

AD _____

Award Number: W81XWH-06-1-0384

TITLE: Role of the PY Motif Containing Protein, WBP-2 in ER, PR Signaling and Breast Tumorigenesis

PRINCIPAL INVESTIGATOR: Sarath C. Dhananjayan
Zafar Nawaz

CONTRACTING ORGANIZATION: University of Miami
Miami, FL 33136

REPORT DATE: March 2008

TYPE OF REPORT: Annual Summary

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE 01-03-2008		2. REPORT TYPE Annual Summary		3. DATES COVERED 1 Mar 2007 – 28 Feb 2008	
4. TITLE AND SUBTITLE Role of the PY Motif Containing Protein, WBP-2 in ER, PR Signaling and Breast Tumorigenesis				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-06-1-0384	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Sarath C. Dhananjayan Zafar Nawaz Email: sdhananjayan@med.miami.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Miami Miami, FL 33136				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT Our data demonstrates that WBP-2 is recruited onto the hormone responsive promoters in the presence of hormone and it specifically enhances the transactivation functions of PR and ER. Our data also demonstrates that WBP-2 contains an intrinsic activation domain and the cPPXY of WBP-2 is essential for its coactivation and intrinsic activation functions. Our preliminary data also demonstrates that the WBP-2 binding protein, YAP1 enhances PR and ER transactivation but YAP1's coactivation function is absolutely dependent on WBP-2. Furthermore, cPPXY motif of WBP-2 and WW-domain of YAP1 is required for YAP1 to work as a transcriptional coactivator. Additionally, our data also indicate that the coactivation functions of WBP-2 and YAP1 are suppressed by WWOX1, suggesting that WWOX1 may regulate the transactivation functions of ER and PR by antagonizing the functions of WBP-2 and YAP1. Taken together our data established the role of WBP-2 and YAP1 as coactivators and WWOX1 as a repressor for ER and PR transactivation pathways.					
15. SUBJECT TERMS ESTROGEN RECEPTOR, PROGESTERONE RECEPTOR, WW-DOMAIN BINDING PROTEIN-2, YES-ASSOCIATED PROTEIN 1, WW-DOMAIN CONTAINING OXIDOREDUCTASE 1, CHROMATIN IMMUNOPRECIPITATION ASSAY					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			USAMRMC
			UU	29	19b. TELEPHONE NUMBER (include area code)

Table of Contents

	<u>Page</u>
Introduction.....	2-7
Body.....	8-13
Key Research Accomplishments.....	14
Reportable Outcomes.....	14
Conclusion.....	14
References.....	15-19
Appendix I	20-28

Introduction

Estrogen (ER) and Progesterone (PR) Receptors: Estrogen receptor- α (ER) and progesterone receptor (PR) are members of a superfamily of hormone-regulated transcription factors that stimulate gene expression in response to estrogens and progesterones respectively. These receptors contain common structural motifs, which include a less well-conserved amino-terminal activation function (AF-1) that affects transcription efficiency, a central DNA-binding domain, which mediates receptor binding to specific DNA enhancer sequences and determines target gene specificity, and a carboxy-terminal hormone-binding domain (HBD). The HBD contains activation function-2 (AF-2), the region that mediates the hormone-dependent activation function of receptors. In order to activate gene transcription, the ER and PR undergo a series of well-defined steps. When bound to hormone, these receptors undergo a conformational change, dissociation from cellular chaperones, receptor dimerization, phosphorylation, interaction with coactivators and recruitment of chromatin modifying enzyme activities such as histone acetyl transferase activity (HAT), methyl transferase activity, ATPase activity, ubiquitin-conjugation activity and ubiquitin-protein ligase activity, DNA-binding at an enhancer element of the target gene, and subsequent recruitment of basal transcription factors to form a stable preinitiation complex [37-40]. These events are followed by up- or down-regulation of target gene expression. However, these receptors may also be converted into active forms even in the absence of hormones in target cells. The mechanism of hormone-independent activation of ER and PR has not been understood fully yet but it may involve the bypassing of any one of the above mentioned steps of hormone-dependent activation [41, 42].

Coactivators of ER and PR: Nuclear hormone receptor coactivators are molecules that interact with activated receptors and stimulate receptor-mediated transcription of target genes. There are now ~200 published nuclear hormone receptor coactivators that work with ~ 48 nuclear receptors [6, 8, 9, 11]. The most widely studied coactivators include members of the p160 family of coactivators; SRC-1 (steroid receptor coactivator-1) [43], SRC-2 (TIF-2; transcription intermediary factor-2/GRIP-1; glucocorticoid receptor interacting protein-1) [44, 45], SRC-3 (p/CIP; p300/CBP interacting protein/ACTR; activator of thyroid and retinoid acid receptors/AIB-1; amplified in breast cancer-1/RAC-3; retinoid acid receptor coactivator-3/TRAM-1; thyroid receptor activator molecule-1) [46-49], the CBP (CREB-binding protein)/p300 family [50], coactivator-associated arginine methyltransferase (CARM-1) [51, 52], and E6-AP [10]. We have previously reported the cloning and characterization of E6-AP as a novel dual function steroid hormone receptor coactivator. Additionally, we also demonstrated that the E2 ubiquitin-conjugating enzyme, UbcH7, acts as a coactivator of steroid hormone receptors [53]. Initially, it was thought that coactivators act as adaptors and provide a bridge between DNA binding transcription factors and the general transcription machinery. This simple scenario of coactivator action turned out to be much more complex. It has been shown that coactivators can mediate chromatin modifications either through acetylating reactions mediated by histone acetyl transferases or through nucleosome remodeling complexes [6, 8, 11]. Coactivators are predicted to have many activities in addition to the initiation of transcription, such as mRNA transport from the nucleus, mRNA translation, and posttranslational modifications of the synthesized protein. That coactivators possess stratified actions in the entire process of transcription-translation reflects the fact that they do not act alone but rather as part of multiprotein complexes. These multisubunit entities, containing many individual enzymatic activities, represent a complex machine that is able to

concentrate and link diverse enzymes, and the processes that they regulate, together in one place [6, 8, 11]. In this way, the coactivator complex executes the coactivator's final agenda—that is, to see a particular gene expressed as a mature functional protein.

Transcription is a highly dynamic and orderly process involving many subreactions (multiple steps of initiation, elongation, splicing, and termination) [54]. Given that so many nuclear hormone receptor coactivators have been identified, there is certainly no shortage of them to participate in the wide variety of transcription subreactions. But why would a cell possess such a cumbersome transcriptional apparatus? The answer may lie in the fact that mammals are substantially more complex than organisms such as yeast, worm, and the fruit fly, which have far fewer nuclear receptor coactivators. For instance, only a single nuclear receptor coactivator (Taiman/dAIB1) has been identified in fruit flies so far [55].

For transcription to proceed, there need to be histone modifications (such as acetylation and methylation), ATPase-dependent chromatin remodeling, initiation of transcription, elongation, alternative RNA splicing and mRNA processing, and termination. The focus of coactivator enzymatic activities in these processes has centered on the posttranslational modification of histones and chromatin [56, 57]. However, it is becoming clear that nuclear hormone receptors and their coactivators are also subject to posttranslational modification [58, 59]. The posttranslational targeting of nuclear hormone receptors and their coactivators is important because these modifications influence the expression of functionally related groups of genes. *Identification of new coactivators will provide the prime source for the discovery of new molecular events in transcriptional reactions.*

As mentioned above that coactivator proteins form multiprotein complexes to efficiently regulate target gene transcription. Recently, we have identified novel protein named WBP-2 as an E6-AP interacting protein and show that it specifically modulates ER and PR functions. WBP-2 was previously shown to interact with human YAP1 via the WW-domain of YAP1 protein [12]. The WW-domain is characterized by 35-40 semi-conserved amino acids, which are involved in protein-protein interaction. WBP-2 interacts with the WW-domain via a short proline-rich motif (PPXY) with the consensus sequence of four consecutive prolines followed by tyrosine [16, 31, 35]. It has been speculated that WBP-2 plays role in transcription, but its exact function in steroid hormone receptor-dependent transcription has not been defined yet. Additionally, it has been suggested that YAP1 may also regulate transcription by acting as a coactivator for several transcription factors but what role YAP1 plays in steroid hormone receptor-mediated gene transcription remains unknown. *The main goal of this proposal is to decipher the mechanism by which WBP-2 and YAP1 regulates ER and PR-mediated transcription.*

