can confer resistance to these drugs [79–81]. Both ERK1/2 and AKT activity appear to contribute to the resistant phenotype under these conditions. Other groups have isolated antiestrogen-resistant cell lines following long-term treatment of ER-positive breast cancer cells with either tamoxifen or fulvestrant. In these studies, acquisition of resistance often coincides with overexpression of the EGFR and activation of downstream molecules such as ERK1/2, PI3K, and AKT [59,63,82,83]. Expression of one of the ligands for EGFR, transforming growth factor α (TGF α), is elevated in one of these cell models, resulting in an autocrine loop that is required for maintenance of the resistant phenotype [84]. Unlike the parental cells, which are relatively impervious to EGFR signaling pathways, the resistant clones are highly dependent on these pathways for growth and survival.

While the mechanism(s) by which these receptors promote antiestrogen resistance are not clear, several lines of evidence suggest that they may function to provide survival signals that override apoptotic programs in the cell. First, when EGFR functions

are blocked by either the tyrosine kinase inhibitor Iressa or blocking antibodies, apoptosis is induced [85–88]. Second, the expression and/or activity of several anti-apoptotic molecules such as AKT, Bcl-2 and Bcl-xL are upregulated under conditions of HER2 overexpression in MCF-7 cells [89,90]. However, events considered in other contexts to be generally pro-apoptotic are also associated with overexpression of EGFR or HER2 in human tumors. For example, elevated levels of phosphop38 have been reported in HER2-expressing tumors as well as in tamoxifen-resistant xenografts [29,30]. While its role in regulating apoptosis under these conditions is not clear, activated p38 may promote resistance to tamoxifen through its ability to enhance the nuclear functions of ER [91]. Finally, recent data from our group indicate that the EGFR may work together with the non-receptor protein tyrosine kinase c-Src and the adapter molecule Cas to promote survival [92].

In addition to the direct effect of activated growth factor receptors on cell proliferation and survival, crosstalk between these receptors and ER plays a significant role in regulating the cellular response to antiestrogens. Signaling through EGFR and HER2 can result in downregulation of ER α expression in cultured cells [93]. This inverse relationship has been corroborated in a recent clinical study by the Southwest Oncology Group [27]. Not only does growth factor signaling affect ER levels, but it can also strongly impact ER nuclear functions, at least in part through promoting phosphorylation of the ER, its coactivators, and/or its corepressors (for review, see [94]).

Conversely, ER α appears to regulate growth factor receptor activities through rapid, non-genomic functions. This may be augmented by an accumulation of cytoplasmic ER α at the expense of nuclear ER α under conditions of high HER2 expression [95]. Interestingly, it appears that tamoxifen may be able to function as an agonist for these non-genomic ER activities under some circumstances, perhaps explaining how cells expressing high levels of HER2 and the coactivator AIB1 show increased proliferation in the presence of both estrogen and tamoxifen [90]. Additional support for this crosstalk comes from preclinical studies showing enhanced efficacy and/or delays in the appearance of resistance in the presence of combined therapies that target both tyrosine kinase receptors and estrogen signaling ([87,96]; and see below, Section 4.3).

2.2.2. IGF-1R signaling

The involvement of IGF signaling in antiestrogen resistance is less clear than is the role for EGFR and HER2. While data are mixed about whether IGF-1R levels are elevated or decreased in resistant tumors and cell lines [28,97,98], evidence for a role of IGF-1R signaling in acquired resistance is emerging. This may be mediated at least in part through crosstalk with EGFR, HER2, and/or ER α and likely involves the scaffolding molecule insulin receptor substrate 1 (IRS-1) [77,99,100].

2.3. Cas/c-Src/BCAR3

Numerous cytoplasmic proteins whose functions are coupled to growth factor receptors have been linked to antiestrogen resistance, further supporting a role for these pathways. Two of these molecules, Cas and “Breast Cancer Anti-Estrogen Resistance 3” (BCAR3, also known as AND-34 and Nsp2), were identified in a screen designed to isolate proteins capable of inducing antiestrogen resistance in sensitive MCF-7 and ZR-75-1 cells [101,102]. Interestingly, both of these molecules are expressed and interact with one another in breast cancer cells (Schrecengost and Bouton, unpublished data), supporting a role for this molecular pathway in the promotion of resistance.

2.3.1. Cas and c-Src

Cas functions in a wide variety of cellular processes, including proliferation, survival, cell adhesion, and migration [103,104]. Cas overexpression promotes increased proliferation and inhibition of apoptosis in MCF-7 breast cancer cells treated with tamoxifen [92,105]. This requires the association of Cas with c-Src, as documented by the finding that, unlike wildtype Cas, overexpression of a Cas mutant that does not interact with c-Src does not promote growth in the presence of tamoxifen [92]. Interactions between c-Src and Cas result in activation of c-Src catalytic activity, promotion of serum- and anchorage-independent growth, and enhancement of cellular migration [106,107]. Interestingly, two substrates of c-Src, Tyrosine 845 (Tyr-845) on the EGFR and signal transducer and activator of transcription (STAT) 5b, have been implicated in Cas-dependent tamoxifen resistance [92]. These data suggest a mechanism whereby c-Src activation by Cas results in the phosphorylation of a number of c-Src substrates and subsequent activation of various pro-proliferative and survival

pathways that allow the cells to grow in the presence of tamoxifen. Interestingly, ER nuclear functions remain unaffected by this pathway [92], suggesting that therapies that target the Src-Cas-EGFR-STAT5b signaling axis may be able to synergize with tamoxifen for the treatment of “resistant” tumors.

Several other studies provide additional insight into how c-Src and Cas may contribute to tamoxifen resistance. First, ER α and phosphatidylinositol 3-kinase (PI3K) have been found in the c-Src/Cas complex in T47D breast cancer cells following rapid estrogen stimulation [108]. While it is clear that these complexes help regulate estrogen-dependent cell proliferation and survival, however, it is not yet known whether they are important for promoting resistance to antiestrogens. Second, pharmacological inhibition of c-Src in MCF-7 cells enhances the growth-inhibitory and apoptotic-inducing effects of tamoxifen, further strengthening a role for c-Src in this process [109]. Finally, cells treated over a period of two weeks with tamoxifen or fulvestrant show a c-Src-dependent increase in Cas phosphorylation that correlates with activation of a survival pathway involving AKT [110].

Together, these studies point to a key role for Cas and c-Src in tamoxifen resistance in cell culture. Whether these pathways also play a role in clinical resistance remains to be determined. c-Src/Cas interactions are favored under conditions in which both proteins are expressed at high levels [106]. Thus it is intriguing that a large percentage of human breast tumors contain high levels of both c-Src and Cas, and elevated c-Src kinase activity [33,111,112]. Moreover, c-Src/Cas complexes can be isolated from a majority of breast cancer cell lines (Schrecengost and Bouton, personal communication). This suggests that one of the mechanisms by which c-Src activity may be elevated in breast cancer cells is through its association with Cas. Cas overexpression in human tumors correlates with poor overall and relapse-free survival and a poor intrinsic response to tamoxifen treatment [33]. The majority of the Cas-overexpressing tumors were shown to be ER-positive, suggesting that Cas promotes *de novo* resistance to tamoxifen through mechanisms that do not involve downregulation of the ER. In contrast, Cas expression levels do not appear to correlate with acquired tamoxifen resistance [113]. c-Src and the EGFR are also frequently (~30%) co-overexpressed in breast tumors, while 20–30% co-overexpress HER2 and c-Src [114,115]. Based on these data, it is reasonable to postulate that, in

at least a subset of tumors, tamoxifen resistance may be mediated through a coordinated mechanism involving these receptor tyrosine kinases, c-Src, and Cas.

2.3.2. BCAR3/AND-34

BCAR3/AND-34 (hereafter called BCAR3) is a Cas binding partner that was identified in a genetic screen along with Cas as a molecule whose overexpression conferred antiestrogen resistance [102]. Since its discovery, BCAR3 has been implicated in altering the activity, expression, and intracellular location of many proteins, including Cas, Ras and Rho family GTPases, and Pak1 ([107,116–118]; for review, see [119]). BCAR3 overexpression induces antiestrogen resistance through a process that involves PI3K and Rac1 [118]. Estrogen-independent growth can also be induced by BCAR3 expression, a process that requires R-Ras activation of AKT [120]. While these data suggest that BCAR3 overexpression may promote antiestrogen resistance in cultured cells, however, it is yet to be determined whether this molecule contributes to resistance in the clinical setting.

2.4. PI3K/AKT

Downstream of growth factor receptor and adapter protein activation, signaling intermediates such as AKT play an important role in cell proliferation, survival, and endocrine resistance. AKT, also known as protein kinase B, is activated by PI3K, leading to substrate phosphorylation and cell survival. AKT activity is subsequently counter-balanced by the tumor suppressor PTEN. Historically, all three isoforms of AKT have been considered oncogenic, although recent studies have reported that different isoforms may play distinct roles in invasion and cell migration (discussed in [121]). AKT is most often considered to be a survival factor, preventing apoptosis under conditions of cell stress. However, it has become increasingly clear that AKT also participates in cell cycle regulation (reviewed in [122]). Cell cycle proteins with established connections to antiestrogen resistance, including those that are AKT targets, will be detailed in Section 2.6.

In ER-positive breast cancer cell lines, PI3K-mediated AKT activation is linked to rapid phosphorylation of ER α and ligand-independent receptor activation [123]. AKT can also stimulate ER β transcriptional activity and enhance

coactivator recruitment to this receptor [124]; whether this has an impact on antiestrogen resistance has yet to be determined. Overexpression of constitutively active AKT in breast cancer cell lines can induce estrogen independence and resistance to tamoxifen and fulvestrant [125], while inhibition of PI3K or AKT restores tamoxifen sensitivity [126]. In addition to endocrine agents, activated AKT can also confer resistance to conventional chemotherapeutics such as doxorubicin [127]. The link between AKT and tamoxifen resistance is not solely observed in cell lines engineered to overexpress this kinase; MCF-7 cells selected for resistance to tamoxifen exhibit upregulated endogenous AKT activity that is required for antiestrogen resistance [128,129]. However, a separate study showed that an aromatase-expressing MCF-7 cell line (MCF-7/Ca), when grown as xenografts in nude mice, did not upregulate the PI3K/AKT pathway in the process of acquiring resistance to the aromatase inhibitor letrozole [130].

The mechanisms by which AKT may confer antiestrogen resistance are varied. Phosphorylation of the ER on serine 167 is discussed below (Section 2.5). AKT can also activate the mammalian target of rapamycin (mTOR), which controls cap-dependent translation, cell growth, and survival (recently reviewed in [131]). This can occur in response to activation of AKT by growth factor receptors such as EGFR and IGF-1R; AKT then phosphorylates and inactivates the tuberous sclerosis complex, which normally keeps mTOR kinase activity in check. Rapamycin and its analog CCI-779 are able to restore tamoxifen sensitivity in MCF-7 cells overexpressing constitutively active AKT [132]. Another mTOR inhibitor (RAD-001) has also been shown to inhibit the growth of wild type MCF-7 cells, as well as MCF-7/Aro cells that stably

over-express aromatase [133]. Moreover, a combination of the aromatase inhibitor letrozole with RAD-001 was shown to more effectively inhibit MCF-7/Aro proliferation than either drug alone. However, given that MCF-7/Ca cells with acquired resistance to letrozole do not contain elevated levels of activated AKT [130], it would seem that a definitive role for the PI3K/AKT/mTOR pathway in aromatase inhibitor resistance remains to be determined. Nonetheless, mTOR and rapamycin-like drugs that target mTOR are considered to be attractive molecular targets and therapeutic agents, respectively, for treating breast cancer (reviewed by [134,135]; see Section 4.3).

2.5. ER phosphorylation

The estrogen receptor is a target of serine/threonine as well as tyrosine phosphorylation (reviewed in [136]). While ER phosphorylation is a key element of non-genomic or rapid estrogen actions, several studies have shown that it can also affect conventional, transcription-mediated events. Reported ER phosphorylation sites include tyrosine 537, serines 104, 106, 118, 167, 236, and 305, and threonine 311 (Fig. 2, and discussed in [137]).

Tyr-537, which is located in the activation function-2 (AF-2) domain, is phosphorylated by Src family kinases [138]. Mutation of Tyr-537 to phenylalanine increases basal ER transcriptional activity without changing the DNA binding capacity of the receptor [139]. Mutation of Tyr-537 to either serine or alanine also results in estrogen-independent transcription, but both 4-hydroxytamoxifen and fulvestrant inhibit this activity [140,141]. Although the full function(s) of Tyr-537 phosphorylation are still under investigation, the evidence that “constitutively active” Tyr-537 mutants are still inhibited

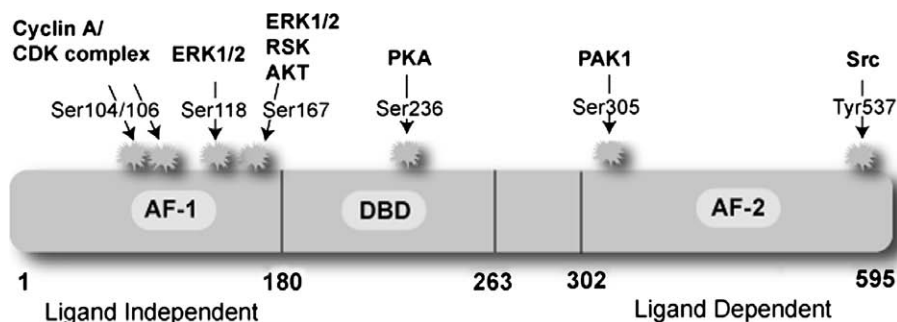


Fig. 2. ER α phosphorylation. Domain structure of ER α showing phosphorylation sites and the kinases that mediate these phosphorylations.

by antiestrogens argues against a critical role for this residue in endocrine therapy resistance. It has also yet to be determined whether Tyr-537 phosphorylation is observed in breast tumors.

