



A Generic Modeling Approach to Biomass Dynamics of *Sagittaria latifolia* and *Spartina alterniflora*

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PURPOSE: This technical note describes an ecological modeling approach that can be used to explore relationships between species of emergent aquatic vegetation communities and their environmental conditions. The modeling approach was used to evaluate the potential persistence of two desired — and quantitatively important — rhizomatous plant species under various climatological conditions: *Sagittaria latifolia*, common in freshwater systems, can produce tubers as well as rhizomes; *Spartina alterniflora* is typical for coastal marshes. Both species are endemic to the United States.

BACKGROUND: Emergent aquatic vegetation may play important roles in aquatic ecosystems. Functions attributed to “desirable” species are: stabilization of sediment and shores, amelioration of water transparency, regulation of nutrient availability in the water column and service as a habitat and food source for invertebrates, fish and waterfowl. Conversely, effects attributed to “nuisance” or “invasive” species are: excessive biomass production that interferes with human utilization of water resources, or displacement of desirable indigenous communities. Distribution and abundance of emergent vegetation in large water bodies in the Mississippi River System (UMRS) and Coastal Louisiana (CL) have changed for decades and declined in recent years. In general, variation in environmental factors such as water depth, temperature, clarity, current, wave action, and substrate characteristics would be expected to affect the distribution and production of emergent macrophytes (Gosselink and Turner 1978). *Sagittaria latifolia* (Broadleaf arrowhead) is a desired and dominant species in the UMRS, where it provides a significant annual autochthonous input (Eckblad et al. 1977). In this river system, changes resulting from the man-made modification of the hydrologic cycle include installing a system of dams in the 1930s, navigation pools with artificially-maintained high water levels, island loss due to erosion, and increased sedimentation (Bellrose et al. 1979; Eckblad et al. 1977). Environmental changes (such as increased water level and turbidity) resulting from the operation of this navigation system for barge transportation of bulk commodities, have been listed as contributing to the decline of *S. latifolia*. The latter statement is confirmed by the fact that experimental decreases in water level during the summer growth season in UMRS Pool 5 led to the increased abundance and distribution of emergent vegetation (Kenow et al. 2007). *Spartina alterniflora* (Smooth cordgrass) is a desired species in CL, where it dominates large portions of the salt marshes because of its high primary production (Kirby and Gosselink 1976; Darby and Turner 2008). Its spatial distribution is limited to the coastal areas along the entire Atlantic and the Southern Pacific seaboards of the United States. Large-scale diebacks have occurred and have been attributed to multiple causes: permanent dieback to prolonged flooding of the subsiding marsh surface (Webb et al. 1995); temporary dieback to drought from which the

Report Documentation Page				Form Approved OMB No. 0704-0188	
Public reporting burden for the collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to a penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number.					
1. REPORT DATE JAN 2011		2. REPORT TYPE		3. DATES COVERED 00-00-2011 to 00-00-2011	
4. TITLE AND SUBTITLE A Generic Modeling Approach to Biomass Dynamics of Sagittaria latifolia and Spartina alterniflora				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S)				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) U.S. Army Engineer Research and Development Center, System-Wide Water Resources Program (SWWRP), 3909 Halls Ferry Road, Vicksburg, MS, 39180				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES)				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION/AVAILABILITY STATEMENT Approved for public release; distribution unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT					
15. SUBJECT TERMS					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT Same as Report (SAR)	18. NUMBER OF PAGES 23	19a. NAME OF RESPONSIBLE PERSON
a. REPORT unclassified	b. ABSTRACT unclassified	c. THIS PAGE unclassified			

vegetation could recover rapidly by regrowth from rhizomes or more slowly in the absence of the rhizomes by seedling recruitment in the opened areas (Edwards et al. 2005).