WBP-2 as a Coactivator: WBP-2 is a 30 kDa protein that contains three proline-rich motifs known as PPXY motifs. WBP-2 was originally identified as a putative ligand that binds to the WW-domain of YAP1 with relatively high affinity and specificity. The PPXY motifs of WBP-2 are distinct from the PXXP ligand consensus sequence of Src homology domain 3 (SH3) domains [16, 31, 35]. The exact function of the WBP-2 and its PPXY motifs remain unknown. Recently, we cloned WBP-2 as E6-AP interacting protein. Despite the fact that E6-AP does not contain a WW-domain, our data clearly demonstrate that WBP-2 can also interact with proteins that do not contain a WW-domain suggesting a new type of protein-protein interaction between PPXY motif containing protein and *HECT* (homologous to the E6-AP carboxy terminus) domain con-

taining proteins [60, 61]. Additionally, we also found that WBP-2 specifically directly interacts with ER and PR. Our observation that WBP-2 can interact with proteins that do not contain WW-domain is also consistent with a previously published study that suggests WBP-2 interacts with the transcription factor Pax8 [14]. Previously, it has been suggested that WBP-2 act as a transcriptional adaptor for Pax8 [14]. We have demonstrated a role for WBP-2 in steroid hormone receptor functions. We have shown that WBP-2 specifically enhances the hormone-dependent transcriptional activity of ER and PR suggesting that WBP-2 can act as transcriptional coactivator of selected steroid hormone receptors. Furthermore, we have also demonstrated that the most carboxy terminal PPXY motif (cPPXY) of WBP-2 is required for its coactivation function, suggesting a role for the PPXY motif in transcription. Together, our results indicate the role of WBP-2 as a coactivator in modulating ER and PR functions [12]. Previously, it has been postulated that the PPXY motif plays an important role in transcriptional activation since the PPXY motif is present in the transcriptional activation domains of a wide range of transcription factors including c-Jun [62], AP-2 [63], NF-E2 [22], C/EBP α [64] and PEBP2/CBF [30]. Our data indicate a role for the PPXY motif and WBP-2 in ER and PR transactivation, but the exact mechanism by which WBP-2 acts as a coactivator and the role of the PPXY motif in ER and PR-dependent gene activation is not fully understood. Furthermore, *the actual target proteins of the PPXY motif that confer transcription stimulation activity have not yet been identified*. Therefore, there is a clear need to understand the role of WBP-2 and its PPXY motif in ER and PR-mediated gene activation and transcription.

WW-Domain Containing Proteins as Modulators of ER and PR Function: Specific protein-protein interactions and multiprotein complexes are important for a multitude of cellular processes including gene transcription. As mentioned above that WBP-2 binds to proteins that contain WW-domain. WW-domains are small protein modules composed of 38-40 amino acids and fold as a stable, triple stranded beta-sheet. The name refers to two signature tryptophan (W) residues that are spaced 20-22 amino acids apart and are present in most of the WW-domains. In some instances, however, the first or the second conserved tryptophan is substituted by other aromatic residues. WW-domains bind to their ligands via the proline rich peptide motifs. WW-domains can be grouped into four classes according to their ligand binding preference. Class I includes WW-domains that bind to the PPXY core sequence which is present in WBP-2. The example of Group I WW-domain containing protein is YAP1. Class II WW-domains prefer ligands containing a stretch of prolines interrupted by a leucine. Class III includes WW-domains interacting with proline-rich sequence that contains arginines or lysines. WW-domains binding phosphoserine or phosphothreonine followed by a proline residue are grouped in class IV. WW-domains are found in both cytoplasmic and nuclear proteins, WW-domains containing proteins are involved in a wide variety of cellular processes, including ubiquitination, nuclear signaling, cell cycle control, transcriptional regulation and the recruitment of signaling proteins [28, 31, 35]. Since, WW-domains bind to the PPXY motif and WBP-2 has been shown to contain PPXY motifs, an important step toward characterizing coactivation function of WBP-2 is to identify to which particular WW-domains it bind, and hence determine the mechanism by which it act as a coactivator and also find out with which signaling pathway it is involved. In order to identify the WW-domains that could interact with the WBP-2 protein, we utilized Panomics' TranSignal WW-domain Array (Panomics Inc., CA, USA). This array contains 67 different human WW-domains from 42 different proteins. Our screening data suggest that WBP-2 interacts with a

wide variety of WW-domain containing proteins (please see preliminary data section) including YAP1, a transcriptional coactivator and WWOX1, a tumor suppressor.

YAP1 was originally identified as a protein binding to the SH3 domain of the Yes proto-oncogene product that belongs to the *src* family of protein-tyrosine kinases. YAP1 is a 65 kDa protein with a well characterized WW-domain [13]. Recently, a splicing variant that encodes human YAP with two WW-domains has been identified. YAP1 with a single WW-domain, known previously as human YAP, is renamed as YAP1 and the new YAP with two WW-domains is named as YAP2. The difference in the amino acid sequence between YAP1 and YAP2 is an insertion of the additional WW-domain in YAP2. YAP1 and 2 contain activation domain which is similar to VP-16 activation domain. Roles of YAP1 and 2 in transcription are not well defined [21]. Recently, YAP1 and YAP2 have been shown to be transcriptional coactivators for several genes but their transcriptional coactivation functions are dependent on PPXY motif containing proteins as in the case of ErbB4 (PPXY motif containing protein) signaling [21]. YAP1 and YAP2 interact with ErbB4 and are recruited to its specific target gene promoter. It has been suggested that YAP2 is a stronger coactivator of transcription than YAP1. Furthermore, phosphorylation of YAP1 and YAP2 by specific kinases regulates their cellular distribution and transcriptional coactivation functions. It is shown that Akt-dependent phosphorylation of YAP1 and YAP2 at Serine 127 induces the interaction of YAP1 with 14-3-3 and promote YAP1 and YAP2 localization to the cytoplasm resulting in loss from the nucleus where they functions as coactivators of transcription [65]. Our preliminary data also indicate a role for YAP1 in steroid hormone receptor-mediated transactivation function. We show that YAP1 expression itself has no significant effect on ER and PR-mediated transcription [12], however, when YAP1 was coexpressed with WBP-2, it could selectively modulate ER and PR-dependent gene transcription. Furthermore, mutation (W199F) in the WW-domain of YAP1 abolishes its coactivation functions [21]. The coactivation function of YAP1 is strictly dependent on the PPXY motif of WBP-2. *The mode of action of WBP-2:YAP1 complex in ER and PR-mediated transactivation is not yet understood.* In this proposal, *we intend to dissect the molecular mechanism and significance of this interaction with relation to ER and PR function.*

As mentioned above, in addition to YAP1, we have also identified another WW-domain containing protein, WWOX1 (a tumor suppressor) as a WBP-2 interacting protein in our WW-domain array. WWOX1 was originally cloned as a putative tumor suppressor gene that spans one of the most common active fragile sites in the human genome, FRA16D. WWOX1 is located on chromosome 16q23.3 and exhibits genomic alterations in several cancer types and a recent study showed a possible involvement of methylation in the regulation of WWOX1 expression. WWOX1 expression level is high in normal tissues; its expression is highest in hormonally regulated normal tissues such as testis, prostate and ovary suggesting that WWOX1 may play an important role in hormone regulated cancers [32, 33, 66-71]. This suggestion is supported by the recent publications which show that WWOX1 expression is frequently altered in tumor tissues and introduction of WWOX1 into WWOX1-negative breast and prostate tumor cells resulted in tumor suppression and apoptosis both *in vitro* and *in vivo* [72]. The WWOX1 contains two WW-domains. The tandem WW-domains of WWOX1 play an important role in WWOX1 function. Both WW-domains of WWOX1 contain a central core of two consecutive aromatic amino acids and therefore belong to the class I specificity of domains, which recognize ligands with the PPXY consensus motif. Recent mapping of the WW-domain in the human proteome identified a

repertoire of PPXY-containing ligands that bind to individual domains of WWOX1. The first WW-domain of WWOX1 bound 18 and the second WW-domain bound 16 ligands, all with PPXY consensus. The mapping data clearly documented that although the second WW-domain of WWOX1 contains a tyrosine in the place of the second conserved tryptophan, the signature residue directly involved in ligand binding, the specificity of the "WY domain" toward PPXY core motif was not changed [33]. Previously, it has been shown that WWOX1 interacts with PPXY motif containing proteins, p73 and AP-2 and suppresses their transcriptional activities [36, 66, 73, 74]. Most recently, it has been shown that WWOX1 interacts with another PPXY motif containing protein, ErbB-4 and compete with YAP1 for ErbB-4 binding and suppress the coactivation functions of YAP1 [36]. Since, WWOX1 and YAP1 have the similar tandem WW-domains and both interact with common protein, WBP-2 but have different effects on transactivation, in this proposal, *we will examine whether WWOX1 and YAP1 expression have opposite effects on ER and PR-dependent gene transcription.*

PPXY Motif and WW-Domain; Possible Function in Gene Regulation: Protein-protein interaction modules are important for the proper signal transduction process in any cell. There are numerous such modules that mediate various processes from cell cycle progression to arrest and eventually apoptosis. One such module, the PPXY motif and WW-domain (PPXY-WW module) has gained prominence in the last decade. WW-domains mediate their interactions with proteins that contain a short PPXY motif. PPXY motif containing proteins essentially act as ligands for WW-domain containing proteins. The K_d of interaction for PPXY motif and WW-domain complex formation is in the high nM to low μ M values. Phosphorylation of the terminal tyrosine in the PPXY motif and phosphorylation of WW-domains containing proteins by specific kinases abolishes their interaction, suggesting that this modification represent a negative regulation mechanism for PPXY-WW module. Although the precise physiological roles of the PPXY motif and WW-domain remain undetermined, their presence in diverse proteins involved in signaling, regulatory, transcription and cytoskeletal functions, as well as their rapidly emerging role in signaling mechanisms that underlie several human diseases, clearly underscores their importance. Protein-protein interactions involving WW-domains and PPXY motif have been implicated in many diseases, including muscular dystrophy, Liddle's syndrome, Alzheimer's, Huntington disease and Cancers [16, 31, 35, 75].