The effect of serine phosphorylation on ER functions is better established than is tyrosine phosphorylation and it is more likely to contribute to the resistant phenotype. Ser-104 and Ser-106 are located in the AF-1 domain and are targets of the cyclin A/CDK2 complex; mutation of these sites to alanine prevents cyclin A-mediated enhancement of ER transcriptional activity [142]. Conversely, cyclin A enhances ER transcriptional activity in the presence of tamoxifen. Together, these data suggest that inappropriate cyclin A/CDK2-mediated phosphorylation of Ser-104 and Ser-106 may play a role in endocrine resistance in breast tumors. Cyclin A has been implicated as an independent prognostic factor for poor outcome in tamoxifen-treated breast cancer patients [143].

Ser-167, also located within the AF-1 domain, was initially proposed to be phosphorylated in response to estrogen binding [144]. However, subsequent studies have revealed that its phosphorylation is ligand-independent, and that this site is a major target of ERK, RSK, and AKT kinases (reviewed in [136]). Overexpression of either EGFR or AKT increases Ser-167 phosphorylation and reduces tamoxifen sensitivity, whereas RNAi-mediated inhibition of AKT abrogates Ser-167 phosphorylation and restores tamoxifen sensitivity [145]. *In vitro*, AKT-mediated phosphorylation of Ser-167 can increase ER binding to DNA and enhances the interaction of ER with the coactivator SRC3 in the presence of estrogen [146]. Despite the evidence that Ser-167 phosphorylation appears to play an important role in promoting antiestrogen resistance in cell culture, however, clinical evidence of a definitive role for phosphorylated Ser-167 in antiestrogen resistance is less convincing (see Section 4.1).

Ser-236, located within the DNA binding domain of the receptor, has been shown to be phosphorylated by protein kinase A (PKA) [147]. While phosphorylation at other sites seems to positively regulate ER transcriptional activity, PKA phosphorylation at Ser-236 appears to inhibit receptor dimerization in the absence of ligand. This suggests that Ser-236 represents a negative regulatory site that has yet to be implicated in the process of tamoxifen resistance.

Ser-305 is located in the hinge region of the receptor, and its phosphorylation by Pak1 has

been shown to increase ER transcriptional activity [148]. Substitution of glutamic acid for serine at this site recapitulates increased receptor activation and induces expression of the ER target gene cyclin D1 [149]. Pak1 is a well-known mediator of cell survival and migration, and has itself recently been shown to contribute to tamoxifen resistance. Pak1 overexpression in MCF-7 cells enhances cyclin D1 expression and prevents tamoxifen-mediated inhibition of ER activity [150,151]. Conversely, lower expression of Pak1 in primary breast tumors is associated with improved disease-free survival in patients treated with tamoxifen [32]. Pak1 co-immunoprecipitation with ER is enhanced by tamoxifen treatment of resistant cell lines [150,151]. Pak1-mediated stimulation of ER transcriptional activity through Ser-305 is reported to require processive phosphorylation of Ser-118, suggesting that these two sites work together to regulate ER activity. Interestingly, Ser-305 can also be phosphorylated by PKA. While PKA-mediated phosphorylation of Ser-236 appears to negatively regulate receptor activities (see above), phosphorylation of Ser-305 by PKA is thought to prevent tamoxifen from inducing the inactive conformation of the ER [152].

Ser-118 is perhaps the best studied site of ER phosphorylation and is widely considered to be a target of ERK1/2, although other kinases such as the TFIIF cyclin-dependent kinase, cyclin-dependent kinase 7 (CDK7), and glycogen synthase kinase-3 (GSK-3) may also phosphorylate this site [153–156]. Interestingly, estrogen-induced phosphorylation of Ser-118 occurs within 10 min of stimulation, and this can be inhibited by 4-hydroxytamoxifen but not fulvestrant [155]. Recent studies by Likhite et al. have helped to clarify the function of Ser-118 phosphorylation in tamoxifen-sensitive and resistant cells with respect to hormone, DNA, and coregulator binding [146]. Stable overexpression of HER2 in MCF-7 cells, which induces tamoxifen resistance, also results in increased basal phosphorylation of Ser-118 that is no longer enhanced by estrogen stimulation. Using purified wild type and serine-mutated ER in *in vitro* kinase assays, this group subsequently demonstrated that phosphorylation of Ser-118 by ERK1/2 specifically enhances recruitment of the coactivator SRC3 to estrogen-bound receptor, but has no effect on the affinity of ER for estrogen or DNA. In contrast, phosphorylation at Ser-118 reduced the association of

ER with trans-hydroxytamoxifen, which has the potential to attenuate the growth inhibitory effects of this antiestrogen.

The role of Ser-118 phosphorylation in antiestrogen resistant cell culture models is not fully understood. Kuske et al. have recently reported that estrogen-induced Ser-118 phosphorylation levels are comparable in parental MCF-7 cells and the estrogen-independent MCF-7/LCC1 cell line, but that basal and estrogen-induced levels of phospho-Ser-118 are significantly reduced in the antiestrogen-resistant MCF-7/LCC9 cell line [65]. In studies by other investigators, baseline phosphorylation of Ser-118 is increased in tamoxifen-resistant MCF-7 cells as compared to the parental cell line and is reduced by MEK and EGFR tyrosine kinase inhibitors, which also inhibit cell growth [157]. Finally, Weitsman et al. suggest that, although both HER2 overexpression and activation of the MAPK pathway can induce antiestrogen resistance, neither has any effect on Ser-118 phosphorylation of ER in MCF-7 cells [158]. Instead, they suggest that the activity of I κ B kinase- α (IKK α) is involved. A potential role for IKK α in ER phosphorylation has only recently been suggested [159], and further studies are needed to clarify its role in antiestrogen resistance.

2.6. Cell cycle regulators

Cyclins, cyclin-dependent kinases (CDKs), and CDK inhibitors are the major regulators of cell cycle progression. Antiestrogens like tamoxifen induce cell cycle arrest during the G1 phase; therefore, genes that control cell cycle progression have the potential to significantly impact drug sensitivity and resistance. In breast cancer, normal expression and function of cyclins D1 and E, and the CDK inhibitors p21 and p27, are often specifically disrupted. Caldon et al. have recently published a comprehensive review of cell cycle control in breast cancer that summarizes how cyclins and CDK inhibitors influence cancer progression and may serve as markers of treatment outcome [160]. Butt et al. have also published a summary of the literature concerning the role of cyclins D1 and E in endocrine resistance [161]. Given the timeliness of these reviews, we will only summarize key information about these molecules and focus on the most recent literature in this area.

2.6.1. Cyclin D1

Cyclin D1 is a well-studied contributor to antiestrogen resistance in breast cancer. Essential for early progression through the G1 phase, cyclin D1 both modulates CDK4/6-dependent phosphorylation of the tumor suppressor Rb and sequesters the CDK inhibitors p21 and p27 [160,162]. Cyclin D1 is a direct transcriptional target of estrogen signaling, where its induction is mediated by ER/AP-1 and ER/Sp1 transcription complexes; conversely, antiestrogens like tamoxifen inhibit cyclin D1 expression (discussed in [161]). AKT can indirectly affect cyclin D1 expression by phosphorylating and inhibiting the activity of GSK3 β , which normally induces cyclin D1 degradation [163]. In cell culture, stable overexpression of cyclin D1 can confer resistance to tamoxifen as well as fulvestrant [164]. Moreover, some cells that have acquired resistance to tamoxifen show increased expression of cyclin D1 [165]. However, these cells are still growth-inhibited by fulvestrant.

Recent studies have shed some light on potential mechanisms of cyclin D1 upregulation in the context of antiestrogen resistance, some of which involve crosstalk between growth factor receptors and other signaling molecules. Tamoxifen-resistant MCF-7 cells overexpressing HER2 upregulate cyclin D1 [90]. Similarly, the tamoxifen-resistant phenotype of MCF-7 cells overexpressing Pak1 is partly due to the increased expression of cyclin D1 [32]. BCAR3 overexpression in MCF-7 also induces cyclin D1 promoter activity, coincident with the adoption of antiestrogen resistance [117]. In some instances, tamoxifen has been reported to stimulate breast cancer, although this has been observed more widely in cell culture and xenograft studies than in the clinic. Under conditions in which tamoxifen or raloxifene functions as an ER agonist, cyclin D1 regulates cell cycle progression much as it does in parental MCF-7 cells stimulated by estrogen. For example, cyclin D1 expression was found to be significantly elevated in raloxifene-stimulated MCF-7 xenografts [166]. Treatment with the HER2 inhibitor trastuzumab decreased cyclin D1 expression in these xenografts and concomitantly inhibited tumor growth. Similarly, cyclin D1 was shown to be essential for cell cycle progression induced by tamoxifen in resistant MCF-7 cells grown in tissue culture [167].

Evidence for a role of cyclin D1 in clinical endocrine resistance is still emerging (reviewed in [160]). Cyclin D1 is overexpressed in ~50% of breast

cancers. Some studies have reported that cyclin D1 overexpression is more often observed in highly differentiated, slower growing tumors than in highly proliferative tumors. Other studies report that cyclin D1 amplification is more frequent in ER-positive tumors, whereas overexpression in the absence of amplification is more common in ER-negative tumors [168]. Not surprisingly, it is in the ER-positive cases where cyclin D1 amplification is associated with a worse outcome. How these findings will affect the clinical management of ER-positive breast tumors and future use of tamoxifen therapy remains to be determined.

2.6.2. Cyclin E

In response to inhibition of Rb by cyclin D1 early in G1 phase, the transcription factor E2F strongly induces the expression of cyclin E, which associates with CDK2 to form an active complex that promotes entry into S phase [160]. Cyclin E/CDK2 activity is subsequently counterbalanced by the CDK inhibitors p21 and p27. While not a direct target of the ER, estrogen-dependent induction of c-Myc can activate cyclin E complexes by stimulating release of the inhibitor p21. CDC25A can also participate in estrogen-mediated cyclin E activation [169].

The contribution of cyclin E to tamoxifen resistance is less clear than for cyclin D1. This may be due to the presence of multiple low molecular weight forms of cyclin E that are generated by post-translational proteolytic cleavage events (reviewed in [170]). Low molecular weight cyclin E variants, which exhibit greater activity than full-length cyclin E, can induce antiestrogen resistance in MCF-7 cells [171]. Further studies are required to clarify the mechanism(s) by which low molecular weight cyclin E is induced in breast cancer, and its function in antiestrogen resistance. Other groups have reported that full-length cyclin E can also confer resistance to tamoxifen and fulvestrant [164,172].

2.6.3. p21 and p27

The CDK inhibitors p21 and p27 are important negative regulators of cell cycle progression that counteract the activities of cyclin D1 and cyclin E. p27 is largely regulated in a post-transcriptional manner, while p21 is primarily controlled at the level of transcription (reviewed in [160]). In MCF-7 cells, both tamoxifen and fulvestrant increase the expression of p21 and p27 during cell cycle arrest [173]. p27 induction by tamoxifen induces quies-

cence in breast cancer cells and prevents a positive proliferative response following re-stimulation by growth factors such as IGF-I and EGF [174]. Combined treatment with tamoxifen and the farnesyl transferase inhibitor FTI-277, which causes a potent G1 arrest, maintains the expression and association of p21 and p27 with cyclin E/CDK2 complexes [23]. These two studies suggest that preservation of p27 and p21 is critical for the inhibitory effects of antiestrogens like tamoxifen in controlling breast cancer cell growth. In contrast, downregulation of these CDK inhibitors prevents the growth inhibitory effects of these drugs [173].

p21 and p27 functions are regulated in part by AKT through phosphorylation of residues within their nuclear localization sequences, leading to sequestration in the cytoplasm where they are unable to interact with cyclin E and CDK2 (discussed in [175]). c-Myc also plays a role in regulating p21, although the mechanism is less clear. Most significantly, c-Myc appears to block tamoxifen- and fulvestrant-induced expression of p21 [176]. In MCF-7/LCC9 breast cancer cells that have acquired resistance to fulvestrant and cross-resistance to tamoxifen, p21 expression is no longer regulated by estrogens and antiestrogens [176]. Studies of other models of antiestrogen resistance also support the idea that p21 and/or p27 play important roles in antiestrogen sensitivity and resistance. Unlike parental MCF-7 cells in which tamoxifen or fulvestrant treatment increases p27 binding to cyclin E, antiestrogens do not alter p27 binding in the tamoxifen-resistant LY-2 variant [177]. These LY-2 cells display constitutive activation of MEK and ERK1/2, and inhibition of these kinases partially restores antiestrogen sensitivity and p27 function. Other cell culture models of resistance, such as MCF-7/LCC2 and MCF-7/HER2-18 cells, exhibit increased MEK/ERK activity, although whether this affects antiestrogen resistance through modulating p27 remains to be determined [177]. These data supporting a role for p21 and p27 in antiestrogen responses and resistance are corroborated by clinical evidence (see Section 4.1).

3. Endocrine therapies; effect on motility/invasion/metastasis

Metastatic breast cancer is ultimately the greatest cause of disease mortality. Tamoxifen is beneficial for decreasing secondary disease occurrence, both at local and distant sites [178]. However, the major

phenotype of endocrine resistance is cancer recurrence. While clinical data have not definitively linked antiestrogen resistance with metastasis, many of the proteins/pathways discussed above in Section 2 that are involved in circumventing antiestrogen-induced blocks in cell cycle progression and survival are also implicated in increased migration and metastasis of breast cancer cells.

Hiscox et al. have begun to tackle the molecular relationship between antiestrogen resistance and invasion/metastasis [179]. They found that MCF-7 cells selected for tamoxifen resistance showed enhanced basal motility and an invasive phenotype. Treatment of these cells with the pharmacological inhibitors gefitinib and AZD0530 inhibited migration and invasion, indicating that the invasive phenotype of these resistant cells was dependent on EGFR and c-Src, respectively. The ability of tumor cells to metastasize requires multiple genotypic and phenotypic changes. Signaling perturbations associated with the epithelial to mesenchymal transition (EMT) contribute significantly to invasive and metastatic disease [180]. These include increased expression of extracellular matrix proteins, down-regulation of E-cadherin, disruption of adherens junctions complexes, accumulation of β -catenin and NF- κ B in the nucleus, and transcription of target genes such as c-Myc and cyclin D1 [181,182]. Interestingly, the tamoxifen-resistant MCF-7 cells investigated by Hiscox et al. displayed an altered morphology reminiscent of an EMT, marked by an elongated, fibroblast-like phenotype that lacked characteristic cell-cell contacts [179]. Tyrosine phosphorylation of β -catenin was elevated in the tamoxifen-resistant cells, coincident with accumulation of the protein, loss of its association with E-cadherin, increased interactions with EGFR, and increased transcription of c-Myc and cyclin D1. These effects were reversed in the presence of gefitinib, supporting a mechanism whereby the EGFR drives phosphorylation of β -catenin and activation of c-Src to initiate EMT-mediated metastatic progression.