Besides light (as affected by water level and turbidity), nitrogen (N) and phosphorus (P) are generally believed to be the most important limiting factors in aquatic systems (Hutchinson, 1975). Relationships between biomass nutrient concentrations and nutrient limitation are complex. Biomass nutrient concentrations tend to be positively correlated with nutrient supply when all other resources are sufficiently available (Guesewell and Koerselman 2002). A low concentration of N in plant biomass should reflect a low availability of N to this plant and, therefore, indicate that additional supply of N would increase the plants' biomass production. By definition, this means that N is limiting (Vitousek and Howarth 1991). If two or more nutrients (e.g., N and P) are in short supply, their availability relative to each other is likely to determine which of them is limiting. Thus, the molar ratio of N:P, rather than the individual concentrations, should indicate limitation (Koerselman and Meuleman 1996), with tissue N:P ratios less than 14 indicative of N-limited growth in terrestrial plants (Aerts and Chapin 2000). Nutrient-related growth limitation of emergent plants in natural systems has been reported, but N:P ratios have not usually been determined. In *S. latifolia*, N may limit growth under natural conditions since results of a short-term pot experiment indicated that this plant depleted the exchangeable N in natural sediments within four weeks, but left substantial P levels (Barko et al. 1988; Barko and Smart 1983). In *S. alterniflora*, it was suggested that N also limits growth under natural conditions, based on the vegetation response to N fertilization in the field (Valiela and Teal 1974; Gallagher 1975). In contrast, the average N:P ratio of 16:1 in aboveground biomass and of 37:1 in belowground biomass suggested P limitation in belowground biomass at this CL site (Darby and Turner 2008). However, growth limitation by nutrient availability may be even more complicated in plants exposed to different salinity levels, as illustrated by the results of a pot experiment in which the critical N:P ratios were determined in *S. alterniflora*. In these plants, growth was limited by N with tissue N:P ratios ≤ 13 and aboveground biomass was correlated with interstitial sediment-N concentration, but growth rate was affected by salinity (Smart and Barko 1980). The relationships between the potential persistence of both plant species, nutrient limitation, and salinity have not yet been unequivocally elucidated; consequently, placeholders have been included in the model in anticipation of supporting data.

Simulation models that include descriptions of aquatic vegetation responses to changes in physical and chemical conditions in various climates can be valuable tools for water resource managers. These models can be used to evaluate key environmental conditions in which the vegetation would persist or produce excessive biomass, with ensuing consequences for the systems in which they grow. Additionally, the models may provide insight as to how the vegetation would be affected by different management scenarios (Carr et al. 1997; Best et al. 2001; Karunaratne and Asaeda 2002; Asaeda et al. 2008). In this paper, a dynamic simulation modelling approach to emergent plant biomass formation is summarized, with light and temperature as driving variables, and including descriptions of plant responses to human influences such as management measures resulting in changes in turbidity, mechanical harvesting, grazing, and flooding. Calibration of plant responses to current velocity and nutrient limitation will be added later on when calibration values become available. This modelling approach was applied to submersed aquatic vegetation (SAV) also (Best and Boyd 2008). The approach is mathematically similar to those followed in other models for emergent vegetation, such as those developed for *Phragmites australis* (Ondok 1973; Asaeda and Karunaratne 2000; Asaeda et al. 2008) and for *S. alterniflora*