Many new roles of the PPXY motif and WW-domain in the nucleus and transcription are just emerging. It is interesting to note that the PPXY motif is found in the transcriptional activation domains of many transcription factors and mutations in the PPXY motif either reduce or abolish their transcriptional activities. This observation suggests that the PPXY motif plays a role in mediating transcription stimulation by interacting with WW-domain containing proteins which act as transcriptional coactivator. Since, the interaction between WW-domain and PPXY motif is highly specific, it suggests that PPXY-WW complex is a more specialized coactivator complex of a subset of transcription factors [16, 21, 31, 35, 75]. This suggestion was supported by our observation which shows that PPXY motif containing protein, WBP-2 and WW-domain containing protein, YAP1 specifically coactivate ER and PR-dependent transcription where as this protein complex has no significant effect on the transactivation functions of other receptors. In contrast to this complex, it has been shown that other transcriptional coactivator complexes such as CBP, SRC-1 and p/CAF (p300/CBP associated factor) coactivates the transcriptional activities of a variety of receptors without exerting any specificity [6]. In addition to transcription stimula-

tion, the PPXY motif has also been shown to suppress transcription after interacting with certain WW-domain containing proteins. These observations suggest that *PPXY-WW module is involved in gene transcription but the actual mechanism by which PPXY-WW complex modulates transcription remains unknown.*

Significance: Coactivators play important roles in diverse pathological processes, such as cancer, inherited genetic diseases, metabolic disorders, and inflammation [6]. There is little doubt that we have much to learn about the biologically diverse roles of coactivators and that we have only scratched the surface of this expansive coactivator cosmos. Therefore, characterizing the mechanism of action of coactivator proteins will provide the prime source for the discovery of new molecular events in transcriptional reactions *and their role in cellular, physiological and pathological processes.* Collectively, our preliminary data suggests that WBP-2 (PPXY motif), YAP1 (WW-domain) and WWOX1 (WW-domain) are key regulators of ER and PR transactivation function but the precise roles of the PPXY motif and WW-domain containing proteins in steroid hormone receptor signaling [12], cell growth and carcinogenesis remain undetermined. Protein-protein interactions involving WW-domains and PPXY motif have been implicated in many diseases, including hormone regulated cancers. But the exact mechanism by which they regulate transcription, cell function and growth are largely unknown. Novel concepts and approaches to elucidate the molecular mechanisms by which PPXY motif and WW-domain containing proteins regulates hormone-dependent gene transcription are proposed here. Thus, accomplishing the specific aims outlined in this proposal will address the novel roles of the PPXY-WW complex in ER and PR function and will provide new and timely insights into the mechanism of action of PPXY motif and WW-domains in ER and PR signaling and cellular pathways that are regulated by these modulatory proteins.

Body

In the original proposal, we proposed to dissect the role of WBP-2 in ER and PR signaling and breast tumorigenesis.

- A. Role of PY motif containing protein, WBP-2 in ER and PR signaling**
- B. Role of WW-domain containing proteins, YAP and WWOX1 in SHR function**
- C. Expression analysis of endogenous WBP-2 protein**

A. Role of PY motif containing protein, WBP-2 in ER and PR signaling

This aim has been successfully completed and the findings were reported in the previous annual report. The major outcomes of this aim were also published

B. Role of WW-domain containing proteins, YAP and WWOX1 in SHR function

The third specific aim is intricately related to the second aim so we are pursuing both the aims simultaneously. As a consequence of the modification of aim two the scope of this aim is extended to include the newly identified WW-domain containing proteins that may be involved in the mechanism of action of WBP-2 protein.

I. Identification of WBP-2 Binding Proteins:

Specific protein-protein interactions and multiprotein complexes are important for a multitude of cellular processes including gene transcription. As mentioned above WBP-2 via its PPXY motifs binds to proteins that contain WW-domain. WW-domains are found in both cytoplasmic and nuclear proteins, WW-domains containing proteins are involved in ubiquitination, nuclear signaling, cell cycle control, transcriptional regulation and the recruitment of signaling proteins. Since, WW-domains bind to the PPXY motif and WBP-2 has been shown to contain PPXY motifs, an important step toward characterizing coactivation function of WBP-2 is to identify to which particular WW-domain containing protein it binds, and hence determine the mechanism by which it acts as a coactivator. In order to identify the possible WW-domain containing proteins that could interact with the WBP-2 protein, we utilized Panomics' TranSignal WW-domain Array (Panomics Inc., CA, USA). This array contains 67 different human WW-domains from 42 different proteins. The arrays are made using the recombinant conserved binding sites of individual WW-domains fused with GST. Proteins are affinity purified and immobilized onto a membrane. Each WW-domain is spotted in duplicate. In order to identify the WW-domain(s) that interact with WBP-2, the full-length wild-type WBP-2 protein containing flag tag was expressed in bacteria. Afterward, WBP-2 protein was purified on flag beads and incubated with TranSignal WW-domain Array membranes. The protein-protein interaction was visualized by using HRP-based chemiluminescence detection. The resulting interacting proteins have been tabulated in table-1. Our screening data suggest that WBP-2 interacts with a wide variety of WW-domain containing proteins including YAP1, a transcriptional coactivator and WWOX1, a tumor suppressor (Fig. 1).

Table 1 List of WBP-2 interacting proteins

Gene Symbol	WW-domain(s)	Interaction	Protein name	Function
SMURF1	D1	-	WW-domain containing protein 1	E3 ubiquitin ligase
	D2	+		E3 ubiquitin ligase
SMURF2	D1	-		E3 ubiquitin ligase
	D2	+		E3 ubiquitin ligase
WWP1	D1	++		Nedd-4-like ubiquitin protein ligase
	D2	++		Nedd-4-like ubiquitin protein ligase
	D3	++		Nedd-4-like ubiquitin protein ligase
	D4	++		Nedd-4-like ubiquitin protein ligase
NEDD4	D1	++	Neuronal precursor	Ubiquitin protein

	D2	++	cell-expressed developmentally downregulated 4	ligase
	D3	++		
	D4	++		
Caveolin-3		+		Membrane protein
PABPN1		+	Poly(A)-binding protein	
GAS7		+	Growth-arrest specific 7 isoform b	
YAP		++	yes-associated protein	Transcriptional Coactivator
BAIAP1	D1	-	Membrane associated guanylate kinase-1	
	D2	+		
ITCH	D1	+	Atropin-1 interacting protein 4	E3 ubiquitin ligase
	D2	+		
	D3	+		
	D4	+		
TAZ		++	Transcriptional coactivator with PDZ domain	Transcriptional coactivator
WWOX1	D1	++	WW-domain containing oxidoreductase, isoform 1	Tumor suppressor
	D2	-/+		
APBB3		+	FE65-like protein	

II. PY3 motif of WBP-2 is essential for its interaction with YAP:

Initially, Sudol et al, identified WBP-2 as a binding partner for YAP using a functional screen. Our WW-domain array also confirms this observation that WBP-2 interacts with YAP. Next we wanted to know if this interaction is mediated by the PY motif of WBP-2 and the WW-domain of YAP, for this purpose we used both wild-type and coactivation function-dead PY3 motif mutant WBP-2 in GST pull-down assays. Figure 2, demonstrates that while the wild-type WBP-2 was able to interact with YAP, mutation of the PY3 motif of WBP-2 completely abrogated this interaction. Together with our previous observations we concluded that the PY3 motif of WBP-2, though not essential for its interaction with the receptors was indispensable for its interaction with the WW-domain containing protein YAP.

III. The WW-Domain Containing Protein, YAP1, Modulates Progesterone Receptor Transcriptional Activity via the WBP-2 Protein:

Since, WBP-2 binds to the WW-domain containing protein, YAP1 and YAP1 has been shown to be a transcriptional coactivator. Thus, we wanted to know whether YAP1 modulates steroid receptor-dependent target gene expression. To determine the role of YAP1 in steroid hormone receptor transactivation, HeLa cells were co-transfected with mammalian expression plasmids for the PR and ER receptors along with reporter plasmids containing their cognate hormone response element, with or without an expression vector for YAP1. *YAP1 alone did not affect PR-mediated transactivation either in the absence or presence of hormone.* In contrast, *when YAP1 was coexpressed with WBP-2 the hormone-dependent transcriptional activity of PR was significantly enhanced (~24-fold) (Fig. 3).* Similarly, YAP1 alone did not activate the ER-

mediated transactivation but when co-expressed with WBP-2 enhanced ER-mediated transactivation (data not shown). This activity was higher than the observed coactivation with WBP-2 alone (Fig. 3). ***These data suggest that YAP1 can modulate the ligand-dependent transcriptional activity of PR and ER but only via WBP-2.*** The PPXY motifs of WBP-2 have been shown to interact with the WW-domain of YAP1. Since our data revealed that the cPPXY of WBP-2 was required for its coactivation function, we next asked whether the cPPXY of WBP-2 is required for YAP1 to function as a coactivator for ER and PR. When coexpressed together, wild-type WBP-2 and wild-type YAP1 greatly enhanced the transactivation function of PR (Fig. 3). In contrast, the cPPXY mutant WBP-2 and wild-type YAP1 also failed to enhance the transcriptional activity of PR (Fig. 3). Our data demonstrate that *the cPPXY motif of the WBP-2 protein is required for YAP1 to function as a transcriptional secondary coactivator.* This data is consistent with previously published reports that YAP1 stimulates gene transcription by binding to the PPXY motif of ErbB4 protein. In order to determine whether the WW-domain of YAP1 is required for its transcriptional coactivator function, a WW-domain mutant (W199F) was utilized. This YAP1 mutant has been shown to be inactive in its ability to bind to PPXY motif. When coexpressed together, wild-type WBP-2 and WW-domain mutant YAP1, the WW-domain mutant YAP1 fail to act as a coactivator (data not shown). These data suggest that *WW-domain of YAP1 is required for it to function as a transcriptional coactivator.* In summary, our preliminary data *substantiate the role of WBP-2 (contains PPXY motif) and YAP1 (contains WW-domain) in female steroid hormone receptor function.* Based on our data, we postulate that *the cPPXY motif of WBP-2 binds to the WW-domain of YAP1 and recruits YAP1 to the target gene promoter by interacting with receptor. When the receptor-WBP-2-YAP1 complex is recruited to hormone responsive promoters, it acts at one of the many substeps required to modulate the transactivation functions of ER and PR- responsive target genes.*