Many of the other molecules that have been mentioned above in the context of promoting antiestrogen resistance also play a role in promoting invasiveness. For example, Pak1 is functionally linked to regulation of the actin cytoskeleton through Rho family GTPases [150]. The *Pak1* gene is closely linked to *CCND1* (cyclin D1) at 11q13, a region that is often amplified in breast cancers [183]. Similarly, amplification of the gene encoding c-Myc in breast tumors appears to correlate with

the transition from *in situ* to invasive carcinoma [38,184]. Finally, in a study of matched primary and metastatic breast tumors, Bcl-2 expression was found to be elevated in metastatic tumors coincident with a retention of ER α expression [38]. These results conflicted with those from a similar study performed on paired primary and recurrent tamoxifen-resistant tumors, which found that the recurrent tumors did not show a correlation between ER status and Bcl-2 expression [30]. However, there was a correlation between ER α and phosphorylated p38 and ERK in these relapsed specimens. These discrepancies may be due to differences in inclusion criteria and the source of recurrent tissue between studies. The former study evaluated mostly tumors at distant sites after discontinuation of hormonal therapy, while the latter study evaluated locoregional recurrences while the patients continued therapy.

4. Antiestrogen resistance: implications for treatment

4.1. Prediction of response

It is clear from the discussion above that resistance to endocrine therapy likely occurs through diverse mechanisms that vary from patient to patient. Consequently, it is imperative to develop molecular signatures that can predict the likelihood of response and potential for relapse of individual tumors. Initially, predictive markers for tamoxifen resistance included expression of ER and receptor tyrosine kinases such as EGFR and HER2. As our understanding of antiestrogen resistance has evolved, these markers have broadened. Currently, the overall expression of proteins, protein modifications, and intracellular localization are all being analyzed for predictive value of response to endocrine therapy.

As mentioned above, survival pathways regulated by PI3K, AKT, PTEN, and Bcl-2 family members play an important role in antiestrogen resistance. Studies are currently underway to determine whether increased activation and/or expression of AKT, decreased activity of PTEN, or altered expression of Bad could be predictive markers for response to anti-hormonal treatments. For example, a retrospective analysis of 402 ER α -positive breast carcinoma biopsies treated with tamoxifen for a median of 5 years revealed that high expression of AKT2, as measured by immunohistochemistry, was predictive of improved overall survival [31]. Patients who relapsed following

tamoxifen treatment were more likely to have low AKT2 expression and high cytoplasmic phospho-AKT (Ser-473). Interestingly, expression of AKT1 and AKT3 did not have a significant predictive correlation. Bad expression could also predict tamoxifen-treatment outcome. Patients in this same clinical study whose tumors had high levels of Bad expression had improved disease-free survival rates compared to patients whose tumors had low levels of Bad [36]. There was no association between phospho-Bad (Ser-112), Bax, Bcl-2 or Bcl-xL expression and response to tamoxifen treatment in this group of patients. A related study examined 252 primary ER α human breast carcinomas to determine whether activated AKT or loss of heterozygosity (LOH) of PTEN was predictive of response to tamoxifen [185]. Phospho-AKT correlated with HER2 status in this study, as did PTEN LOH. From this patient cohort, 36 patient samples were used to examine the relationship between activated AKT and the efficacy of endocrine therapy for metastatic breast cancer. Patients with phospho-AKT-positive tumors received less benefit from endocrine therapy than did patients with phospho-AKT-negative tumors. A third study evaluated PTEN expression in 100 patients with ER α breast carcinomas who had been treated with tamoxifen [186]. High levels of PTEN were recorded for 81% of patients who had non-recurrent disease and only 41% of patients with recurrent disease. Conversely, low PTEN expression was associated with a shorter relapse free survival and reduced disease-specific survival. PTEN expression was an even stronger predictor of relapse and survival when the analysis was restrained to only stage I breast cancer patients.

Like the PI3K/AKT survival pathway, cyclin D1 expression is associated with multiple mechanisms of antiestrogen resistance. Patients with ER α -positive tumors and high levels of cyclin D1 mRNA had a worse prognosis for overall survival [187]. In a small subset of these patients who were treated with endocrine therapy, an apparent association between high cyclin D1 mRNA and decreased duration of response was made. Similar results were obtained by evaluating protein levels of cyclin D1 in 248 post-menopausal patients who had been randomized into 2-year tamoxifen treatment or no-treatment groups [188]. Interestingly, the 55 strongly ER α -positive patients with tumors showing moderate or low cyclin D1 levels responded to tamoxifen treatment, whereas the 46 patients with both high ER α and cyclin D1 expression in their

tumors showed no difference in survival between tamoxifen and no treatment.

The association between cyclin E levels and antiestrogen response is even less clear. Some of the clinical studies focused on cyclin E have shown a correlation between the presence of low molecular weight and/or full-length cyclin E and poor survival, while others have not found such an association ([189] and discussed in [161]). A recent study examining the expression of cyclin E and two proteases implicated in the generation of low molecular weight cyclin E (proteinase 3 and neutrophil elastase) in 205 breast cancer patients revealed that cyclin E was significantly associated with poor disease-free survival in the entire cohort, and with tamoxifen resistance in the 110 treated patients [190]. However, expression of neutrophil elastase was not associated with poor outcome or treatment resistance. Further studies are required to clarify the role of these variants in clinical endocrine resistance.

Coincident with cyclin D1 and cyclin E expression, p21 and p27 levels may also prove to be predictive for clinical response to antiestrogen resistance. High levels of p27 expression in a group of 512 patients with premenopausal breast cancer patients treated with tamoxifen and the gonadotropin-releasing hormone agonist goserelin were strongly associated with improved overall and disease-free survival [191]. Data for p21 are less consistent, perhaps because they may be confounded by the complex regulation of p21 expression and contributions from other proteins such as p53 and IRF-1 that may independently affect breast cancer outcome (discussed in [20,161]). It may also be that subcellular localization of p21, rather than overall expression levels, will prove to be a better marker of therapeutic response and treatment outcome. In line with this suggestion, Perez-Tenorio et al. have recently reported that cytoplasmic p21 is associated with increased AKT activity and poor tamoxifen response in a cohort of 280 women [192].

As mentioned above, serine phosphorylation of ER α may also prove to be useful for predicting therapeutic response (see Section 2.5). Recent studies have centered on the role of Ser-118 phosphorylation in predicting clinical outcome in breast cancer patients. In a series of 117 lymph node-negative primary breast tumors from patients treated with tamoxifen following surgery with or without radiation, Murphy et al. observed Ser-118 phosphorylation only in tumors with positive estrogen binding activity [193]. Phosphorylated Ser-118 was also

observed more frequently in PR-positive tumors than PR-negative tumors, and patients with tumors positive for Ser-118 phosphorylation showed increased disease-free survival. Bergqvist et al. reported that increased MAPK activation and expression of phosphorylated Ser-118 correlated with one another and also predicted better survival [194]. However, overexpression of HER2 was inversely correlated with activated ERK1/2 and phosphorylated Ser-118, suggesting that at least in this cohort, HER2 activation does not drive phosphorylation and hyperactivation of ER. Moreover, this apparent improved outcome observed in patients with Ser-118 phosphorylation appears to contradict some of the cell culture experiments that showed a link between Ser-118 phosphorylation and antiestrogen resistance (see Section 2.5). This discrepancy may be explained by the possibility that Ser-118 phosphorylation of the ER may have distinct functions in different phases of breast cancer; there is some evidence to suggest that it may be associated with a better outcome prior to endocrine therapy but a worse outcome following acquisition of tamoxifen resistance [35].

The correlation between Ser-167 phosphorylation and clinical response to antiestrogens is equally complex. Kirkegaard et al. found that high levels of active AKT correlated with reduced survival and increased Ser-167 phosphorylation in tamoxifen-treated patients [31]. However, Yamashita et al. reported that phosphorylated Ser-167, along with expression of ER and PR, were associated with an improved response to endocrine therapy and survival [195]. The fact that Ser-167 can be phosphorylated by kinases other than AKT may suggest that it is AKT rather than phosphorylated Ser-167 that is responsible for the poor outcome in the Kirkegaard study [31].

In addition to the ER α modulation, PR status has also been suggested to be predictive of response to antiestrogens. As discussed above (Section 2.1), the presence of PR is reflective of a functional ER pathway because PR synthesis is initiated by ER-induced transcription. Bardou et al. analyzed two databases containing information on ER and PR status of a large number of breast cancer patients [196]. They found that PR expression was associated with disease-free survival independently of ER α status. However, ER α -positive/PR-positive tumors received the greatest benefit from adjuvant hormonal therapy when both ER α and PR were taken into consideration, while loss of expression of either

receptor reduced responsiveness by nearly half. Additional studies have also found that ER/PR-positive breast tumors have significant improvement in prognosis after either adjuvant tamoxifen or letrozole treatments [197,198]. This supports the hypothesis that ER α positive tumors that lack PR expression are less likely to respond to anti-hormonal therapy because they are no longer dependent on estrogen for survival. Conversely, ER β expression appears to be predictive of a positive response to endocrine therapy and better overall survival [6]. This is potentially due to the putative tumor suppressor functions of ER β discussed above (see Section 1), and provides the rationale for developing SERMs that exhibit selective activities to ER subtypes (see Section 1.1.1).

4.2. Molecular profiling studies of antiestrogen resistance

Gene expression analysis and molecular profiling have added to our understanding of the mechanisms of antiestrogen response and resistance. Several groups have used these approaches on tumor samples to develop gene signatures that can predict clinical responses to antiestrogen treatment [199,200]. However, in some instances, these profiles have not been validated by independent studies [201]. Expression profiling of cell culture and xenograft models has also been extensively utilized to help clarify the molecular actions of antiestrogens and aromatase inhibitors. Several of these studies have confirmed biological evidence indicating that SERMs like tamoxifen and raloxifene can behave as both antagonists and agonists of the ER, whereas fulvestrant is a pure antagonist. For example, expression of cell cycle-associated genes such as cyclin A1 and CDC25B was unaffected by estrogen, tamoxifen, or raloxifene but strongly regulated by fulvestrant [202]. The effects of tamoxifen and raloxifene were also similar to estrogen with respect to expression of Bcl-2-interacting killer (BIK), an estrogen-inhibited, pro-apoptotic gene that has been demonstrated to play a critical role in fulvestrant-induced breast cancer cell death [203].

Other studies have examined regulation of gene expression by letrozole and anastrozole as compared to tamoxifen [204]. As might be expected, the effects of these aromatase inhibitors were more similar to each other than they were to tamoxifen. However, the major functional classes of genes regulated by all three drugs were comparable, and

included apoptosis, Wnt, and MAPK signaling pathways. This suggests that, rather than looking at individual genes, these approaches may be most useful for identifying functional groupings of genes that work together in specific signaling pathways to regulate responses to antiestrogens. A recent study in MCF-7 cells comparing acquired resistance to tamoxifen and fulvestrant found that tamoxifen resistance was strongly associated with changes in the PKA pathway, MAPK phosphatases, and calcium binding proteins. In contrast, downregulation of ER and major alterations in growth factor receptor and interferon signaling were observed in cells resistant to fulvestrant [60]. Interferon-inducible genes have also been found to be differentially regulated in the MaCa 3366/TAM xenograft model of acquired tamoxifen resistance when compared to the tamoxifen-sensitive MaCa 3366 counterpart [205]. Interestingly, loss of the putative tumor suppressor interferon regulatory factor 1 (IRF-1) has been specifically implicated in fulvestrant resistance [25] and is required for tamoxifen-induced cell death of normal mammary epithelial cells [206]. Another interferon-induced gene, calgranulin A (S100A8) [207], is upregulated in MCF-7, T47D, and ZR-75-1-derived models of fulvestrant resistance [59], providing additional support for a role of interferon-inducible genes in antiestrogen resistance.

While expression profiling has provided insight into molecular pathways that may contribute to antiestrogen resistance, there are some challenges associated with these approaches that must be carefully considered. Multiple expression array platforms and technologies can make comparing the findings of different investigators difficult, especially at the level of individual genes. Statistical analysis of these data is also very complex, and the risk of excluding biologically relevant genes while including others that do not contribute to the phenotype is a significant one. Unfortunately, these challenges are greatly magnified in the context of analysis of tumor tissues [208–210]. Nonetheless, these approaches can be quite powerful, particularly when one considers functional grouping of genes rather than individual molecules.

4.3. Combinatorial therapies

Strategies for treating breast tumors have significantly broadened in recent years because of a growing appreciation for the fact that resistance to endocrine therapies can and will invariably arise

through a variety of distinct mechanisms. Our increasing understanding of the molecular basis for these failures has brought to the forefront the possibility of using combination therapies for treating breast cancer. There are two separate rationales for using combination therapies. First, targeting multiple pathways at once can potentially delay or halt the acquisition of a resistant phenotype. Alternatively, treatments can be administered to effectively resensitize tumors to endocrine therapies for which they are resistant. In many cases, the practical strategies for addressing these distinct phenomena are the same. For example, as discussed above, growth factor receptor signaling is often increased in antiestrogen-resistant breast tumors, leading to upregulation of proliferation and survival pathways that may be distinct from estrogen signaling per se. Thus, treatment with gefitinib or trastuzumab to inhibit the EGFR or HER2, respectively, along with hormonal therapies would potentially block both the pro-proliferative activity of the ER and the proliferation/survival pathways activated by the growth factor receptors. Some of the current trials evaluating the efficacy of these combinations include a Phase II randomized trial (Translational Oncology Research International trial) that is investigating fulvestrant and trastuzumab in combination as a first-line treatment for patients with ER α /HER2-positive breast tumors, a comparative study evaluating anastrozole versus fulvestrant with the addition of gefitinib in women with ER α -positive tumors who have metastatic disease and have failed chemotherapy and/or endocrine therapy, and a combination of fulvestrant with the dual inhibitor lapatinib in postmenopausal women with ER α -positive and EGFR-and/or HER2-positive tumors who have had previous exposure to an aromatase inhibitor (reviewed in [211,212]).