(Morris 1989; Dai and Wiegert 1996a). The approach describes plant morphology and biomass formation in relative detail, but it differs from them in that it relates ecophysiological processes to developmental cycle, using the model to simulate plant communities in different climates. In this construct, aerial shoots absorb CO₂ from air, and submerged shoots absorb it from water where CO₂ availability is assumed to be typical for hard water with an alkalinity between 0 and 300 mg L⁻¹ and a circumneutral pH; effects of changes in CO₂ availability are not included. The model species are *S. latifolia* and *S. alterniflora*; both plants have similarities in growth strategy, but are significantly different in morphology and physiology. The model has been calibrated, tested for sensitivity and validated against field data for both species. Both species have the capacity to persist in eutrophic, shallow water bodies with fluctuating water levels; both are capable of forming substantial above- and belowground biomass, thereby functioning as important marsh-characteristic elements. Important physiological differences are that *S. latifolia* fixes carbon via the C₃ photosynthetic pathway in contrast to *S. Alterniflora*, which uses the C₄ pathway; *S. latifolia* has a higher potential photosynthetic rate at light saturation for aerial shoots, and a higher species-characteristic light extinction coefficient than *S. alterniflora*; *S. latifolia* can form tubers (i.e., organs), enabling survival during adverse conditions such as drought and cold; *S. latifolia* is sensitive to increased salinity. These are characteristics which make *S. latifolia* a species that may predominate in freshwater and coastal marshes with a relatively low salinity, whereas *S. alterniflora* grows well in coastal marshes of higher salinity. With both species having different strategies to survive adverse conditions, changes in spatial distribution and replacement of one species by the other (the latter in coastal areas only) over a period of one to several years can be expected. Consequently, the described ecological model provides an ideal means to investigate the effects of relatively short-term changes in environmental conditions on the potential persistence of these two emergent species in shallow water bodies as part of restoration plans, provided detailed information on environmental conditions is available.

Besides model calibration and validation, two other aspects of the relationship between important representatives of rhizomatous plant species and environmental conditions were investigated in the present study. A dynamic ecological modeling approach was used: (i) persistence at various flood and drought conditions, and (ii) persistence under more southern climatological conditions than at the calibration site.

ECOLOGICAL MODELING APPROACH: This ecological model type simulates the carbon flow mass balance of typical emergent vegetation on a 1-m² sediment/soil with an overlying water column (Figure 1). Growth is considered as the plant dry matter accumulation, including rhizomes, and, if present, subterranean tubers, in an environment where N and P may be limiting under the prevailing weather conditions. At least one plant cohort waxes and wanes per season in different climatological conditions, varying from temperate to tropical. The rate of dry matter accumulation is a function of irradiance, temperature, CO₂ availability and plant characteristics. The rate of CO₂ assimilation (photosynthesis) of the plant community depends on the radiant energy absorbed by the canopy. The daily rate of gross CO₂ assimilation of the community is calculated from the absorbed radiation, the photosynthetic characteristics of leaves and the CO₂ availability. Calculations are executed in a set of subroutines added to the model. Part of the carbohydrates produced is used to maintain the existing biomass. The remaining carbohydrates are converted into structural dry matter (plant organs). In the conversion process, part of the weight is lost in respiration. The dry matter produced is partitioned among the various plant organs using partitioning factors, defined as a function of the phenological cycle of the community. The

dry weights (DW) of the plant organs are obtained by integration of their growth rates over time. The plant may over-winter through rhizomes and/or tubers in the sediment without or with plant biomass present. Rhizomes may persist when a critical rhizome mass is maintained. Tubers are depleted and disintegrate in the summer following the season in which they were formed. All calculations are performed on an m^2 basis. Since environmental factors and plant growth characteristics vary with plant height and water depth, in the model the growth-related processes of the aboveground plant biomass and the water column have been partitioned in 0.10-m depth layers. A relational diagram is presented in Figure 2. Seed formation has not been included in the model, because its role in maintaining established emergent plant communities in a temperate climate is minimal. Dispersal and colonization of new habitats are recognized, important characteristics of emergent plants. The latter processes, however, are better described using other modelling approaches (based on logistic regression or on descriptions of population dynamics varying in time and space), as described by Scheffer (1991).