IV. YAP Enhances Endogenous Estrogen Receptor Target Gene Activation via the WBP-2 Protein

To further substantiate that YAP and WBP-2 synergistically enhance ER and PR transactivation functions as observed by reporter gene assays, we performed real-time PCR (QPCR) analysis of ER α target gene pS2 in MCF-7 cells. MCF-7 cells were co-transfected with either control plasmid or expression plasmids of wild-type WBP-2, PY3 motif mutant WBP-2, and wild-type YAP alone or in combination. Cells were treated with either vehicle or estradiol (E₂) and expression of estrogen-regulated gene, pS2 was measured by QPCR. As control, we also examined the mRNA levels of ER, WBP-2 and YAP. There was no change observed in ER mRNA levels (data not shown) whereas, the mRNA levels of exogenously expressed WBP-2 (wild-type and PY3 mutant) and wild-type YAP were significantly increased in cells that were transfected with their expression plasmid compared to that of cells transfected with control plasmids (Figure 4A). As shown in Figure 4, WBP-2 enhanced the relative mRNA levels of ER α target gene, pS2 in the presence of estradiol but YAP alone showed no enhancement. Whereas, when YAP was coexpressed with wild-type WBP-2, the hormone-dependent transcriptional activity of ER α was synergistically enhanced, more than that observed with WBP-2 alone (Figure 4B). Furthermore, this synergistic enhancement of pS2 relative mRNA levels was not observed when YAP was coexpressed with the PY3 motif mutant WBP-2. Taken together, both our reporter gene assays and endogenous target gene assays demonstrate that YAP can function as a secondary transcriptional coactivator (co-coactivator) of ER α in a WBP-2 dependent manner (Figure 4B).

V. Estrogen induces association of ER α with WBP-2 and YAP at ER α responsive gene promoter

Since WBP-2 and YAP interact with each other and synergistically enhance ER α transactivation functions, we next investigated the recruitment of WBP-2 and YAP to ER α responsive gene promoters by quantitative chromatin immunoprecipitation (ChIP) assays. YAP has been shown to be a potential transcriptional coactivator of various other transcription factors including p73 and RUNX2. We demonstrated previously the hormone dependent recruitment of WBP-2 to the ER α responsive pS2 promoter by classical ChIP assays in MCF-7 cells. To further investigate the association of ER α with WBP-2 and YAP at ER α responsive pS2 promoter we performed quantitative re-ChIP assays in MCF-7 cells. Cross-linked and sheared DNA-protein complexes from MCF-7 cells were immunoprecipitated with either non-specific purified IgG (Mock) or ER- α specific antibody. After immunoprecipitation the cross-linked immunocomplexes from each of the primary ChIPs were eluted and subjected to another round of immunoprecipitation with antibodies specific for WBP-2 or YAP. The immunocomplexes after re-ChIP were eluted, reverse cross-linked and the associated genomic DNA fragments were analyzed by QPCR with pS2 promoter specific primers.

We observed that in ER/WBP-2 ChIP assays where ER α antibody was used in the first ChIP and WBP-2 specific antibody was used in the re-ChIP, there was a significant enrichment in E₂-induced association of WBP-2 with ER α in comparison with the mock ChIP (IgG/WBP-2) as given by the increased association of pS2 promoter locus in QPCR assays (Figure 5). Similar results were observed with ER/YAP ChIP when compared to its mock (IgG/YAP) ChIP (Figure 5). These observations suggest that WBP-2 and YAP (as separate entities) are recruited to and are associated with ER α at the pS2 promoter locus in an estrogen dependent manner.

VI. Estrogen-induced recruitment of WBP-2 and YAP onto ER α responsive pS2 promoter is mutually interdependent

Our initial observations showed that YAP1 acts as a secondary coactivator of ER α only when co-expressed with WBP-2. We also show that estrogen enhances the recruitment of YAP1 to ER α responsive pS2 promoter. To test further if the recruitment of YAP1 is also dependent on WBP-2, we performed quantitative re-ChIP assays as described above in MCF-7 cells that were treated with siRNAs against YAP1 and WBP-2. As control in these assays, MCF-7 cells were treated with a non-specific scrambled siRNA.

In cells treated with non-specific scrambled siRNA, estrogen-induced association of ER α with WBP-2 (ER α /WBP-2) and YAP1 (ER α /YAP1) were consistent with our earlier observations when compared to their respective mock ChIP assays. Whereas in cells treated with siRNA against WBP-2, the E₂-induced enhancement of association of YAP1 with ER α was completely lost, suggesting that the recruitment and association of YAP1 with ER α at the pS2 promoter locus is dependent on the normal endogenous expression levels of WBP-2. Interestingly, the converse relationship also holds true, where the association of WBP-2 with ER α and its recruitment to the pS2 promoter locus was abolished in cells that were treated with siRNA against YAP1 (Figure 6). These observations suggest an intriguing possibility where the recruitment of WBP-2 and YAP1 to the pS2 promoter locus and their association with ER α may be mutually interdependent of their normal physiological expression levels in MCF-7 cells.

VII. WWOX1 suppresses the transcriptional coactivation functions of WBP-2 and YAP1

As mentioned above, we have also identified another WW-domain containing protein, WWOX1 as a WBP-2 interacting protein in our WW-domain array. WWOX1 interacts with WBP-2 via its first WW-domain. WWOX1 was originally cloned as a putative tumor suppressor gene and it has been suggested that WWOX1 may play an important role in hormone regulated cancers [32, 33, 70, 78, 79]. Previously, it has been shown that WWOX1 interacts with PPXY motif containing proteins, p73 and AP-2 and suppresses their transcriptional activities. Most recently, it has been shown that WWOX1 interacts with another PPXY motif containing protein, ErbB-4 and compete with YAP1 for ErbB-4 binding and suppress the coactivation functions of YAP1 [74]. Since, WWOX1 and YAP1 have the similar tandem WW-domains and both interact with common protein, WBP-2; we ask whether WWOX1 and YAP1 expression have opposite effects on ER and PR-dependent gene transcription. To determine the role of WWOX1 in steroid hormone receptor transactivation, MCF-7 cells were co-transfected with ER-responsive reporter plasmid along with expression vectors for either WBP-2, YAP1, and WWOX1, WBP-2 and WWOX1 or WBP-2, YAP1 and WWOX1. As shown before, WBP-2 coactivates the transactivation functions of ER. Furthermore, YAP1 and WWOX1 alone had no significant effect on ER function. But when YAP1 was coexpressed with WBP-2 the hormone-dependent transcriptional activity of ER was synergistically enhanced (Fig. 7). In contrast, expression of WWOX1 significantly reduced the coactivation functions of WBP-2. Similarly, WWOX1 also significantly suppressed WBP-2-YAP1-mediated transcriptional activities of ER and PR in a dose-dependent manner (Fig. 7). To determine whether the WW-domain of WWOX1 is required for its transcriptional suppression function, a WW-domain mutant was utilized in which the tryptophan 33 within WW-domain 1 was mutated to arginine. This WWOX1 mutant has been shown to be inactive in its ability to bind to PPXY motif. When coexpressed along with wild-type WBP-2 and wild-type YAP1, this mutant had no significant effect on WBP-2 and YAP1's coactivation functions (Data not shown). Identical results were obtained with PR (data not shown). These data indicate that *the coactivation functions of WBP-2 and YAP1 are suppressed by WWOX1, suggesting that WWOX1 may regulate the transactivation functions of ER and PR by antagonizing the functions of WBP-2 and YAP1.*

C. Expression analysis of endogenous WBP-2 protein

In light of the recent modifications to the original proposal we have accommodated and updated the first aim of the proposal. In the original proposal we intended to analyze only the expression profile of WBP-2 in various cancer cell lines and human breast tissue arrays. Our current understanding is that YAP1 and WWOX1 may play vital roles in the regulations and function of WBP-2, furthermore YAP1 has been shown to be amplified in various breast cancers and intriguingly, WWOX1 has been shown to be a potent tumor suppressor [74]. Given these interesting facts we propose to analyze the expression of WBP-2, YAP1 and WWOX1 in correlation with the ER and PR expression status of various breast cancer cell lines as well as many breast tumor-arrays which may shed some light on the complexity of WBP-2 function and the possible roles of YAP1 and WWOX1 in breast cancers.