In addition to targeting growth factor receptors, several clinical trials have been initiated to investigate the effect of farnesyl transferase inhibitors such as tipifarnib in combination with endocrine therapies. However, these trials have been somewhat disappointing in that they have not thus far corroborated pre-clinical models showing that tipifarnib in combination with tamoxifen resulted in greater tumor regression than either treatment alone [213]. Additional studies are underway that examine the efficacy of combining tipifarnib with AIs (reviewed in [214]).

Clinical studies with mTOR antagonists in combination with hormonal therapies are also ongoing.

As a single agent, CCI-779 has been somewhat effective in Phase II studies of metastatic breast cancer [215]. Currently, Phase II and Phase III trials are testing either CCI-779 or RAD-001 in combination with letrozole, as compared to letrozole alone. Interim results show that the combination of CCI-779 and letrozole translates into a clinical benefit rate of 77% versus 66% for letrozole alone, although these values have not yet reached statistical significance [135].

Finally, extensive efforts are being made to determine whether pre-clinical data showing that tumors resistant to one class of hormonal therapy may still retain sensitivity to alternate endocrine therapies hold true in the clinic. Results of the Phase II Swiss Group for Clinical Research Trial (SAKK 21/00), which compared the efficacy of fulvestrant treatment in AI-sensitive versus AI-resistant patients, were recently published (reviewed in [216]). Overall fulvestrant yielded a clinical benefit (measured as an objective response or stable disease >24 weeks) in 30% of patients, irrespective of whether they had previously been treated with AIs. There are currently multiple planned or ongoing clinical trials combining AIs with antiestrogens. These include Phase II and Phase III trials evaluating use as first line and neoadjuvant treatment (reviewed in [214]).

Acknowledgments

The authors acknowledge support of our research from the Susan G. Komen Breast Cancer Foundation (PDF0503551) and Department of Defense Breast Cancer Research Program (BC051851) to RBR, the Department of Defense Breast Cancer Research Program (BC050339) to RSS, the National Cancer Institute institutional training Grant (T32 CA009109) for R.S.S. and M.S.G.; and the National Institutes of Health (R01 CA 096846 to AHB and R01 CA 123037 to Sarah J. Parsons, AHB Co-PI).

References

- [1] P. Ascenzi, A. Bocedi, M. Marino, Structure–function relationship of estrogen receptor alpha and beta: impact on human health, *Mol. Aspects Med.* 27 (2006) 299–402.
- [2] E. Anderson, R.B. Clarke, Steroid receptors and cell cycle in normal mammary epithelium, *J. Mammary Gland Biol. Neoplasia* 9 (2004) 3–13.
- [3] V. Speirs, G.P. Skliris, S.E. Burdall, P.J. Carder, Distinct expression patterns of ER alpha and ER beta in normal human mammary gland, *J. Clin. Pathol.* 55 (2002) 371–374.
- [4] C. Forster, S. Makela, A. Warri, S. Kietz, D. Becker, K. Hultenby, et al., Involvement of estrogen receptor beta in terminal differentiation of mammary gland epithelium, *Proc. Natl. Acad. Sci. USA* 99 (2002) 15578–15583.
- [5] S. Saji, M. Hirose, M. Toi, Clinical significance of estrogen receptor beta in breast cancer, *Cancer Chemother. Pharmacol.* 56 (Suppl. 1) (2005) 21–26.
- [6] L.C. Murphy, P.H. Watson, Is oestrogen receptor-beta a predictor of endocrine therapy responsiveness in human breast cancer?, *Endocr. Relat. Cancer* 13 (2006) 327–334.
- [7] J. Matthews, B. Wihlen, M. Tujague, J. Wan, A. Strom, J.A. Gustafsson, Estrogen receptor (ER) beta modulates ER alpha-mediated transcriptional activation by altering the recruitment of c-fos and c-jun to estrogen-responsive promoters, *Mol. Endocrinol.* 20 (2006) 534–543.
- [8] L.A. Helguero, M.H. Faulds, J.A. Gustafsson, L.A. Haldosen, Estrogen receptors alpha (ER alpha) and beta (ER beta) differentially regulate proliferation and apoptosis of the normal murine mammary epithelial cell line HC11, *Oncogene* 24 (2005) 6605–6616.
- [9] F. Stossi, D.H. Barnett, J. Frasar, B. Komm, C.R. Lyttle, B.S. Katzenellenbogen, Transcriptional profiling of estrogen-regulated gene expression via estrogen receptor (ER) alpha or ER beta in human osteosarcoma cells: distinct and common target genes for these receptors, *Endocrinology* 145 (2004) 3473–3486.
- [10] E.C. Chang, J. Frasar, B. Komm, B.S. Katzenellenbogen, Impact of estrogen receptor beta on gene networks regulated by estrogen receptor alpha in breast cancer cells, *Endocrinology* 147 (2006) 4831–4842.
- [11] J.S. Lewis, V.C. Jordan, Selective estrogen receptor modulators (SERMs): mechanisms of anticarcinogenesis and drug resistance, *Mutat. Res.* 591 (2005) 247–263.
- [12] B. Fisher, J.P. Costantino, D.L. Wickerham, C.K. Redmond, M. Kavanah, W.M. Cronin, et al., Tamoxifen for prevention of breast cancer: Report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study, *J. Natl. Cancer Inst.* 90 (1998) 1371–1388.
- [13] G.H. Veeneman, Non-steroidal subtype selective estrogens, *Curr. Med. Chem.* 12 (2005) 1077–1136.
- [14] A. Escande, A. Pillon, N. Servant, J.P. Cravedi, F. Larrea, P. Muhn, et al., Evaluation of ligand selectivity using reporter cell lines stably expressing estrogen receptor alpha or beta, *Biochem. Pharmacol.* 71 (2006) 1459–1469.
- [15] T.A. Blizzard, C. Gude, J.D. Morgan 2nd, W. Chan, E.T. Birzin, M. Mojena, et al., Androstenediol analogs as ER-beta-selective SERMs, *Bioorg. Med. Chem. Lett.* 16 (2006) 834–838.
- [16] A.U. Buzdar, TAS-108: a novel steroidal antiestrogen, *Clin. Cancer Res.* 11 (2005) 906S–908S.
- [17] A. Howell, Pure oestrogen antagonists for the treatment of advanced breast cancer, *Endocr. Relat. Cancer* 13 (2006) 689–706.
- [18] C.K. Osborne, A. Wakeling, R.I. Nicholson, Fulvestrant: an oestrogen receptor antagonist with a novel mechanism of action, *Br. J. Cancer* 90 (Suppl. 1) (2004) S2–S6.
- [19] R.W. Brueggemeier, Update on the use of aromatase inhibitors in breast cancer, *Expert Opin. Pharmacother.* 7 (2006) 1919–1930.
- [20] R.B. Riggins, A.H. Bouton, M.C. Liu, R. Clarke, Antiestrogens, aromatase inhibitors, and apoptosis in breast cancer, *Vitam. Horm.* 71 (2005) 201–237.

- [21] S. Chen, S. Masri, X. Wang, S. Phung, Y.C. Yuan, X. Wu, What do we know about the mechanisms of aromatase inhibitor resistance?, *J. Steroid Biochem. Mol. Biol.* 102 (2006) 232–240.
- [22] J.S. Carroll, O.W. Prall, E.A. Musgrove, R.L. Sutherland, A pure estrogen antagonist inhibits cyclin E-CDK2 activity in MCF-7 breast cancer cells and induces accumulation of p130-E2F4 complexes characteristic of quiescence, *J. Biol. Chem.* 275 (2000) 38221–38229.
- [23] S.F. Doisneau-Sixou, P. Cestac, J.C. Faye, G. Favre, R.L. Sutherland, Additive effects of tamoxifen and the farnesyl transferase inhibitor FTI-277 on inhibition of MCF-7 breast cancer cell-cycle progression, *Int. J. Cancer* 106 (2003) 789–798.
- [24] A. Thiantanawat, B.J. Long, A.M. Brodie, Signaling pathways of apoptosis activated by aromatase inhibitors and antiestrogens, *Cancer Res.* 63 (2003) 8037–8050.
- [25] K.B. Bouker, T.C. Skaar, D.R. Fernandez, K.A. O'Brien, R.B. Riggins, D. Cao, et al., Interferon regulatory factor-1 mediates the proapoptotic but not cell cycle arrest effects of the steroidal antiestrogen ICI 182,780 (faslodex, fulvestrant), *Cancer Res.* 64 (2004) 4030–4039.
- [26] S.F. Doisneau-Sixou, C.M. Sergio, J.S. Carroll, R. Hui, E.A. Musgrove, R.L. Sutherland, Estrogen and antiestrogen regulation of cell cycle progression in breast cancer cells, *Endocr. Relat. Cancer* 10 (2003) 179–186.
- [27] G. Arpino, S.J. Green, D.C. Allred, D. Lew, S. Martino, C.K. Osborne, et al., HER-2 amplification, HER-1 expression, and tamoxifen response in estrogen receptor-positive metastatic breast cancer: a Southwest Oncology Group Study, *Clin. Cancer Res.* 10 (2004) 5670–5676.
- [28] J.M. Gee, J.F. Robertson, E. Gutteridge, I.O. Ellis, S.E. Pinder, M. Rubini, et al., Epidermal growth factor receptor/HER2/insulin-like growth factor receptor signaling and oestrogen receptor activity in clinical breast cancer, *Endocr. Relat. Cancer* 12 (Suppl. 1) (2005) S99–S111.
- [29] M. Dowsett, S. Johnston, L.A. Martin, J. Salter, M. Hills, S. Detre, et al., Growth factor signalling and response to endocrine therapy: The Royal Marsden Experience, *Endocr. Relat. Cancer* 12 (Suppl. 1) (2005) S113–S117.
- [30] M.C. Gutierrez, S. Detre, S. Johnston, S.K. Mohsin, J. Shou, D.C. Allred, et al., Molecular changes in tamoxifen-resistant breast cancer: relationship between estrogen receptor, HER-2, and p38 mitogen-activated protein kinase, *J. Clin. Oncol.* 23 (2005) 2469–2476.
- [31] T. Kirkegaard, C.J. Witton, L.M. McGlynn, S.M. Tovey, B. Dunne, A. Lyon, et al., AKT activation predicts outcome in breast cancer patients treated with tamoxifen, *J. Pathol.* 207 (2005) 139–146.
- [32] C. Holm, S. Rayala, K. Jirstrom, O. Stal, R. Kumar, G. Landberg, Association between Pak1 expression and subcellular localization and tamoxifen resistance in breast cancer patients, *J. Natl. Cancer Inst.* 98 (2006) 671–680.
- [33] S. van der Flier, A. Brinkman, M.P. Look, E.M. Kok, M.E. Meijer-van Gelder, J.G. Klijn, et al., BCAR1/p130^{Cas} protein and primary breast cancer: prognosis and response to tamoxifen treatment, *J. Natl. Cancer Inst.* 92 (2000) 120–127.
- [34] C.K. Osborne, V. Bardou, T.A. Hopp, G.C. Chamness, S.G. Hilsenbeck, S.A. Fuqua, et al., Role of the estrogen receptor coactivator AIB1 (SRC-3) and HER-2/neu in tamoxifen resistance in breast cancer, *J. Natl. Cancer Inst.* 95 (2003) 353–361.
- [35] N. Sarwar, J.S. Kim, J. Jiang, D. Peston, H.D. Sinnett, P. Madden, et al., Phosphorylation of ER alpha at serine 118 in primary breast cancer and in tamoxifen-resistant tumours is indicative of a complex role for ER alpha phosphorylation in breast cancer progression, *Endocr. Relat. Cancer* 13 (2006) 851–861.
- [36] E. Cannings, T. Kirkegaard, S.M. Tovey, B. Dunne, T.G. Cooke, J.M. Bartlett, Bad expression predicts outcome in patients treated with tamoxifen, *Breast Cancer Res. Treat.* 102 (2007) 173–179.
- [37] S.P. Linke, T.M. Bremer, C.D. Herold, G. Sauter, C. Diamond, A multimarker model to predict outcome in tamoxifen-treated breast cancer patients, *Clin. Cancer Res.* 12 (2006) 1175–1183.
- [38] M.D. Planas-Silva, R.D. Bruggeman, R.T. Grenko, J.S. Smith, Overexpression of c-Myc and Bcl-2 during progression and distant metastasis of hormone-treated breast cancer, *Exp. Mol. Pathol.* 82 (2007) 85–90.
- [39] E. Rahko, G. Blanco, R. Bloigu, Y. Soini, A. Talvensaari-Mattila, A. Jukkola, Adverse outcome and resistance to adjuvant antiestrogen therapy in node-positive postmenopausal breast cancer patients—the role of p53, *Breast* 15 (2006) 69–75.
- [40] G.G. Kuiper, E. Enmark, M. Peltö-Huikko, S. Nilsson, J.A. Gustafsson, Cloning of a novel receptor expressed in rat prostate and ovary, *Proc. Natl. Acad. Sci. USA* 93 (1996) 5925–5930.
- [41] D.P. McDonnell, The molecular determinants of estrogen receptor pharmacology, *Maturitas* 48 (Suppl. 1) (2004) S7–S12.
- [42] K. Paech, P. Webb, G.G. Kuiper, S. Nilsson, J. Gustafsson, P.J. Kushner, et al., Differential ligand activation of estrogen receptors ER alpha and ER beta at AP1 sites, *Science* 277 (1997) 1508–1510.
- [43] A. Zou, K.B. Marschke, K.E. Arnold, E.M. Berger, P. Fitzgerald, D.E. Mais, et al., Estrogen receptor beta activates the human retinoic acid receptor alpha-1 promoter in response to tamoxifen and other estrogen receptor antagonists, but not in response to estrogen, *Mol. Endocrinol.* 13 (1999) 418–430.
- [44] C.L. Smith, B.W. O'Malley, Coregulator function: a key to understanding tissue specificity of selective receptor modulators, *Endocr. Rev.* 25 (2004) 45–71.
- [45] J.M. Hall, D.P. McDonnell, Coregulators in nuclear estrogen receptor action: from concept to therapeutic targeting, *Mol. Interv.* 5 (2005) 343–357.
- [46] R. Kumar, A.E. Gururaj, R.K. Vadlamudi, S.K. Rayala, The clinical relevance of steroid hormone receptor corepressors, *Clin. Cancer Res.* 11 (2005) 2822–2831.
- [47] I. Girault, I. Bieche, R. Lidereau, Role of estrogen receptor alpha transcriptional coregulators in tamoxifen resistance in breast cancer, *Maturitas* 54 (2006) 342–351.
- [48] R. Clarke, F. Leonessa, J.N. Welch, T.C. Skaar, Cellular and molecular pharmacology of antiestrogen action and resistance, *Pharmacol. Rev.* 53 (2001) 25–71.
- [49] S.F. Honig, Treatment of metastatic disease: hormonal therapy and chemotherapy, in: J.R. Harris, M.E. Lippman, M. Morrow, S. Helman (Eds.), *Diseases of the Breast*, Lippincott-Raven, Philadelphia, 1996, pp. 669–734.