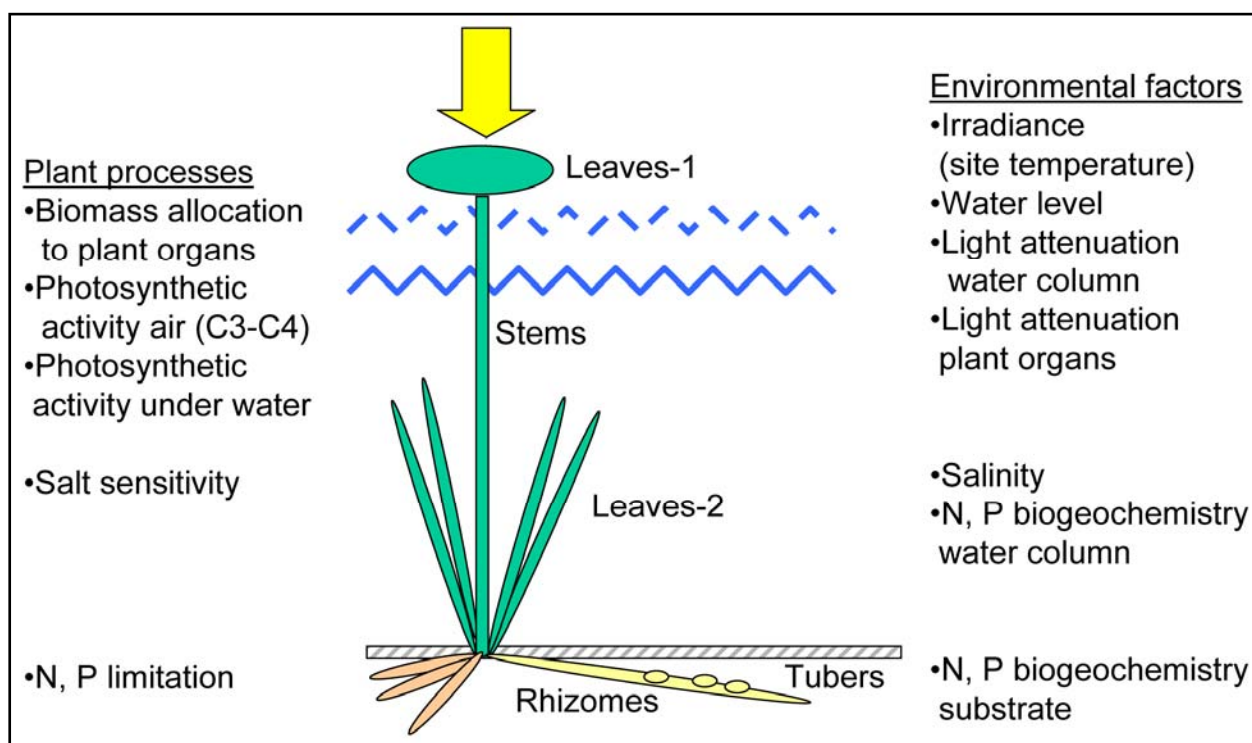


Figure 1. Schematic generic emergent plant growth model.

General features of the model include that it:

- Is operational in a one-dimensional (quasi two-dimensional) configuration
- Follows a state variable approach
- Provides that the state variable selected may be individually activated or deactivated
- Performs integration using the Runge-Kutta method
- Computes photosynthesis per second and other masses per day