Key Research Accomplishments

- Establishing that WBP-2 is a coactivator of ER and PR
- The PY motif of WBP-2 is essential for its coactivation function
- Identification of YAP1 and WWOX1 as WBP-2 interacting proteins
- YAP1 acts as transcriptional secondary coactivator of ER and PR that is strictly dependent of WBP-2
- WW-domain of YAP and the PY motif of WBP-2 are essential for their coactivation activities
- WBP-2 and YAP are recruited to ER-responsive promoter
- Estrogen dependent recruitment and association of WBP-2 with ER is dependent on YAP, and vice versa.
- WWOX1 may act as a transcriptional repressor of WBP-2

Reportable Outcomes

1. The first part of the this work has been published “Dhananjayan, S.C., et al., *WW Domain Binding Protein-2, an E6-Associated Protein Interacting Protein, Acts as a Coactivator of Estrogen and Progesterone Receptors*. Mol Endocrinol, 2006. **20**(10): p. 2343-2354” (Please refer to previous Annual Report).
2. The second part of this report has been accepted to be presented at the Annual Endocrine Society Meeting, ENDO 2007 (June 2-5th), in Toronto, Canada (Please refer to previous Annual Report).

Conclusions

Our data demonstrates that WBP-2 is recruited onto the hormone responsive promoters in the presence of hormone and it specifically enhances the transactivation functions of PR and ER. Our data also demonstrates that WBP-2 contains an intrinsic activation domain and the cPPXY of WBP-2 is essential for its coactivation and intrinsic activation functions. Our preliminary data also demonstrates that the WBP-2 binding protein, YAP1 enhances PR and ER transactivation but YAP1's coactivation function is absolutely dependent on WBP-2. Furthermore, cPPXY motif of WBP-2 and WW-domain of YAP1 is required for YAP1 to work as a transcriptional coactivator. Additionally, our data also indicate that the coactivation functions of WBP-2 and YAP1 are suppressed by WWOX1, suggesting that WWOX1 may regulates the transactivation functions of ER and PR by antagonizing the functions of WBP-2 and YAP1 (Fig. 8). Taken together our data established the role of WBP-2 and YAP1 as coactivators and WWOX1 as a repressor for ER and PR transactivation pathways.

References

1. Edwards, D.P., *The role of coactivators and corepressors in the biology and mechanism of action of steroid hormone receptors*. J Mammary Gland Biol Neoplasia, 2000. **5**(3): p. 307-24.
2. Eelen, G., et al., *Vitamin D analogs and coactivators*. Anticancer Res, 2006. **26**(4A): p. 2717-21.
3. Gao, X., B.W. Loggie, and Z. Nawaz, *The roles of sex steroid receptor coregulators in cancer*. Mol Cancer, 2002. **1**: p. 7.
4. Gao, X. and Z. Nawaz, *Progesterone receptors - animal models and cell signaling in breast cancer: Role of steroid receptor coactivators and corepressors of progesterone receptors in breast cancer*. Breast Cancer Res, 2002. **4**(5): p. 182-6.
5. Li, X., D.M. Lonard, and B.W. O'Malley, *A contemporary understanding of progesterone receptor function*. Mech Ageing Dev, 2004. **125**(10-11): p. 669-78.
6. Lonard, D.M. and B.W. O'Malley, *The expanding cosmos of nuclear receptor coactivators*. Cell, 2006. **125**(3): p. 411-4.
7. Mahajan, M.A. and H.H. Samuels, *Nuclear hormone receptor coregulator: role in hormone action, metabolism, growth, and development*. Endocr Rev, 2005. **26**(4): p. 583-97.
8. Nawaz, Z. and B.W. O'Malley, *Urban renewal in the nucleus: is protein turnover by proteasomes absolutely required for nuclear receptor-regulated transcription?* Mol Endocrinol, 2004. **18**(3): p. 493-9.
9. Nishihara, E., B.W. O'Malley, and J. Xu, *Nuclear receptor coregulators are new players in nervous system development and function*. Mol Neurobiol, 2004. **30**(3): p. 307-25.
10. Nawaz, Z., et al., *The Angelman syndrome-associated protein, E6-AP, is a coactivator for the nuclear hormone receptor superfamily*. Mol Cell Biol, 1999. **19**(2): p. 1182-9.
11. McKenna, N.J., et al., *Nuclear receptor coactivators: multiple enzymes, multiple complexes, multiple functions*. J Steroid Biochem Mol Biol, 1999. **69**(1-6): p. 3-12.
12. Dhananjayan, S.C., et al., *WW Domain Binding Protein-2, an E6-Associated Protein Interacting Protein, Acts as a Coactivator of Estrogen and Progesterone Receptors*. Mol Endocrinol, 2006. **20**(10): p. 2343-2354.
13. Chen, H.I. and M. Sudol, *The WW domain of Yes-associated protein binds a proline-rich ligand that differs from the consensus established for Src homology 3-binding modules*. Proc Natl Acad Sci U S A, 1995. **92**(17): p. 7819-23.
14. Nitsch, R., et al., *WBP-2, a WW domain binding protein, interacts with the thyroid-specific transcription factor Pax8*. Biochem J, 2004. **377**(Pt 3): p. 553-60.
15. Pirozzi, G., et al., *Identification of novel human WW domain-containing proteins by cloning of ligand targets*. J Biol Chem, 1997. **272**(23): p. 14611-6.
16. Sudol, M., et al., *Characterization of a novel protein-binding module--the WW domain*. FEBS Lett, 1995. **369**(1): p. 67-71.
17. Guillebault, D., et al., *Role of nuclear WW domains and proline-rich proteins in dinoflagellate transcription*. Protist, 2001. **152**(2): p. 127-38.
18. Helton, E.S., J. Zhu, and X. Chen, *The unique NH2-terminally deleted (DeltaN) residues, the PXXP motif, and the PPXY motif are required for the transcriptional activity of the DeltaN variant of p63*. J Biol Chem, 2006. **281**(5): p. 2533-42.

19. Ingham, R.J., et al., *WW domains provide a platform for the assembly of multiprotein networks*. Mol Cell Biol, 2005. **25**(16): p. 7092-106.
20. Irie, T., et al., *Budding of PPxY-containing rhabdoviruses is not dependent on host proteins TGS101 and VPS4A*. J Virol, 2004. **78**(6): p. 2657-65.
21. Komuro, A., et al., *WW domain-containing protein YAP associates with ErbB-4 and acts as a co-transcriptional activator for the carboxyl-terminal fragment of ErbB-4 that translocates to the nucleus*. J Biol Chem, 2003. **278**(35): p. 33334-41.
22. Mosser, E.A., et al., *Physical and functional interactions between the transactivation domain of the hematopoietic transcription factor NF-E2 and WW domains*. Biochemistry, 1998. **37**(39): p. 13686-95.
23. Strano, S., et al., *Physical interaction with Yes-associated protein enhances p73 transcriptional activity*. J Biol Chem, 2001. **276**(18): p. 15164-73.
24. Bedford, M.T., D.C. Chan, and P. Leder, *FBP WW domains and the Abl SH3 domain bind to a specific class of proline-rich ligands*. Embo J, 1997. **16**(9): p. 2376-83.
25. Bedford, M.T., et al., *A novel pro-Arg motif recognized by WW domains*. J Biol Chem, 2000. **275**(14): p. 10359-69.
26. Kanai, F., et al., *TAZ: a novel transcriptional co-activator regulated by interactions with 14-3-3 and PDZ domain proteins*. Embo J, 2000. **19**(24): p. 6778-91.
27. Macias, M.J., et al., *Structure of the WW domain of a kinase-associated protein complexed with a proline-rich peptide*. Nature, 1996. **382**(6592): p. 646-9.
28. Otte, L., et al., *WW domain sequence activity relationships identified using ligand recognition propensities of 42 WW domains*. Protein Sci, 2003. **12**(3): p. 491-500.
29. Pereboev, A.V., et al., *Epitopes in the interacting regions of beta-dystroglycan (PPxY motif) and dystrophin (WW domain)*. Biochim Biophys Acta, 2001. **1527**(1-2): p. 54-60.
30. Yagi, R., et al., *A WW domain-containing yes-associated protein (YAP) is a novel transcriptional co-activator*. Embo J, 1999. **18**(9): p. 2551-62.
31. Sudol, M., et al., *WW or WoW: the WW domains in a union of bliss*. IUBMB Life, 2005. **57**(12): p. 773-8.
32. Ludes-Meyers, J.H., et al., *WFOX, the common chromosomal fragile site, FRA16D, cancer gene*. Cytogenet Genome Res, 2003. **100**(1-4): p. 101-10.
33. Ludes-Meyers, J.H., et al., *WFOX binds the specific proline-rich ligand PPXY: identification of candidate interacting proteins*. Oncogene, 2004. **23**(29): p. 5049-55.
34. Kato, Y., et al., *Common mechanism of ligand recognition by group II/III WW domains: redefining their functional classification*. J Biol Chem, 2004. **279**(30): p. 31833-41.
35. Sudol, M. and T. Hunter, *NeW wrinkles for an old domain*. Cell, 2000. **103**(7): p. 1001-4.
36. Aqeilan, R.I., et al., *WW domain-containing proteins, WFOX and YAP, compete for interaction with ErbB-4 and modulate its transcriptional function*. Cancer Res, 2005. **65**(15): p. 6764-72.
37. Ozawa, H., *Steroid Hormones, their receptors and neuroendocrine system*. J Nippon Med Sch, 2005. **72**(6): p. 316-25.
38. Shupnik, M.A., *Crosstalk between steroid receptors and the c-Src-receptor tyrosine kinase pathways: implications for cell proliferation*. Oncogene, 2004. **23**(48): p. 7979-89.
39. Walters, M.R. and I. Nemere, *Receptors for steroid hormones: membrane-associated and nuclear forms*. Cell Mol Life Sci, 2004. **61**(18): p. 2309-21.
40. Clarke, R.B., E. Anderson, and A. Howell, *Steroid receptors in human breast cancer*. Trends Endocrinol Metab, 2004. **15**(7): p. 316-23.