- [50] Y.L. Ottaviano, J.P. Issa, F.F. Parl, H.S. Smith, S.B. Baylin, N.E. Davidson, Methylation of the estrogen receptor gene CpG island marks loss of estrogen receptor expression in human breast cancer cells, *Cancer Res.* 54 (1994) 2552–2555.
- [51] N. Roodi, L.R. Bailey, W.Y. Kao, C.S. Verrier, C.J. Yee, W.D. Dupont, et al., Estrogen receptor gene analysis in estrogen receptor-positive and receptor-negative primary breast cancer, *J. Natl. Cancer Inst.* 87 (1995) 446–451.
- [52] D. Sharma, N.K. Saxena, N.E. Davidson, P.M. Vertino, Restoration of tamoxifen sensitivity in estrogen receptor-negative breast cancer cells: tamoxifen-bound reactivated ER recruits distinctive corepressor complexes, *Cancer Res.* 66 (2006) 6370–6378.
- [53] L. Giacinti, P.P. Claudio, M. Lopez, A. Giordano, Epigenetic information and estrogen receptor alpha expression in breast cancer, *Oncologist* 11 (2006) 1–8.
- [54] M. Garcia, D. Derocq, G. Freiss, H. Rochefort, Activation of estrogen receptor transfected into a receptor-negative breast cancer cell line decreases the metastatic and invasive potential of the cells, *Proc. Natl. Acad. Sci. USA* 89 (1992) 11538–11542.
- [55] S.R. Johnston, G. Saccani-Jotti, I.E. Smith, J. Salter, J. Newby, M. Coppen, et al., Changes in estrogen receptor, progesterone receptor, and PS2 expression in tamoxifen-resistant human breast cancer, *Cancer Res.* 55 (1995) 3331–3338.
- [56] C.A. Encarnacion, D.R. Ciocca, W.L. McGuire, G.M. Clark, S.A. Fuqua, C.K. Osborne, Measurement of steroid hormone receptors in breast cancer patients on tamoxifen, *Breast Cancer Res. Treat.* 26 (1993) 237–246.
- [57] R. Clarke, T.C. Skaar, K.B. Bouker, N. Davis, Y.R. Lee, J.N. Welch, et al., Molecular and pharmacological aspects of antiestrogen resistance, *J. Steroid Biochem. Mol. Biol.* 76 (2001) 71–84.
- [58] H.W. van den Berg, M. Lynch, J. Martin, J. Nelson, G.R. Dickson, A.D. Crockard, Characterisation of a tamoxifen-resistant variant of the ZR-75-1 human breast cancer cell line (ZR-75-9a1) and ability of the resistant phenotype, *Br. J. Cancer* 59 (1989) 522–526.
- [59] A. Sommer, J. Hoffmann, R.B. Lichtner, M.R. Schneider, K. Parczyk, Studies on the development of resistance to the pure antiestrogen faslodex in three human breast cancer cell lines, *J. Steroid Biochem. Mol. Biol.* 85 (2003) 33–47.
- [60] M. Fan, P.S. Yan, C. Hartman-Frey, L. Chen, H. Paik, S.L. Oyer, et al., Diverse gene expression and DNA methylation profiles correlate with differential adaptation of breast cancer cells to the antiestrogens tamoxifen and fulvestrant, *Cancer Res.* 66 (2006) 11954–11966.
- [61] S.S. Larsen, M.W. Madsen, B.L. Jensen, A.E. Lykkesfeldt, Resistance of human breast-cancer cells to the pure steroidal anti-estrogen ICI 182,780 is not associated with a general loss of estrogen-receptor expression or lack of estrogen responsiveness, *Int. J. Cancer* 72 (1997) 1129–1136.
- [62] B.L. Jensen, J. Skouv, B.K. Lundholt, A.E. Lykkesfeldt, Differential regulation of specific genes in MCF-7 and the ICI 182780-resistant cell line MCF-7/182R-6, *Br. J. Cancer* 79 (1999) 386–392.
- [63] N. Brunner, B. Boysen, S. Jirus, T.C. Skaar, C. Holst-Hansen, J. Lippman, et al., MCF7/LCC9: an antiestrogen-resistant MCF-7 variant in which acquired resistance to the steroidal antiestrogen ICI 182,780 confers an early cross-resistance to the nonsteroidal antiestrogen tamoxifen, *Cancer Res.* 57 (1997) 3486–3493.
- [64] N. Brunner, V. Boulay, A. Fojo, C.E. Freter, M.E. Lippman, R. Clarke, Acquisition of hormone-independent growth in MCF-7 cells is accompanied by increased expression of estrogen-regulated genes but without detectable DNA amplifications, *Cancer Res.* 53 (1993) 283–290.
- [65] B. Kuske, C. Naughton, K. Moore, K.G. Macleod, W.R. Miller, R. Clarke, et al., Endocrine therapy resistance can be associated with high estrogen receptor α (ER α) expression and reduced ER α phosphorylation in breast cancer models, *Endocr. Relat. Cancer* 13 (2006) 1121–1133.
- [66] R.B. Riggins, A. Zwart, R. Nehra, R. Clarke, The nuclear factor kappa B inhibitor parthenolide restores ICI 182,780 (faslodex; fulvestrant)-induced apoptosis in antiestrogen-resistant breast cancer cells, *Mol. Cancer Ther.* 4 (2005) 33–41.
- [67] A. Howell, Fulvestrant ('faslodex'): current and future role in breast cancer management, *Crit. Rev. Oncol. Hematol.* 57 (2006) 265–273.
- [68] M.L. Graham 2nd., N.L. Krett, L.A. Miller, K.K. Leslie, D.F. Gordon, W.M. Wood, et al., T47DCO cells, genetically unstable and containing estrogen receptor mutations, are a model for the progression of breast cancers to hormone resistance, *Cancer Res.* 50 (1990) 6208–6217.
- [69] M.L. Graham 2nd., J.A. Smith, P.B. Jewett, K.B. Horwitz, Heterogeneity of progesterone receptor content and remodeling by tamoxifen characterize subpopulations of cultured human breast cancer cells: analysis by quantitative dual parameter flow cytometry, *Cancer Res.* 52 (1992) 593–602.
- [70] M.H. Herynk, S.A. Fuqua, Estrogen receptor mutations in human disease, *Endocr. Rev.* 25 (2004) 869–898.
- [71] J.J. Pink, S.Q. Wu, D.M. Wolf, M.M. Bilimoria, V.C. Jordan, A novel 80 kDa human estrogen receptor containing a duplication of exons 6 and 7, *Nucleic Acids Res.* 24 (1996) 962–969.
- [72] P.S. Karnik, S. Kulkarni, X.P. Liu, G.T. Budd, R.M. Bukowski, Estrogen receptor mutations in tamoxifen-resistant breast cancer, *Cancer Res.* 54 (1994) 349–353.
- [73] Q.X. Zhang, A. Borg, D.M. Wolf, S. Oesterreich, S.A. Fuqua, An estrogen receptor mutant with strong hormone-independent activity from a metastatic breast cancer, *Cancer Res.* 57 (1997) 1244–1249.
- [74] S.A. Fuqua, C. Wiltchke, Q.X. Zhang, A. Borg, C.G. Castles, W.E. Friedrichs, et al., A hypersensitive estrogen receptor-alpha mutation in premalignant breast lesions, *Cancer Res.* 60 (2000) 4026–4029.
- [75] Z. Zhang, H. Yamashita, T. Toyama, Y. Omoto, H. Sugiura, Y. Hara, et al., Estrogen receptor alpha mutation (A-to-G transition at nucleotide 908) is not found in different types of breast lesions from Japanese women, *Breast Cancer* 10 (2003) 70–73.
- [76] P.M. Navolanic, L.S. Steelman, J.A. McCubrey, EGFR family signaling and its association with breast cancer development and resistance to chemotherapy, *Int. J. Oncol.* 22 (2003) 237–252.
- [77] R.I. Nicholson, I.R. Hutcheson, J.M. Knowlden, H.E. Jones, M.E. Harper, N. Jordan, et al., Nonendocrine pathways and endocrine resistance: observations with antiestrogens and signal transduction inhibitors in combination, *Clin. Cancer Res.* 10 (2004) 346S–354S.

- [78] R.I. Nicholson, I.R. Hutcheson, D. Britton, J.M. Knowlden, H.E. Jones, M.E. Harper, et al., Growth factor signalling networks in breast cancer and resistance to endocrine agents: new therapeutic strategies, *J. Steroid Biochem. Mol. Biol.* 93 (2005) 257–262.
- [79] H. Kurokawa, A.E. Lenferink, J.F. Simpson, P.I. Pisacane, M.X. Sliwkowski, J.T. Forbes, et al., Inhibition of HER2/neu (ErbB-2) and mitogen-activated protein kinases enhances tamoxifen action against HER2-overexpressing, tamoxifen-resistant breast cancer cells, *Cancer Res.* 60 (2000) 5887–5894.
- [80] H. Kurokawa, C.L. Arteaga, Inhibition of ErbB receptor (HER) tyrosine kinases as a strategy to abrogate antiestrogen resistance in human breast cancer, *Clin. Cancer Res.* 7 (2001) 4436S–4442S, discussion 4411S–4412S.
- [81] H. Kurokawa, C.L. Arteaga, ErbB (HER) receptors can abrogate antiestrogen action in human breast cancer by multiple signaling mechanisms, *Clin. Cancer Res.* 9 (2003) 511S–515S.
- [82] R.A. McClelland, D. Barrow, T.A. Madden, C.M. Dutkowski, J. Pamment, J.M. Knowlden, et al., Enhanced epidermal growth factor receptor signaling in MCF7 breast cancer cells after long-term culture in the presence of the pure antiestrogen ICI 182,780 (faslodex), *Endocrinology* 142 (2001) 2776–2788.
- [83] J.M. Knowlden, I.R. Hutcheson, H.E. Jones, T. Madden, J.M. Gee, M.E. Harper, et al., Elevated levels of epidermal growth factor receptor/c-ErbB2 heterodimers mediate an autocrine growth regulatory pathway in tamoxifen-resistant MCF-7 cells, *Endocrinology* 144 (2003) 1032–1044.
- [84] I.R. Hutcheson, J.M. Knowlden, T.A. Madden, D. Barrow, J.M. Gee, A.E. Wakeling, et al., Oestrogen receptor-mediated modulation of the EGFR/MAPK pathway in tamoxifen-resistant MCF-7 cells, *Breast Cancer Res. Treat.* 81 (2003) 81–93.
- [85] J. Mendelsohn, J. Baselga, The EGF receptor family as targets for cancer therapy, *Oncogene* 19 (2000) 6550–6565.
- [86] R.I. Yarden, M.A. Wilson, S.A. Chrysogelos, Estrogen suppression of EGFR expression in breast cancer cells: a possible mechanism to modulate growth, *J. Cell. Biochem.* 81 (2001) 232–246.
- [87] J.M. Gee, M.E. Harper, I.R. Hutcheson, T.A. Madden, D. Barrow, J.M. Knowlden, et al., The antiepidermal growth factor receptor agent gefitinib (ZD1839/Iressa) improves antihormone response and prevents development of resistance in breast cancer *in vitro*, *Endocrinology* 144 (2003) 5105–5117.
- [88] S. Okubo, J. Kurebayashi, T. Otsuki, Y. Yamamoto, K. Tanaka, H. Sonoo, Additive antitumour effect of the epidermal growth factor receptor tyrosine kinase inhibitor gefitinib (Iressa, ZD1839) and the antioestrogen fulvestrant (faslodex, ICI 182,780) in breast cancer cells, *Br. J. Cancer* 90 (2004) 236–244.
- [89] R. Kumar, M. Mandal, A. Lipton, H. Harvey, C.B. Thompson, Overexpression of HER2 modulates Bcl-2, Bcl-xL, and tamoxifen-induced apoptosis in human MCF-7 breast cancer cells, *Clin. Cancer Res.* 2 (1996) 1215–1219.
- [90] J. Shou, S. Massarweh, C.K. Osborne, A.E. Wakeling, S. Ali, H. Weiss, et al., Mechanisms of tamoxifen resistance: increased estrogen receptor-HER2/neu cross-talk in ER/HER2-positive breast cancer, *J. Natl. Cancer Inst.* 96 (2004) 926–935.
- [91] H. Lee, W. Bai, Regulation of estrogen receptor nuclear export by ligand-induced and p38-mediated receptor phosphorylation, *Mol. Cell. Biol.* 22 (2002) 5835–5845.
- [92] R.B. Riggins, K.S. Thomas, H.Q. Ta, J. Wen, R.J. Davis, N.R. Schuh, et al., Physical and functional interactions between Cas and c-Src induce tamoxifen resistance of breast cancer cells through pathways involving epidermal growth factor receptor and signal transducer and activator of transcription 5b, *Cancer Res.* 66 (2006) 7007–7015.
- [93] J.N. Holloway, S. Murthy, D. El-Ashry, A cytoplasmic substrate of mitogen-activated protein kinase is responsible for estrogen receptor- α down-regulation in breast cancer cells: the role of nuclear factor- κ B, *Mol. Endocrinol.* 18 (2004) 1396–1410.
- [94] R. Schiff, S.A. Massarweh, J. Shou, L. Bharwani, G. Arpino, M. Rimawi, et al., Advanced concepts in estrogen receptor biology and breast cancer endocrine resistance: implicated role of growth factor signaling and estrogen receptor coregulators, *Cancer Chemother. Pharmacol.* 56 (Suppl. 1) (2005) 10–20.
- [95] Z. Yang, C.J. Barnes, R. Kumar, Human epidermal growth factor receptor 2 status modulates subcellular localization of and interaction with estrogen receptor α in breast cancer cells, *Clin. Cancer Res.* 10 (2004) 3621–3628.
- [96] S. Massarweh, C.K. Osborne, S. Jiang, A.E. Wakeling, M. Rimawi, S.K. Mohsin, et al., Mechanisms of tumor regression and resistance to estrogen deprivation and fulvestrant in a model of estrogen receptor-positive, HER-2/neu-positive breast cancer, *Cancer Res.* 66 (2006) 8266–8273.
- [97] B.L. Brockdorff, I. Heiberg, A.E. Lykkesfeldt, Resistance to different antiestrogens is caused by different multifactorial changes and is associated with reduced expression of IGF receptor 1 α , *Endocr. Relat. Cancer* 10 (2003) 579–590.
- [98] J.M. Knowlden, I.R. Hutcheson, D. Barrow, J.M. Gee, R.I. Nicholson, Insulin-like growth factor-1 receptor signaling in tamoxifen-resistant breast cancer: a supporting role to the epidermal growth factor receptor, *Endocrinology* 146 (2005) 4609–4618.
- [99] R.J. Santen, R.X. Song, Z. Zhang, R. Kumar, M.H. Jeng, A. Masamura, et al., Long-term estradiol deprivation in breast cancer cells up-regulates growth factor signaling and enhances estrogen sensitivity, *Endocr. Relat. Cancer* 12 (Suppl. 1) (2005) S61–S73.
- [100] G. Cesarone, C. Garofalo, M.T. Abrams, O. Igoucheva, V. Alexeev, K. Yoon, et al., RNAi-mediated silencing of insulin receptor substrate 1 (IRS-1) enhances tamoxifen-induced cell death in MCF-7 breast cancer cells, *J. Cell Biochem.* 98 (2006) 440–450.
- [101] L.C. Dorssers, T. van Agthoven, A. Dekker, T.L. van Agthoven, E.M. Kok, Induction of antiestrogen resistance in human breast cancer cells by random insertional mutagenesis using defective retroviruses: identification of BCAR-1, a common integration site, *Mol. Endocrinol.* 7 (1993) 870–878.
- [102] T. van Agthoven, T.L. van Agthoven, A. Dekker, P.J. van der Spek, L. Vreede, L.C. Dorssers, Identification of BCAR3 by a random search for genes involved in antiestrogen resistance of human breast cancer cells, *EMBO J.* 17 (1998) 2799–2808.