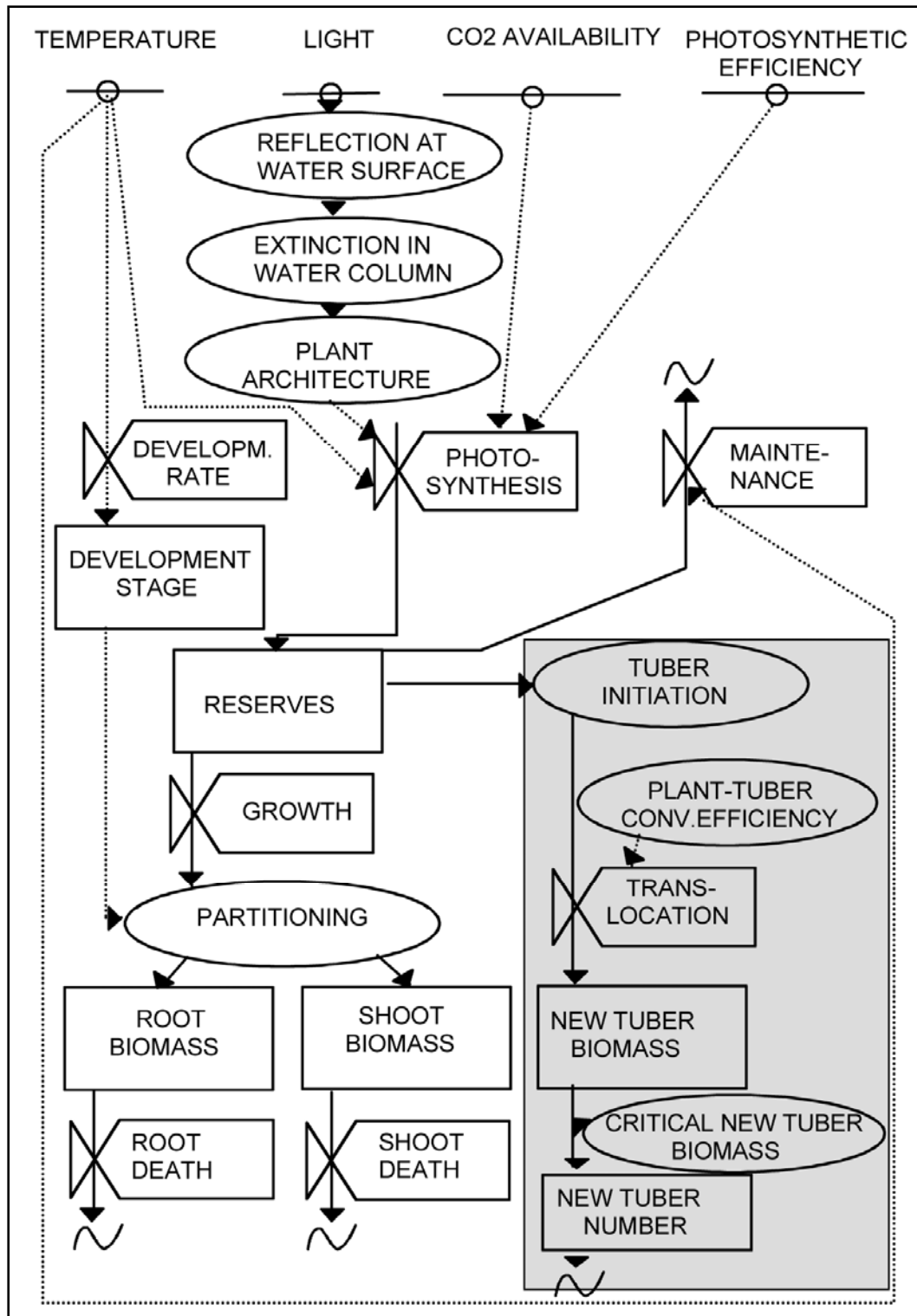


Figure 2. Relational diagram illustrating the following model processes in ARROW: (1) phenological cycle and development; (2) photosynthesis, respiration, and biomass formation; and (3) flowering, translocation, senescence and wintering organs (the latter process in grey background). Rectangles represent quantities (state variables); valve symbols, flows (rate variables); circles, auxiliary variables; underlined variables, driving and other external variables; dashed lines, information flow (symbols according to Forrester 1961).

- Operates as a stand-alone version fitted in a FORTRAN Simulation Environment (FSE) shell (Van Kraalingen 1995). Provides binary and ASCII output files, and graphics that can be viewed within a user-friendly shell. Coded in ANSI Standard FORTRAN F77.

Central Features. Central features of the model are the (1) link between the species-characteristic phenological cycle, physiological processes and environmental conditions and (2) state variable equation determining instantaneous gross photosynthesis.

Species-characteristic Phenological Cycle. The phenology of the plant community, for which the development phase can be used as a measure, is modelled as a sequence of processes that take place over a period of time, punctuated by more or less discrete events. The development phase (DVS) is a state variable in the models. The DVS is dimensionless and its value increases gradually within a growing season. The development rate (DVR) has the dimension d^{-1} . The multiple of rate and time period yields an increment in phase. The response of DVR to temperature in the model is in accordance with the degree-day hypothesis (Thornley and Johnson 1990). Calibration, according to this hypothesis, allows use of the model for the same plant species at various sites with different climates (temperature regime). The relationships between the development phase, day-of-year, and $3^{\circ}C$ day-degree sum for a temperate climate are presented in Table 1.