41. Norman, A.W., M.T. Mizwicki, and D.P. Norman, *Steroid-hormone rapid actions, membrane receptors and a conformational ensemble model*. Nat Rev Drug Discov, 2004. **3**(1): p. 27-41.
42. Lange, C.A., *Making sense of cross-talk between steroid hormone receptors and intracellular signaling pathways: who will have the last word?* Mol Endocrinol, 2004. **18**(2): p. 269-78.
43. Onate, S.A., et al., *Sequence and characterization of a coactivator for the steroid hormone receptor superfamily*. Science, 1995. **270**(5240): p. 1354-7.
44. Ding, X.F., et al., *Nuclear receptor-binding sites of coactivators glucocorticoid receptor interacting protein 1 (GRIP1) and steroid receptor coactivator 1 (SRC-1): multiple motifs with different binding specificities*. Mol Endocrinol, 1998. **12**(2): p. 302-13.
45. Voegel, J.J., et al., *TIF2, a 160 kDa transcriptional mediator for the ligand-dependent activation function AF-2 of nuclear receptors*. Embo J, 1996. **15**(14): p. 3667-75.
46. Anzick, S.L., et al., *AIB1, a steroid receptor coactivator amplified in breast and ovarian cancer*. Science, 1997. **277**(5328): p. 965-8.
47. Chen, H., et al., *Nuclear receptor coactivator ACTR is a novel histone acetyltransferase and forms a multimeric activation complex with P/CAF and CBP/p300*. Cell, 1997. **90**(3): p. 569-80.
48. Li, H., P.J. Gomes, and J.D. Chen, *RAC3, a steroid/nuclear receptor-associated coactivator that is related to SRC-1 and TIF2*. Proc Natl Acad Sci U S A, 1997. **94**(16): p. 8479-84.
49. Takeshita, A., et al., *TRAM-1, A novel 160-kDa thyroid hormone receptor activator molecule, exhibits distinct properties from steroid receptor coactivator-1*. J Biol Chem, 1997. **272**(44): p. 27629-34.
50. Smith, C.L., et al., *CREB binding protein acts synergistically with steroid receptor coactivator-1 to enhance steroid receptor-dependent transcription*. Proc Natl Acad Sci U S A, 1996. **93**(17): p. 8884-8.
51. Chen, S.L., et al., *The coactivator-associated arginine methyltransferase is necessary for muscle differentiation: CARM1 coactivates myocyte enhancer factor-2*. J Biol Chem, 2002. **277**(6): p. 4324-33.
52. Chen, D., S.M. Huang, and M.R. Stallcup, *Synergistic, p160 coactivator-dependent enhancement of estrogen receptor function by CARM1 and p300*. J Biol Chem, 2000. **275**(52): p. 40810-6.
53. Verma, S., et al., *The ubiquitin-conjugating enzyme UBCH7 acts as a coactivator for steroid hormone receptors*. Mol Cell Biol, 2004. **24**(19): p. 8716-26.
54. Thomas, M.C. and C.M. Chiang, *The general transcription machinery and general cofactors*. Crit Rev Biochem Mol Biol, 2006. **41**(3): p. 105-78.
55. Yoshida, H., et al., *Steroid receptor coactivator-3, a homolog of Taiman that controls cell migration in the Drosophila ovary, regulates migration of human ovarian cancer cells*. Mol Cell Endocrinol, 2005. **245**(1-2): p. 77-85.
56. Sharma, R.P., et al., *Chromatin, DNA methylation and neuron gene regulation--the purpose of the package*. J Psychiatry Neurosci, 2005. **30**(4): p. 257-63.
57. van Leeuwen, F. and B. van Steensel, *Histone modifications: from genome-wide maps to functional insights*. Genome Biol, 2005. **6**(6): p. 113.
58. Li, X., et al., *The SRC-3/AIB1 coactivator is degraded in a ubiquitin- and ATP-independent manner by the REGgamma proteasome*. Cell, 2006. **124**(2): p. 381-92.

59. Yan, F., et al., *Specific ubiquitin-conjugating enzymes promote degradation of specific nuclear receptor coactivators*. Mol Endocrinol, 2003. **17**(7): p. 1315-31.
60. Hatakeyama, S., J.P. Jensen, and A.M. Weissman, *Subcellular localization and ubiquitin-conjugating enzyme (E2) interactions of mammalian HECT family ubiquitin protein ligases*. J Biol Chem, 1997. **272**(24): p. 15085-92.
61. Huibregtse, J.M., et al., *A family of proteins structurally and functionally related to the E6-AP ubiquitin-protein ligase*. Proc Natl Acad Sci U S A, 1995. **92**(7): p. 2563-7.
62. Baichwal, V.R. and R. Tjian, *Control of c-Jun activity by interaction of a cell-specific inhibitor with regulatory domain delta: differences between v- and c-Jun*. Cell, 1990. **63**(4): p. 815-25.
63. Williams, T. and R. Tjian, *Analysis of the DNA-binding and activation properties of the human transcription factor AP-2*. Genes Dev, 1991. **5**(4): p. 670-82.
64. Nerlov, C. and E.B. Ziff, *Three levels of functional interaction determine the activity of CCAAT/enhancer binding protein-alpha on the serum albumin promoter*. Genes Dev, 1994. **8**(3): p. 350-62.
65. Basu, S., et al., *Akt phosphorylates the Yes-associated protein, YAP, to induce interaction with 14-3-3 and attenuation of p73-mediated apoptosis*. Mol Cell, 2003. **11**(1): p. 11-23.
66. Aqeilan, R.I., et al., *Loss of WWOX expression in gastric carcinoma*. Clin Cancer Res, 2004. **10**(9): p. 3053-8.
67. Bednarek, A.K., et al., *WWOX, the FRA16D gene, behaves as a suppressor of tumor growth*. Cancer Res, 2001. **61**(22): p. 8068-73.
68. Bednarek, A.K., et al., *WWOX, a novel WW domain-containing protein mapping to human chromosome 16q23.3-24.1, a region frequently affected in breast cancer*. Cancer Res, 2000. **60**(8): p. 2140-5.
69. Fabbri, M., et al., *WWOX gene restoration prevents lung cancer growth in vitro and in vivo*. Proc Natl Acad Sci U S A, 2005. **102**(43): p. 15611-6.
70. Nunez, M.I., et al., *Frequent loss of WWOX expression in breast cancer: correlation with estrogen receptor status*. Breast Cancer Res Treat, 2005. **89**(2): p. 99-105.
71. Yendamuri, S., et al., *WW domain containing oxidoreductase gene expression is altered in non-small cell lung cancer*. Cancer Res, 2003. **63**(4): p. 878-81.
72. Chang, N.S., et al., *Molecular mechanisms underlying WOX1 activation during apoptotic and stress responses*. Biochem Pharmacol, 2003. **66**(8): p. 1347-54.
73. Aqeilan, R.I., et al., *Physical and functional interactions between the Wwox tumor suppressor protein and the AP-2gamma transcription factor*. Cancer Res, 2004. **64**(22): p. 8256-61.
74. Aqeilan, R.I., et al., *Functional association between Wwox tumor suppressor protein and p73, a p53 homolog*. Proc Natl Acad Sci U S A, 2004. **101**(13): p. 4401-6.
75. Sudol, M., K. Sliwa, and T. Russo, *Functions of WW domains in the nucleus*. FEBS Lett, 2001. **490**(3): p. 190-5.
76. Molkentin, J.D., et al., *MEF2B is a potent transactivator expressed in early myogenic lineages*. Mol Cell Biol, 1996. **16**(7): p. 3814-24.
77. Vesque, C. and P. Charnay, *Mapping functional regions of the segment-specific transcription factor Krox-20*. Nucleic Acids Res, 1992. **20**(10): p. 2485-92.
78. Nunez, M.I., et al., *WWOX protein expression varies among ovarian carcinoma histotypes and correlates with less favorable outcome*. BMC Cancer, 2005. **5**(1): p. 64.

79. Qin, H.R., et al., *A role for the WWOX gene in prostate cancer*. Cancer Res, 2006. **66**(13): p. 6477-81.
80. Zaidi, S.K., et al., *Tyrosine phosphorylation controls Runx2-mediated subnuclear targeting of YAP to repress transcription*. Embo J, 2004. **23**(4): p. 790-9.
81. Dennis, A.P., et al., *Inhibition of the 26S proteasome blocks progesterone receptor-dependent transcription through failed recruitment of RNA polymerase II*. J Steroid Biochem Mol Biol, 2005. **94**(4): p. 337-46.
82. Cosgrove, M.S. and C. Wolberger, *How does the histone code work?* Biochem Cell Biol, 2005. **83**(4): p. 468-76.
83. Kiekhäfer, C.M., et al., *A WW domain-binding motif within the activation domain of the hematopoietic transcription factor NF-E2 is essential for establishment of a tissue-specific histone modification pattern*. J Biol Chem, 2004. **279**(9): p. 7456-61.
84. Mellor, J., *It takes a PHD to read the histone code*. Cell, 2006. **126**(1): p. 22-4.