- [103] A.H. Bouton, R.B. Riggins, P.J. Bruce-Staskal, Functions of the adapter protein Cas: signal convergence and the determination of cellular responses, *Oncogene* 20 (2001) 6448–6458.
- [104] P. Defilippi, P. Di Stefano, S. Cabodi, p130cas: a versatile scaffold in signaling networks, *Trends Cell Biol.* 16 (2006) 257–263.
- [105] A. Brinkman, S. van der Flier, E.M. Kok, L.C. Dorssers, BCAR1, a human homologue of the adapter protein p130^{Cas}, and antiestrogen resistance in breast cancer cells, *J. Natl. Cancer Inst.* 92 (2000) 112–120.
- [106] M.R. Burnham, P.J. Bruce-Staskal, M.T. Harte, C.L. Weidow, A. Ma, S.A. Weed, et al., Regulation of c-Src activity and function by the adapter protein Cas, *Mol. Cell Biol.* 20 (2000) 5865–5878.
- [107] R.B. Riggins, L.A. Quilliam, A.H. Bouton, Synergistic promotion of c-Src activation and cell migration by Cas and AND-34/BCAR3, *J. Biol. Chem.* 278 (2003) 28264–28273.
- [108] S. Cabodi, L. Moro, G. Baj, M. Smeriglio, P. Di Stefano, S. Gippone, et al., p130cas interacts with estrogen receptor alpha and modulates non-genomic estrogen signaling in breast cancer cells, *J. Cell Sci.* 117 (2004) 1603–1611.
- [109] M.D. Planas-Silva, K.N. Hamilton, Targeting c-Src kinase enhances tamoxifen's inhibitory effect on cell growth by modulating expression of cell cycle and survival proteins, *Cancer Chemother. Pharmacol.*, in press.
- [110] L.N. Cowell, J.D. Graham, A.H. Bouton, C.L. Clarke, G.M. O'Neill, Tamoxifen treatment promotes phosphorylation of the adhesion molecules, p130^{Cas}/BCAR1, Fak and Src, via an adhesion-dependent pathway, *Oncogene* 25 (2006) 7597–7607.
- [111] A.E. Ottenhoff-Kalf, G. Rijksen, E.A. van Beurden, A. Hennipman, A.A. Michels, G.E. Staal, Characterization of protein tyrosine kinases from human breast cancer: Involvement of the c-Src oncogene product, *Cancer Res.* 52 (1992) 4773–4778.
- [112] B.S. Verbeek, T.M. Vroom, S.S. Adriaansen-Slot, A.E. Ottenhoff-Kalf, J.G. Geertzema, A. Hennipman, et al., c-Src protein expression is increased in human breast cancer. An immunohistochemical and biochemical analysis, *J. Pathol.* 180 (1996) 383–388.
- [113] S. van der Flier, C.M. Chan, A. Brinkman, M. Smid, S.R. Johnston, L.C. Dorssers, et al., BCAR1/p130^{Cas} expression in untreated and acquired tamoxifen-resistant human breast carcinomas, *Int. J. Cancer* 89 (2000) 465–468.
- [114] J.S. Biscardi, A.P. Belsches, S.J. Parsons, Characterization of human epidermal growth factor receptor and c-Src interactions in human breast tumor cells, *Mol. Carcinog.* 21 (1998) 261–272.
- [115] A.P. Belsches-Jablonski, J.S. Biscardi, D.R. Peavy, D.A. Tice, D.A. Romney, S.J. Parsons, Src family kinases and HER2 interactions in human breast cancer cell growth and survival, *Oncogene* 20 (2001) 1465–1475.
- [116] T. Gotoh, D. Cai, X. Tian, L.A. Feig, A. Lerner, p130cas regulates the activity of AND-34, a novel Ral, Rap1, and R-Ras guanine nucleotide exchange factor, *J. Biol. Chem.* 275 (2000) 30118–30123.
- [117] D. Cai, K.N. Felekis, R.I. Near, G.M. O'Neill, J.M. van Seventer, E.A. Golemis, et al., The GDP exchange factor AND-34 is expressed in B cells, associates with HEF1, and activates Cdc42, *J. Immunol.* 170 (2003) 969–978.
- [118] K.N. Felekis, R.P. Narsimhan, R. Near, A.F. Castro, Y. Zheng, L.A. Quilliam, et al., AND-34 activates phosphatidylinositol 3-kinase and induces anti-estrogen resistance in a SH2 and GDP exchange factor-like domain-dependent manner, *Mol. Cancer Res.* 3 (2005) 32–41.
- [119] K. Felekis, L.A. Quilliam, A. Lerner, Characterization of AND-34 function and signaling, *Methods Enzymol.* 407 (2005) 55–63.
- [120] Y. Yu, L.A. Feig, Involvement of R-Ras and Ral GTPases in estrogen-independent proliferation of breast cancer cells, *Oncogene* 21 (2002) 7557–7568.
- [121] A. Toker, M. Yoeli-Lerner, AKT signaling and cancer: surviving but not moving on, *Cancer Res.* 66 (2006) 3963–3966.
- [122] D. Kim, J. Chung, AKT: versatile mediator of cell survival and beyond, *J. Biochem. Mol. Biol.* 35 (2002) 106–115.
- [123] R.A. Campbell, P. Bhat-Nakshatri, N.M. Patel, D. Constantinidou, S. Ali, H. Nakshatri, Phosphatidylinositol 3-kinase/AKT-mediated activation of estrogen receptor alpha: a new model for anti-estrogen resistance, *J. Biol. Chem.* 276 (2001) 9817–9824.
- [124] B.N. Duong, S. Elliott, D.E. Frigo, L.I. Melnik, L. Vanhoy, S. Tomchuck, et al., AKT regulation of estrogen receptor β transcriptional activity in breast cancer, *Cancer Res.* 66 (2006) 8373–8381.
- [125] J. Faridi, L. Wang, G. Endemann, R.A. Roth, Expression of constitutively active AKT-3 in MCF-7 breast cancer cells reverses the estrogen and tamoxifen responsiveness of these cells *in vivo*, *Clin. Cancer Res.* 9 (2003) 2933–2939.
- [126] L.A. DeGraffenried, W.E. Friedrichs, L. Fulcher, G. Fernandes, J.M. Silva, J.M. Peralba, et al., Eicosapentaenoic acid restores tamoxifen sensitivity in breast cancer cells with high AKT activity, *Ann. Oncol.* 14 (2003) 1051–1056.
- [127] A.S. Clark, K. West, S. Streicher, P.A. Dennis, Constitutive and inducible AKT activity promotes resistance to chemotherapy, trastuzumab, or tamoxifen in breast cancer cells, *Mol. Cancer Ther.* 1 (2002) 707–717.
- [128] N.J. Jordan, J.M. Gee, D. Barrow, A.E. Wakeling, R.I. Nicholson, Increased constitutive activity of PKB/AKT in tamoxifen resistant breast cancer MCF-7 cells, *Breast Cancer Res. Treat.* 87 (2004) 167–180.
- [129] T. Frogne, J.S. Jepsen, S.S. Larsen, C.K. Fog, B.L. Brockdorff, A.E. Lykkesfeldt, Antiestrogen-resistant human breast cancer cells require activated protein kinase B/AKT for growth, *Endocr. Relat. Cancer* 12 (2005) 599–614.
- [130] A. Brodie, G. Sabnis, D. Jelovac, Aromatase and breast cancer, *J. Steroid Biochem. Mol. Biol.* 102 (2006) 97–102.
- [131] Y. Mamane, E. Petroulakis, O. LeBacquer, N. Sonenberg, mTOR, translation initiation and cancer, *Oncogene* 25 (2006) 6416–6422.
- [132] L.A. DeGraffenried, W.E. Friedrichs, D.H. Russell, E.J. Donzis, A.K. Middleton, J.M. Silva, et al., Inhibition of mTOR activity restores tamoxifen response in breast cancer cells with aberrant AKT activity, *Clin. Cancer Res.* 10 (2004) 8059–8067.
- [133] A. Boulay, J. Rudloff, J. Ye, S. Zumstein-Mecker, T. O'Reilly, D.B. Evans, et al., Dual inhibition of mTOR and estrogen receptor signaling in vitro induces cell death in models of breast cancer, *Clin. Cancer. Res.* 11 (2005) 5319–5328.