Each simulation starts at the first Julian day (i.e., 1 January, when the DVS has the value of 0.0). For *S. latifolia*, a species that may overwinter with rhizomes and tubers, the simulation starts using a selected rhizome weight and/or tuber bank density/individual tuber weight combination as initial values. Initiation of growth activity occurs by sprouting of the tubers, or sprouting of a fixed number of plants at a $DVS \geq 0.292$. Sprouts of the first plant cohort develop through remobilization of carbohydrates until the tubers or rhizomes are depleted. If the first plant cohort does not succeed in becoming self-supporting and DVS is less than 1.001, a second cohort sprouts from the tuber bank or rhizomes. For *S. alterniflora*, a species that overwinters with rhizomes, the simulation starts using a selected rhizome weight. The DVS values of the phenological processes in *S. alterniflora* differ from those in *S. latifolia* (Table 1).

Instantaneous Gross Photosynthesis and Biomass Formation. Light availability is an important factor influencing the distribution and abundance of aquatic plants. For emergent leaves, light attenuation only within the plant canopy occurs. Leaves submersed in water may have a small part of the irradiance reflected by the water surface, and further attenuation may occur by water and its suspended solids and by the plant itself, either covered by epiphytes or not. Emergent leaves fix carbon with a higher potential photosynthetic rate at light saturation than submersed leaves. Measured daily, total irradiance (wavelength 300-3000 nm) is used as input in the model. Only half of the irradiance reaching the water surface is considered to be photosynthetically active and is, therefore, used as a base for the calculation of CO_2 assimilation. Part of the irradiance (6 percent) can be reflected by the water surface. The subsurface irradiance can be attenuated by dissolved substances and particles (in $mg\ L^{-1}$) within the water column resulting in a site- and season-specific water extinction coefficient (Equation 1). The remaining radiation may be further reduced by epiphyte shading (Equation 1.1). The vertical profiles of the radiation within the plant layers are characterized also. The absorbed irradiance for each horizontal plant layer is derived from these profiles (Equation 1.2). The plant light extinction coefficient, K , is plant species-characteristic and

Table 1. Relationship between plant development phase (DVS), day of year, and 3°C day-degree sum in a temperate climate^a (DVRVT= 0.015; DVRRT= 0.040 for tuberforming species; DVRVT= 0.022; DVRRT= 0.015 for non-tuberforming species; at a reference temperature of 30°C).

Plant developmental phase	<i>S. latifolia</i>			<i>S. alterniflora</i>		
Description	DVS value	Day number	3°C Day-degree sum	DVS value	Day number	3°C Day-degree sum
First Julian day number → sprouting, initiation elongation, leaf expansion COHORT1	0 → 0.291	0 → 129	1 → 341	0 → 0.375	0 → 42	1 → 392
Sprouting, initiation elongation, leaf expansion → floral initiation, anthesis, induction of tuber formation* and senescence COHORT1	0.292 → 1.000	130 → 199	342 → 1682	0.376 → 1.000	43 → 99	393 → 1080
Floral initiation, anthesis, induction of tuber formation* and senescence → translocation, tuber formation* and senescence COHORT1	1.001 → 1.630	199 → 212	1683 → 1955	1.001 → 1.630	100 → 154	1081 → 2171
Translocation, tuber formation* and senescence → senesced COHORT1	1.631 → 2.000	213 → 321	1956 → 2293	1.631 → 2.000	155 → 183	2172 → 2827
Sprouting, initiation elongation, leaf expansion → floral initiation, anthesis, induction of tuber formation* and senescence COHORT2	1.001 → 1.630	199 → 212	1683 → 1955	1.001 → 1.630	100 → 154	1081 → 2171
Floral initiation, anthesis, induction of tuber formation* and senescence → translocation, tuber formation* and senescence COHORT2	1.631 → 2.000	213 → 321	1956 → 2273	1.631 → 2.000	155 → 183	2171 → 2827
Translocation, tuber formation* and senescence → senesced COHORT2	2.001 → 2.570	321 → 365	2274 → 3409	2.001 → 2.570	184 → 365	2828 → 6257
Senesced COHORT 1 and 2	2.570	365	3409	2.570	365	6257
^a Calibration was: for <i>S. latifolia</i> on field data on biomass, water transparency and depth from Upper Mississippi River Pool 9, Iowa, 1982 (Clark and Clay 1985), irradiance from La Crosse, Wisconsin, 1982, water temperature from Lansing, Iowa; for <i>S. alterniflora</i> on field data on biomass from Sapelo Island, Georgia (Dai and Wiegert 1996a,b), irradiance and air temperature from Brunswick, Georgia, 1991, and fixed water transparency and depth. * Tuber formation only in <i>S. latifolia</i> .						