Appendix 1

Figures

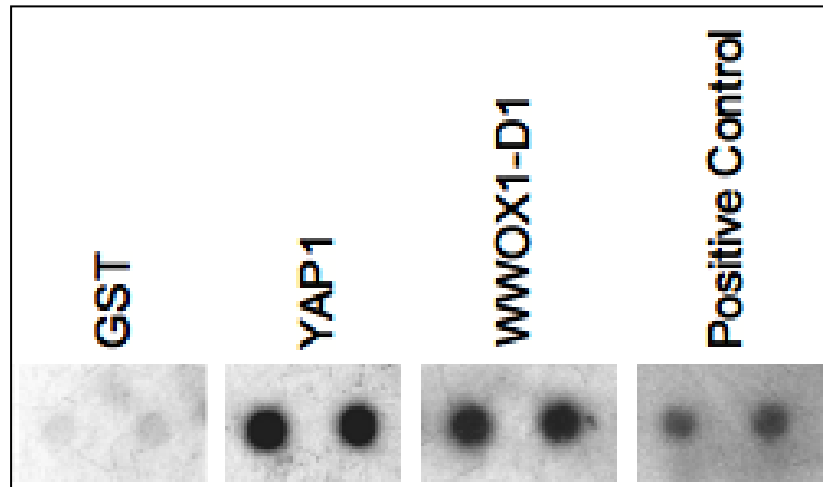


Fig. 1: WBP-2 interacting proteins

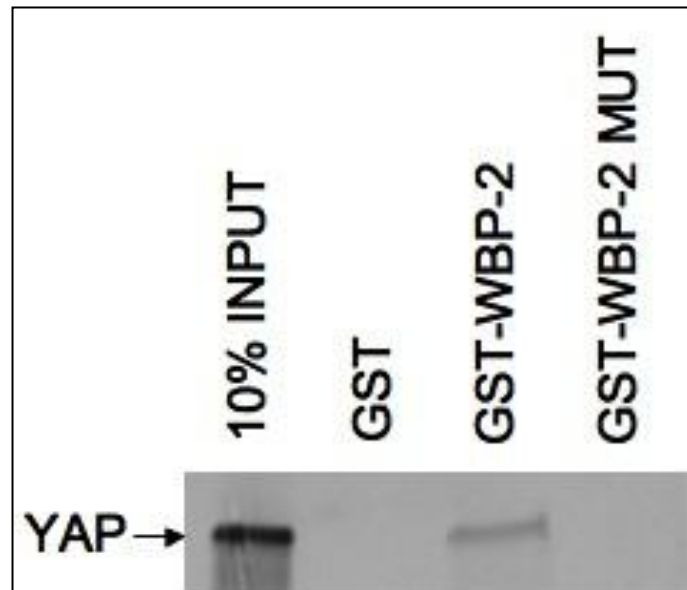


Fig. 2: PY motif of WBP-2 is essential for interaction with YAP1

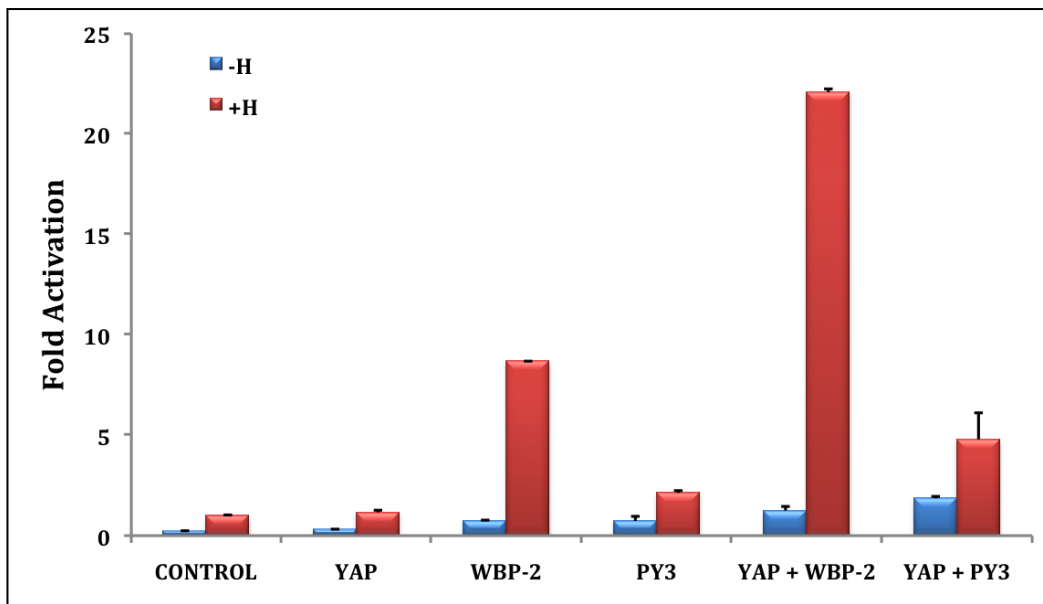


Fig. 3: YAP coactivation function is dependent on the PY motif of WBP-2

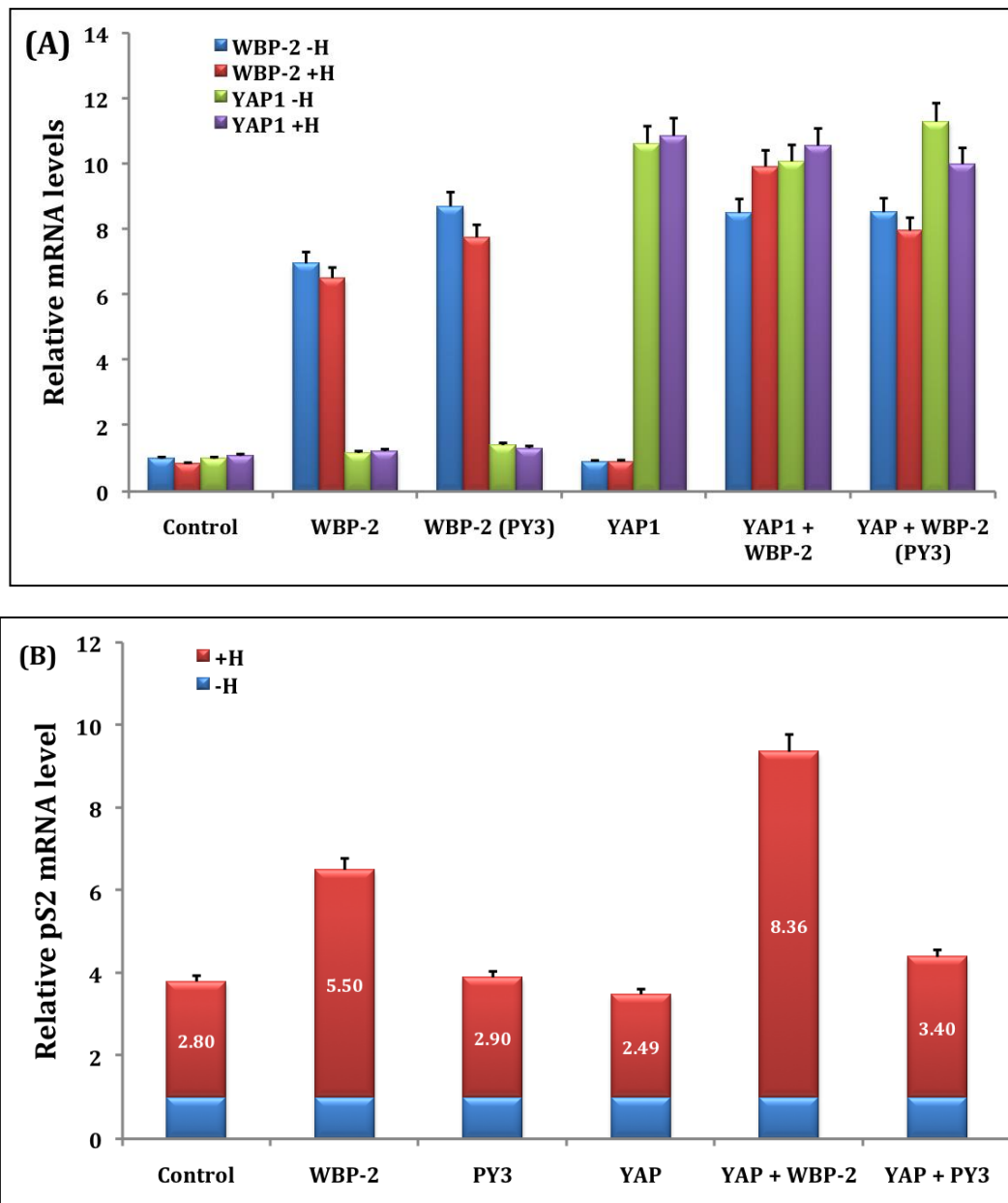


Fig. 4: A. WBP-2 and YAP1 mRNA expression levels after transfection. B. YAP-mediated coactivation of endogenous ER target gene, pS2.