- [134] H. Carraway, M. Hidalgo, New targets for therapy in breast cancer: mammalian target of rapamycin (mTOR) antagonists, *Breast Cancer Res.* 6 (2004) 219–224.
- [135] S.R. Johnston, Targeting downstream effectors of epidermal growth factor receptor/HER2 in breast cancer with either farnesyltransferase inhibitors or mTOR antagonists, *Int. J. Gynecol. Cancer* 16 (Suppl. 2) (2006) 543–548.
- [136] D.A. Lannigan, Estrogen receptor phosphorylation, *Steroids* 68 (2003) 1–9.
- [137] L.C. Murphy, G.E. Weitsman, G.P. Skliris, E.M. Teh, L. Li, B. Peng, et al., Potential role of estrogen receptor alpha (ER alpha) phosphorylated at serine(118) in human breast cancer *in vivo*, *J. Steroid Biochem. Mol. Biol.* 102 (2006) 139–146.
- [138] S.F. Arnold, J.D. Obourn, H. Jaffe, A.C. Notides, Phosphorylation of the human estrogen receptor on tyrosine 537 *in vivo* and by Src family tyrosine kinases *in vitro*, *Mol. Endocrinol.* 9 (1995) 24–33.
- [139] M.R. Yudt, D. Vorojeikina, L. Zhong, D.F. Skafar, S. Sasson, T.A. Gasiewicz, et al., Function of estrogen receptor tyrosine 537 in hormone binding, DNA binding, and transactivation, *Biochemistry* 38 (1999) 14146–14156.
- [140] K.E. Weis, K. Ekena, J.A. Thomas, G. Lazennec, B.S. Katzenellenbogen, Constitutively active human estrogen receptors containing amino acid substitutions for tyrosine 537 in the receptor protein, *Mol. Endocrinol.* 10 (1996) 1388–1398.
- [141] G.B. Tremblay, A. Tremblay, F. Labrie, V. Giguere, Ligand-independent activation of the estrogen receptors alpha and beta by mutations of a conserved tyrosine can be abolished by antiestrogens, *Cancer Res.* 58 (1998) 877–881.
- [142] I. Rogatsky, J.M. Trowbridge, M.J. Garabedian, Potentiation of human estrogen receptor alpha transcriptional activation through phosphorylation of serines 104 and 106 by the cyclin A-CDK2 complex, *J. Biol. Chem.* 274 (1999) 22296–22302.
- [143] R. Michalides, H. van Tinteren, A. Balkenende, J.B. Vermorken, J. Benraadt, J. Huldij, et al., Cyclin A is a prognostic indicator in early stage breast cancer with and without tamoxifen treatment, *Br. J. Cancer* 86 (2002) 402–408.
- [144] S.F. Arnold, J.D. Obourn, H. Jaffe, A.C. Notides, Serine 167 is the major estradiol-induced phosphorylation site on the human estrogen receptor, *Mol. Endocrinol.* 8 (1994) 1208–1214.
- [145] S. Glaros, N. Atanaskova, C. Zhao, D.F. Skafar, K.B. Reddy, Activation function-1 domain of estrogen receptor regulates the agonistic and antagonistic actions of tamoxifen, *Mol. Endocrinol.* 20 (2006) 996–1008.
- [146] V.S. Likhite, F. Stossi, K. Kim, B.S. Katzenellenbogen, J.A. Katzenellenbogen, Kinase-specific phosphorylation of the estrogen receptor changes receptor interactions with ligand, deoxyribonucleic acid, and coregulators associated with alterations in estrogen and tamoxifen activity, *Mol. Endocrinol.* 20 (2006) 3120–3132.
- [147] D. Chen, P.E. Pace, R.C. Coombes, S. Ali, Phosphorylation of human estrogen receptor alpha by protein kinase A regulates dimerization, *Mol. Cell. Biol.* 19 (1999) 1002–1015.
- [148] R.A. Wang, A. Mazumdar, R.K. Vadlamudi, R. Kumar, p21-activated kinase-1 phosphorylates and transactivates estrogen receptor-alpha and promotes hyperplasia in mammary epithelium, *EMBO J.* 21 (2002) 5437–5447.
- [149] S. Balasenthil, C.J. Barnes, S.K. Rayala, R. Kumar, Estrogen receptor activation at serine 305 is sufficient to upregulate cyclin D1 in breast cancer cells, *FEBS Lett.* 567 (2004) 243–247.
- [150] S.K. Rayala, P.R. Molli, R. Kumar, Nuclear p21-activated kinase 1 in breast cancer packs off tamoxifen sensitivity, *Cancer Res.* 66 (2006) 5985–5988.
- [151] S.K. Rayala, A.H. Talukder, S. Balasenthil, R. Tharakan, C.J. Barnes, R.A. Wang, et al., p21-activated kinase 1 regulation of estrogen receptor-alpha activation involves serine 305 activation linked with serine 118 phosphorylation, *Cancer Res.* 66 (2006) 1694–1701.
- [152] R. Michalides, A. Griekspoor, A. Balkenende, D. Verwoerd, L. Janssen, K. Jalink, et al., Tamoxifen resistance by a conformational arrest of the estrogen receptor alpha after PKA activation in breast cancer, *Cancer Cell* 5 (2004) 597–605.
- [153] P.B. Joel, A.M. Traish, D.A. Lannigan, Estradiol-induced phosphorylation of serine 118 in the estrogen receptor is independent of p42/p44 mitogen-activated protein kinase, *J. Biol. Chem.* 273 (1998) 13317–13323.
- [154] D. Chen, T. Riedl, E. Washbrook, P.E. Pace, R.C. Coombes, J.M. Egly, et al., Activation of estrogen receptor alpha by S118 phosphorylation involves a ligand-dependent interaction with TFIIH and participation of CDK7, *Mol. Cell* 6 (2000) 127–137.
- [155] D. Chen, E. Washbrook, N. Sarwar, G.J. Bates, P.E. Pace, V. Thirunuvakkarasu, et al., Phosphorylation of human estrogen receptor alpha at serine 118 by two distinct signal transduction pathways revealed by phosphorylation-specific antisera, *Oncogene* 21 (2002) 4921–4931.
- [156] S. Medunjanin, A. Hermani, B. De Servi, J. Grisouard, G. Rincke, D. Mayer, Glycogen synthase kinase-3 interacts with and phosphorylates estrogen receptor alpha and is involved in the regulation of receptor activity, *J. Biol. Chem.* 280 (2005) 33006–33014.
- [157] D.J. Britton, I.R. Hutcheson, J.M. Knowlden, D. Barrow, M. Giles, R.A. McClelland, et al., Bidirectional cross talk between ER alpha and EGFR signalling pathways regulates tamoxifen-resistant growth, *Breast Cancer Res. Treat.* 96 (2006) 131–146.
- [158] G.E. Weitsman, L. Li, G.P. Skliris, J.R. Davie, K. Ung, Y. Niu, et al., Estrogen receptor-alpha phosphorylated at ser118 is present at the promoters of estrogen-regulated genes and is not altered due to HER-2 overexpression, *Cancer Res.* 66 (2006) 10162–10170.
- [159] K.J. Park, V. Krishnan, B.W. O'Malley, Y. Yamamoto, R.B. Gaynor, Formation of an IKK alpha-dependent transcription complex is required for estrogen receptor-mediated gene activation, *Mol. Cell* 18 (2005) 71–82.
- [160] C.E. Caldon, R.J. Daly, R.L. Sutherland, E.A. Musgrove, Cell cycle control in breast cancer cells, *J. Cell. Biochem.* 97 (2006) 261–274.
- [161] A.J. Butt, C.M. McNeil, E.A. Musgrove, R.L. Sutherland, Downstream targets of growth factor and oestrogen signalling and endocrine resistance: the potential roles of c-Myc, cyclin D1 and cyclin E, *Endocr. Relat. Cancer* 12 (Suppl. 1) (2005) S47–S59.
- [162] C.J. Sherr, J.M. Roberts, CDK inhibitors: positive and negative regulators of G1-phase progression, *Genes Dev.* 13 (1999) 1501–1512.

- [163] C.J. Sherr, J.M. Roberts, Living with or without cyclins and cyclin-dependent kinases, *Genes Dev.* 18 (2004) 2699–2711.
- [164] R. Hui, G.L. Finney, J.S. Carroll, C.S. Lee, E.A. Musgrove, R.L. Sutherland, Constitutive overexpression of cyclin D1 but not cyclin E confers acute resistance to antiestrogens in T-47D breast cancer cells, *Cancer Res.* 62 (2002) 6916–6923.
- [165] R.L. Kilker, M.W. Hartl, T.M. Rutherford, M.D. Planas-Silva, Cyclin D1 expression is dependent on estrogen receptor function in tamoxifen-resistant breast cancer cells, *J. Steroid Biochem. Mol. Biol.* 92 (2004) 63–71.
- [166] R.M. O'Regan, C. Osipo, E. Ariazi, E.S. Lee, K. Meeke, C. Morris, et al., Development and therapeutic options for the treatment of raloxifene-stimulated breast cancer in athymic mice, *Clin. Cancer Res.* 12 (2006) 2255–2263.
- [167] R.L. Kilker, M.D. Planas-Silva, Cyclin D1 is necessary for tamoxifen-induced cell cycle progression in human breast cancer cells, *Cancer Res.* 66 (2006) 11478–11484.
- [168] R.L. Sutherland, E.A. Musgrove, Cyclins and breast cancer, *J. Mammary Gland Biol. Neoplasia* 9 (2004) 95–104.
- [169] J.S. Foster, D.C. Henley, S. Ahamed, J. Wimalasena, Estrogens and cell-cycle regulation in breast cancer, *Trends Endocrinol. Metab.* 12 (2001) 320–327.
- [170] S. Akli, K. Keyomarsi, Low-molecular-weight cyclin E: the missing link between biology and clinical outcome, *Breast Cancer Res.* 6 (2004) 188–191.
- [171] S. Akli, P.J. Zheng, A.S. Multani, H.F. Wingate, S. Pathak, N. Zhang, et al., Tumor-specific low molecular weight forms of cyclin E induce genomic instability and resistance to p21, p27, and antiestrogens in breast cancer, *Cancer Res.* 64 (2004) 3198–3208.
- [172] N.K. Dhillon, M. Mudryj, Ectopic expression of cyclin E in estrogen responsive cells abrogates antiestrogen mediated growth arrest, *Oncogene* 21 (2002) 4626–4634.
- [173] S. Cariou, J.C. Donovan, W.M. Flanagan, A. Milic, N. Bhattacharya, J.M. Slingerland, Down-regulation of p21^{WAF1/CIP1} or p27^{Kip1} abrogates antiestrogen-mediated cell cycle arrest in human breast cancer cells, *Proc. Natl. Acad. Sci. USA* 97 (2000) 9042–9046.
- [174] J.S. Carroll, D.K. Lynch, A. Swarbrick, J.M. Renoir, B. Sarcevic, R.J. Daly, et al., p27(Kip1) induces quiescence and growth factor insensitivity in tamoxifen-treated breast cancer cells, *Cancer Res.* 63 (2003) 4322–4326.
- [175] J. Liang, J.M. Slingerland, Multiple roles of the PI3K/PKB (AKT) pathway in cell cycle progression, *Cell Cycle* 2 (2003) 339–345.
- [176] S. Mukherjee, S.E. Conrad, c-Myc suppresses p21^{WAF1/CIP1} expression during estrogen signaling and antiestrogen resistance in human breast cancer cells, *J. Biol. Chem.* 280 (2005) 17617–17625.
- [177] J.C. Donovan, A. Milic, J.M. Slingerland, Constitutive MEK/MAPK activation leads to p27(Kip1) deregulation and antiestrogen resistance in human breast cancer cells, *J. Biol. Chem.* 276 (2001) 40888–40895.
- [178] M. Dowsett, J. Houghton, C. Iden, J. Salter, J. Farndon, R. A'Hern, et al., Benefit from adjuvant tamoxifen therapy in primary breast cancer patients according to estrogen receptor, progesterone receptor, EGF receptor and HER2 status, *Ann. Oncol.* 17 (2006) 818–826.
- [179] S. Hiscox, L. Morgan, D. Barrow, C. Dutkowskil, A. Wakeling, R.I. Nicholson, Tamoxifen resistance in breast cancer cells is accompanied by an enhanced motile and invasive phenotype: Inhibition by gefitinib ('Iressa', ZD1839), *Clin. Exp. Metastasis* 21 (2004) 201–212.
- [180] Y. Kang, J. Massague, Epithelial-mesenchymal transitions: twist in development and metastasis, *Cell* 118 (2004) 277–279.
- [181] S. Hiscox, L. Morgan, T.P. Green, D. Barrow, J. Gee, R.I. Nicholson, Elevated Src activity promotes cellular invasion and motility in tamoxifen resistant breast cancer cells, *Breast Cancer Res. Treat.* 97 (2006) 263–274.
- [182] J.M. Lee, S. Dedhar, R. Kalluri, E.W. Thompson, The epithelial-mesenchymal transition: new insights in signaling, development, and disease, *J. Cell Biol.* 172 (2006) 973–981.
- [183] P. Schraml, G. Schwerdtfeger, F. Burkhalter, A. Raggi, D. Schmidt, T. Ruffalo, et al., Combined array comparative genomic hybridization and tissue microarray analysis suggest Pak1 at 11q13.5-q14 as a critical oncogene target in ovarian carcinoma, *Am. J. Pathol.* 163 (2003) 985–992.
- [184] E.C. Robanus-Maandag, C.A. Bosch, P.M. Kristel, A.A. Hart, I.F. Faneyte, P.M. Nederlof, et al., Association of c-Myc amplification with progression from the in situ to the invasive stage in c-Myc-amplified breast carcinomas, *J. Pathol.* 201 (2003) 75–82.
- [185] E. Tokunaga, Y. Kimura, K. Mashino, E. Oki, A. Kataoka, S. Ohno, et al., Activation of PI3K/AKT signaling and hormone resistance in breast cancer, *Breast Cancer* 13 (2006) 137–144.
- [186] N. Shoman, S. Klassen, A. McFadden, M.G. Bickis, E. Torlakovic, R. Chibbar, Reduced PTEN expression predicts relapse in patients with breast carcinoma treated by tamoxifen, *Mod. Pathol.* 18 (2005) 250–259.
- [187] F.S. Kenny, R. Hui, E.A. Musgrove, J.M. Gee, R.W. Blamey, R.I. Nicholson, et al., Overexpression of cyclin D1 messenger RNA predicts for poor prognosis in estrogen receptor-positive breast cancer, *Clin. Cancer Res.* 5 (1999) 2069–2076.
- [188] M. Stendahl, A. Kronblad, L. Ryden, S. Emdin, N.O. Bengtsson, G. Landberg, Cyclin D1 overexpression is a negative predictive factor for tamoxifen response in postmenopausal breast cancer patients, *Br. J. Cancer* 90 (2004) 1942–1948.
- [189] P.N. Span, V.C. Tjan-Heijnen, P. Manders, L.V. Beex, C.G. Sweep, Cyclin-E is a strong predictor of endocrine therapy failure in human breast cancer, *Oncogene* 22 (2003) 4898–4904.
- [190] C. Desmedt, F.E. Ouriaghli, V. Durbecq, A. Soree, M.A. Colozza, E. Azambuja, et al., Impact of cyclins E, neutrophil elastase and proteinase 3 expression levels on clinical outcome in primary breast cancer patients, *Int. J. Cancer* 119 (2006) 2539–2545.
- [191] G. Pohl, M. Rudas, O. Dietze, S. Lax, E. Markis, R. Pirker, et al., High p27^{Kip1} expression predicts superior relapse-free and overall survival for premenopausal women with early-stage breast cancer receiving adjuvant treatment with tamoxifen plus goserelin, *J. Clin. Oncol.* 21 (2003) 3594–3600.
- [192] G. Perez-Tenorio, F. Berglund, A. Esguerra Merca, B. Nordenskjold, L.E. Rutqvist, L. Skoog, et al., Cytoplasmic p21^{WAF1/CIP1} correlates with AKT activation and poor response to tamoxifen in breast cancer, *Int. J. Oncol.* 28 (2006) 1031–1042.
- [193] L.C. Murphy, Y. Niu, L. Snell, P. Watson, Phospho-serine-118 estrogen receptor- α expression is associated with