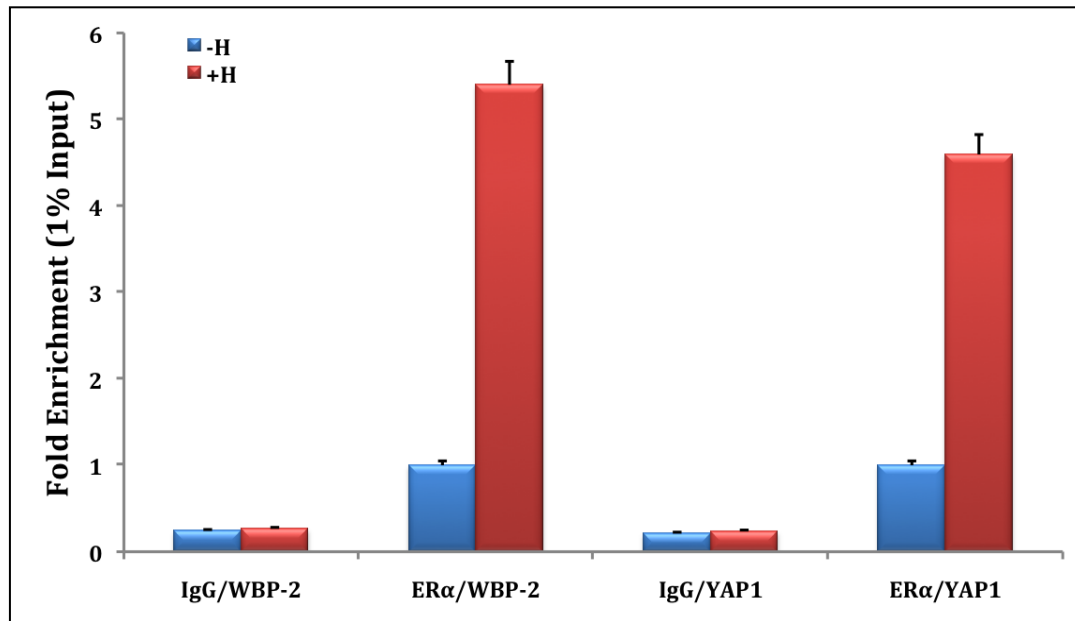


Fig. 5: WBP-2 and YAP are associated with ER α at the pS2 promoter in an estrogen dependent manner.

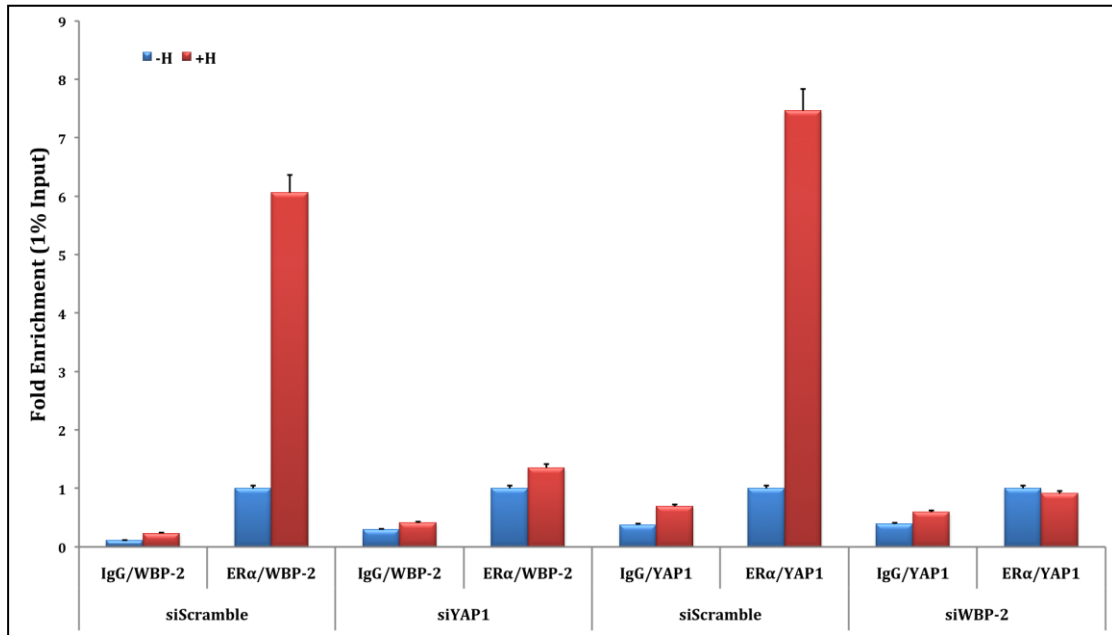


Fig. 6: Estrogen dependent recruitment and association of WBP-2 with ERα is dependent on YAP, and vice versa.

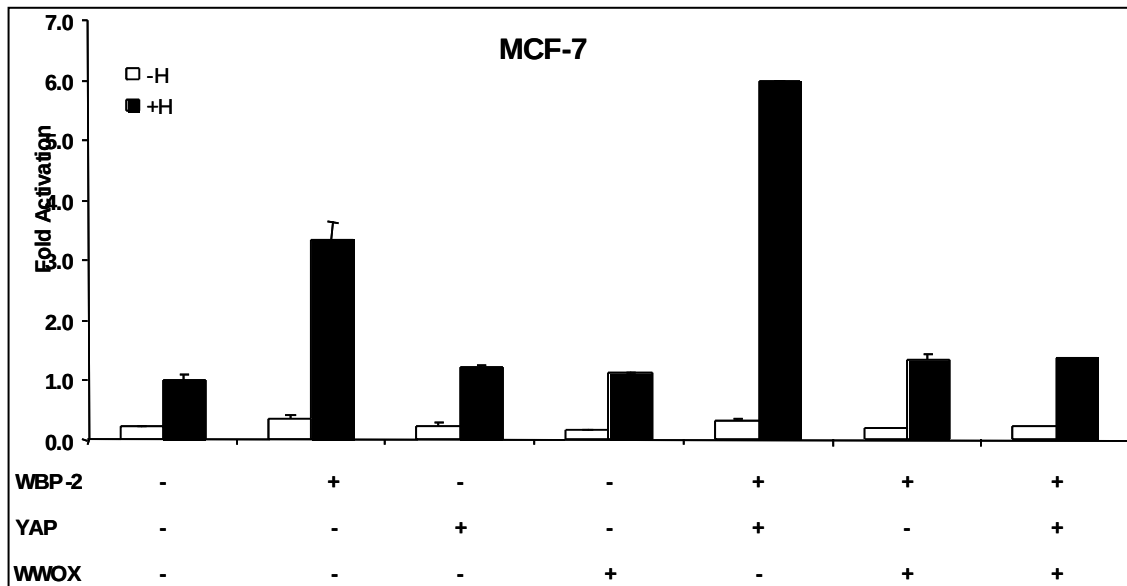


Fig. 7: WWOX1 represses the coactivation functions of WBP-2 and YAP1

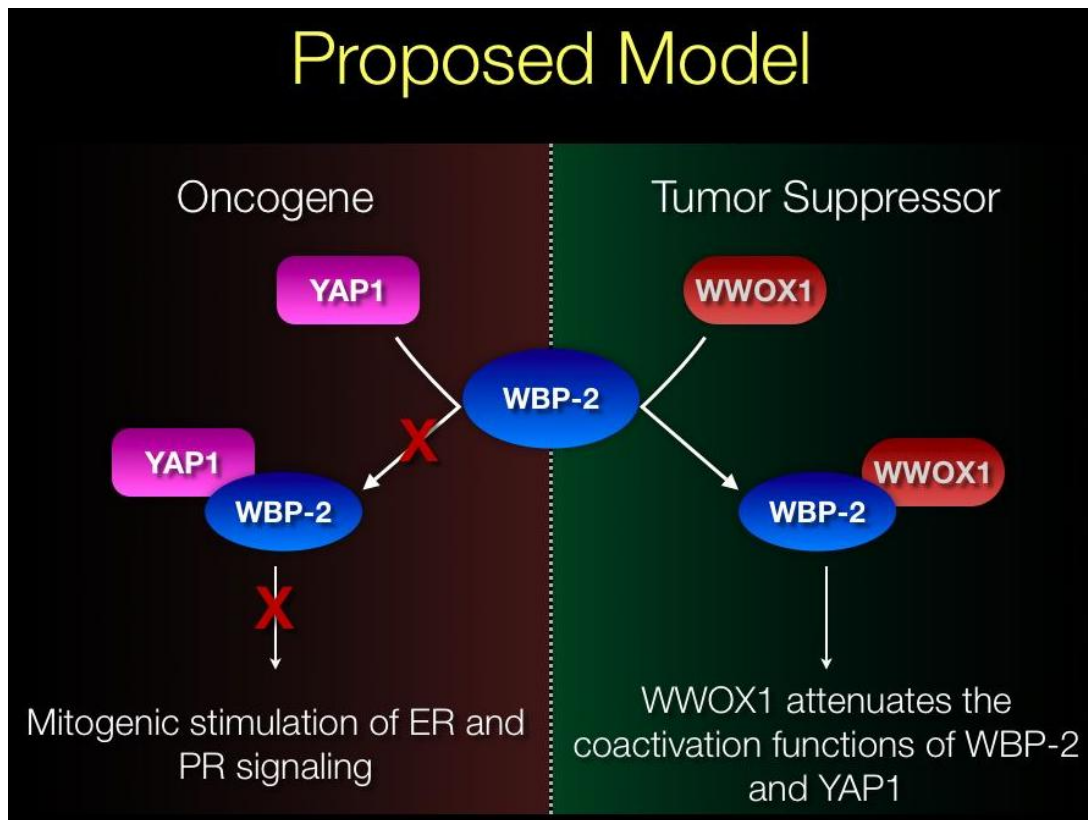


Fig. 8: Proposed model for the role of WW-domain containing proteins, YAP and WWOX1 in WBP-2-mediated ER and PR signaling