- better disease outcome in women treated with tamoxifen, *Clin. Cancer Res.* 10 (2004) 5902–5906.
- [194] J. Bergqvist, G. Elmberger, J. Ohl, B. Linderholm, J. Bjohle, H. Hellborg, et al., Activated ERK1/2 and phosphorylated oestrogen receptor alpha are associated with improved breast cancer survival in women treated with tamoxifen, *Eur. J. Cancer* 42 (2006) 1104–1112.
- [195] H. Yamashita, M. Nishio, S. Kobayashi, Y. Ando, H. Sugiura, Z. Zhang, et al., Phosphorylation of estrogen receptor alpha serine 167 is predictive of response to endocrine therapy and increases postrelapse survival in metastatic breast cancer, *Breast Cancer Res.* 7 (2005) R753–R764.
- [196] V.J. Bardou, G. Arpino, R.M. Elledge, C.K. Osborne, G.M. Clark, Progesterone receptor status significantly improves outcome prediction over estrogen receptor status alone for adjuvant endocrine therapy in two large breast cancer databases, *J. Clin. Oncol.* 21 (2003) 1973–1979.
- [197] M. Ferno, O. Stal, B. Baldetorp, T. Hatschek, A.C. Kallstrom, P. Malmstrom, et al., Results of two or five years of adjuvant tamoxifen correlated to steroid receptor and S-phase levels. South Sweden Breast Cancer Group, and South-East Sweden Breast Cancer Group, *Breast Cancer Res. Treat.* 59 (2000) 69–76.
- [198] M.J. Ellis, A. Coop, B. Singh, Y. Tao, A. Llombart-Cussac, F. Janicke, et al., Letrozole inhibits tumor proliferation more effectively than tamoxifen independent of HER1/2 expression status, *Cancer Res.* 63 (2003) 6523–6531.
- [199] M.P. Jansen, J.A. Foekens, I.L. van Staveren, M.M. Dirkwager-Kiel, K. Ritstier, M.P. Look, et al., Molecular classification of tamoxifen-resistant breast carcinomas by gene expression profiling, *J. Clin. Oncol.* 23 (2005) 732–740.
- [200] M.P. Goetz, V.J. Suman, J.N. Ingle, A.M. Nibbe, D.W. Visscher, C.A. Reynolds, et al., A two-gene expression ratio of homeobox 13 and interleukin-17B receptor for prediction of recurrence and survival in women receiving adjuvant tamoxifen, *Clin. Cancer Res.* 12 (2006) 2080–2087.
- [201] J.F. Reid, L. Lusa, L. De Cecco, D. Coradini, S. Veneroni, M.G. Daidone, et al., Limits of predictive models using microarray data for breast cancer clinical treatment outcome, *J. Natl. Cancer Inst.* 97 (2005) 927–930.
- [202] J. Frasor, F. Stossi, J.M. Danes, B. Komm, C.R. Lyttle, B.S. Katzenellenbogen, Selective estrogen receptor modulators: discrimination of agonistic versus antagonistic activities by gene expression profiling in breast cancer cells, *Cancer Res.* 64 (2004) 1522–1533.
- [203] J. Hur, J. Chesnes, K.R. Coser, R.S. Lee, P. Geck, K.J. Isselbacher, et al., The Bik BH3-only protein is induced in estrogen-starved and antiestrogen-exposed breast cancer cells and provokes apoptosis, *Proc. Natl. Acad. Sci. USA* 101 (2004) 2351–2356.
- [204] T. Itoh, K. Karlsberg, I. Kijima, Y.C. Yuan, D. Smith, J. Ye, et al., Letrozole-, anastrozole- and tamoxifen-responsive genes in MCF-7aro cells: a microarray approach, *Mol. Cancer Res.* 3 (2005) 203–218.
- [205] M. Becker, A. Sommer, J.R. Kratzschmar, H. Seidel, H.D. Pohlenz, I. Fichtner, Distinct gene expression patterns in a tamoxifen-sensitive human mammary carcinoma xenograft and its tamoxifen-resistant subline MaCa 3366/TAM, *Mol. Cancer Ther.* 4 (2005) 151–168.
- [206] M.L. Bowie, E.C. Dietze, J. Delrow, G.R. Bean, M.M. Troch, R.J. Marjoram, et al., Interferon-regulatory factor-1 is critical for tamoxifen-mediated apoptosis in human mammary epithelial cells, *Oncogene* 23 (2004) 8743–8755.
- [207] K. Xu, C.L. Geczy, IFN-gamma and TNF regulate macrophage expression of the chemotactic S100 protein S100A8, *J. Immunol.* 164 (2000) 4916–4923.
- [208] S.S. Jeffrey, P.E. Lonning, B.E. Hillner, Genomics-based prognosis and therapeutic prediction in breast cancer, *J. Natl. Compr. Canc. Netw.* 3 (2005) 291–300.
- [209] C. Fan, D.S. Oh, L. Wessels, B. Weigelt, D.S. Nuyten, A.B. Nobel, et al., Concordance among gene-expression-based predictors for breast cancer, *N. Engl. J. Med.* 355 (2006) 560–569.
- [210] S. Paik, Methods for gene expression profiling in clinical trials of adjuvant breast cancer therapy, *Clin. Cancer Res.* 12 (2006) 1019S–1023S.
- [211] M. Dowsett, R.I. Nicholson, R.J. Pietras, Biological characteristics of the pure antiestrogen fulvestrant: overcoming endocrine resistance, *Breast Cancer Res. Treat.* 93 Suppl. 1 (2005) S11–S18.
- [212] S.R. Johnston, Clinical efforts to combine endocrine agents with targeted therapies against epidermal growth factor receptor/human epidermal growth factor receptor 2 and mammalian target of rapamycin in breast cancer, *Clin. Cancer Res.* 12 (2006) 1061S–1068S.
- [213] J. Head, S.R. Johnston, New targets for therapy in breast cancer: farnesyltransferase inhibitors, *Breast Cancer Res.* 6 (2004) 262–268.
- [214] A. Leary, M. Dowsett, Combination therapy with aromatase inhibitors: the next era of breast cancer treatment?, *Br J. Cancer* 95 (2006) 661–666.
- [215] S. Chan, M.E. Scheulen, S. Johnston, K. Mross, F. Cardoso, C. Dittich, et al., Phase II study of temsirolimus (CCI-779), a novel inhibitor of mTOR, in heavily pre-treated patients with locally advanced or metastatic breast cancer, *J. Clin. Oncol.* 23 (2005) 5314–5322.
- [216] L. Perey, R. Paridaens, H. Hawle, K. Zaman, F. Nole, H. Wildiers, et al., Clinical benefit of fulvestrant in postmenopausal women with advanced breast cancer and primary or acquired resistance to aromatase inhibitors: final results of phase II Swiss Group for Clinical Cancer Research Trial (SAKK 21/00), *Ann. Oncol.* 18 (2007) 64–69.

BIOGRAPHICAL SKETCH

Provide the following information for the key personnel and other significant contributors in the order listed on Form Page 2.

Follow this format for each person. **DO NOT EXCEED FOUR PAGES.**

NAME Rebecca B. Riggins	POSITION TITLE Research Assistant Professor of Oncology		
eRA COMMONS USER NAME rbiggins			
EDUCATION/TRAINING (<i>Begin with baccalaureate or other initial professional education, such as nursing, and include postdoctoral training.</i>)			
INSTITUTION AND LOCATION	DEGREE (if applicable)	YEAR(s)	FIELD OF STUDY
Hood College, Frederick MD, USA	B.A.	1998	Biochemistry, <i>cum laude</i>
University of Virginia, Charlottesville VA, USA	Ph.D.	2003	Microbiology
Georgetown University, Washington DC, USA	postdoc	2003-2006	Oncology

A. Positions and HonorsPositions

2007 – present Research Assistant Professor, Division of Molecular Endocrinology, Nutrition, and Obesity, Dept. of Oncology, Georgetown University, Washington DC

Honors

7/2001 Selected for the Edward A. Smuckler Pathobiology of Cancer Workshop, Keystone CO (sponsored by AACR)

7/2005 Selected for the 2005 Susan G. Komen Breast Cancer Career Mentoring Program for Postdoctoral Fellows. 4th Annual Future of Breast Cancer Congress: Individualizing Breast Cancer Treatment, Southampton, Bermuda

Other Experience and Professional Memberships

2001 – present Member, American Association for Cancer Research (AACR) and Women in Cancer Research (WICR)

2005 – present Manuscript reviewer, *Cancer Research*, *Clinical Cancer Research* and *Nutrition & Cancer*

2006 Grant reviewer, Dutch Cancer Society (KWF Kankerbestrijding)

B. Publications (in chronological order)Peer-Reviewed Original Articles

- 1.) **R.B. Riggins**, R.M. DeBerry, M.D. Toosarvandani, and A.H. Bouton. Association of Cas and Phosphatidylinositol 3-kinase (PI3K) in *crk*-transformed cells. ***Molecular Cancer Research***, 1: 428-37 (2003).
- 2.) **R.B. Riggins**, L.A. Quilliam, and A.H. Bouton. Synergistic promotion of c-Src activation and cell migration by Cas and AND-34/BCAR3. ***J. Biological Chemistry***, 278(30): 28264-73 (2003).
- 3.) K.B. Bouker, T.C. Skaar, D.R. Fernandez, K.A. O'Brien, **R.B. Riggins**, D. Cao and R. Clarke. Interferon regulatory factor-1 regulates the proapoptotic but not cell cycle arrest effects of the steroidal antiestrogen ICI 182,780 (Faslodex, Fulvestrant). ***Cancer Research***, 64: 4030-39 (2004).
- 4.) **R.B. Riggins**, A. Zwart, R. Nehra, and R. Clarke. The NFkB inhibitor parthenolide restores ICI 182,780 (Faslodex; Fulvestrant)-induced apoptosis in antiestrogen-resistant breast cancer cells. ***Molecular Cancer Therapeutics***, 4: 33-41 (2005).
- 5.) K.B. Bouker, T.C. Skaar, **R.B. Riggins**, D.S. Harburger, D.R. Fernandez, A. Zwart, A. Wang, and R. Clarke. Interferon regulatory factor-1 (IRF-1) exhibits tumor suppressor activities in breast cancer associated with caspase activation and induction of apoptosis. ***Carcinogenesis***, 26: 1527-35 (2005).

- 6.) ***R.B. Riggins**, K. Thomas, N. Schuh, S.S. Donelan, H. Ta, M.A. Gibson, M.A. Shupnik, C.M. Silva, R. Clarke, S.J. Parsons, and A.H. Bouton. Physical and functional interactions between Cas and c-Src induce Tamoxifen resistance of breast cancer cells through EGFR and STAT5b. **Cancer Research**, 66:7007-15 (2006). *article was featured in Cancer Research Highlights.
- 7.) K.B. Bouker, T.C. Skaar, D.S. Harburger, **R.B. Riggins**, D.R. Fernandez, A. Zwart, and R. Clarke. The A4396G Polymorphism in Interferon Regulatory Factor-1 is Frequently Expressed in Breast Cancer Cell Lines. **Cancer Genetics and Cytogenetics**, 175:61-4 (2007).
- 8.) R.S. Schrecengost, **R.B. Riggins**, K.S. Thomas, M.S. Guerrero, and A.H. Bouton. BCAR3 expression regulates breast cancer cell migration through promotion of p130Cas localization and membrane ruffling. **Cancer Research**, 67:6174-82 (2007).
- 9.) B.P. Gomez, **R.B. Riggins**, A.N. Shajahan, U. Klimach, A. Wang, A.C. Crawford, Y. Zhu, A. Zwart, M. Wang and R. Clarke. Human X-box binding protein-1 confers both estrogen-independence and antiestrogen resistance in breast cancer cell lines. **FASEB Journal** (2007 Jul 27, Epub ahead of print: <http://www.fasebj.org/cgi/content/abstract/fj.06-7990comv1>).

Reviews and Book Chapters

- 1.) A.H. Bouton, **R.B. Riggins**, and P.J. Bruce-Staskal. Functions of the Adapter Protein Cas: Signal Convergence and the Determination of Cellular Responses. **Oncogene**, 20: 6448-58 (2001).
- 2.) L. Hilakivi-Clarke, C. Wang, M. Kalil, **R. Riggins**, and R.G. Pestell. Cell cycle control in breast cancer: the role of dietary intervention. **Endocrine Related Cancer**, 11 (4): 603-22 (2004).
- 3.) **R.B. Riggins**, A.H. Bouton, M.C. Liu, and R. Clarke. Antiestrogens, aromatase inhibitors, and apoptosis in breast cancer. **Vitamins and Hormones**, 71:201-37 (2005).
- 4.) **R.B. Riggins**, R.S. Schrecengost, M.S. Guerrero, and A.H. Bouton. Pathways to Tamoxifen Resistance. **Cancer Letters** 256: 1-24 (2007).
- 5.) A.N. Shajahan, **R.B. Riggins**, and R. Clarke. Apoptosis, cell death, and breast cancer. **Breast Cancer: Prognosis, Treatment, and Prevention**, 2nd edition, (in press, 2007).

C. Research Support

Completed Research Support

(No # assigned) Riggins (PI) 7/2004 – 6/2005

Cancer Research Fellowship, Ladies Auxiliary to the Veterans of Foreign Wars, USA

“Role of IRF-1 in Breast Cancer Antiestrogen Resistance”

This project was designed to identify which apoptotic genes are affected by IRF-1 downregulation in breast cancer cells with acquired antiestrogen resistance.

Role: PI/Postdoctoral Fellow

PDF0503551 Clarke (PI) 5/2005 – 4/2007

Susan G. Komen Breast Cancer Foundation Postdoctoral Fellowship

“IRF-1-dependent apoptosis and antiestrogen sensitivity in breast cancer”

The primary goals of this project are to determine how IRF-1 controls apoptosis through the BCL2 gene family, and ascertain whether restoration of IRF-1 expression restores antiestrogen sensitivity to resistant breast cancer cells.

Role: Postdoctoral Fellow

BC051851 Riggins (PI) 9/2006 – 9/2007

Department of Defense Breast Cancer Research Program Concept Award

“ERR Gamma: Does an Orphan Nuclear Receptor Link Steroid Hormone Biogenesis to Endocrine Resistance?”

This study will determine whether ERRγ expression and activity affects cholesterol synthesis in breast cancer cells.

Role: PI