assumed to be constant throughout the year. The incoming irradiance is attenuated by the shoots, part of which is absorbed by the photosynthetic plant organs: i.e., the leaves. Instantaneous rates of gross assimilation are calculated from the absorbed light energy and the photosynthesis light response of individual shoots, here used synonymously to leaves. The photosynthesis-light response of leaves is described by Equation 1.3. In the photosynthesis-light response equation, the value of potential photosynthetic activity at light saturation (AMX) is characteristic for submersed plants, and the $AMX2$ and the initial light-use efficiency (EE) typical for C_3 plants in *S. latifolia* and for C_4 plants in *S. alterniflora*. AMX and $AMX2$ are affected by temperature via a fitted, relative function, $AMTMPT$, accounting for the measured effect of daytime temperature, and enabling the calculation of the actual photosynthesis rate ($AMAX$). $AMAX$ can be affected by tissue N:P ratio via a species-characteristic, fitted, relative, function $NPREDF$, accounting for the still-to-be-measured effect of tissue N:P ratio on plant biomass production, here used synonymously for

photosynthesis. *AMAX* may also be affected by current velocity via a species-characteristic, fitted, relative, function, *REDAM1*, accounting for the measured effect of current velocity on *AMX*. Senescence may affect *AMX*. Substituting the appropriate value for the absorbed photosynthetically active radiation yields the assimilation rate for each specific shoot layer. The instantaneous rate of gross assimilation over the height of the vegetation is calculated by relating the assimilation rate per layer to the species-characteristic biomass distribution and by subsequent integration of all vegetation layers. The daily gross assimilation rate is calculated by using the Gaussian integration method. A portion of the carbohydrates formed is respired in maintenance of existing plant components and during the formation of new plant components (i.e. growth). After flowering, downward translocation of assimilates start filling the rhizomes; tubers may be induced and formed under a specific combination of temperature and day length, and senescence sets in. The *S. latifolia* model application (ARROW) was calibrated on data pertaining to a *S. latifolia* vegetation in Upper Mississippi River System Pool 9, IA, USA, 1982 (Clark and Clay 1985). The *S. alterniflora* model application (CORDG) was calibrated on data pertaining to a *S. alterniflora* vegetation on Sapelo Island, Georgia, USA, 1991 (Dai and Wiegert 1996a, b). The models simulated the dynamics of plant, rhizome and tuber biomass and tuber numbers, the latter for *S. latifolia* only, for the calibration and validation sites well over a period of one to five years. The models have been used to simulate plant, rhizome and tuber biomass and tuber numbers for other sites with temperate and tropical climates as well. Key model equations dealing with photosynthesis and nutrient limitation are provided in Appendix A, and parameters, variables, and constants are provided in Table 2. More detailed descriptions of the equations involved and model applications can be found in Best and Boyd (2007, 2008). Executable versions of the models are available at <http://el.ercd.usace.army.mil/products.cfm?Topic=model&Type=aquatic>.

$$IRZ_{i+1} = IRZ_i \times e^{(-TL \times L - K \times SC_i)} \quad (1)$$

$$IABS_i = \frac{(IRZ_i - IRZ_{i+1}) \times SC_i \times K}{(K \times SC_i + TL \times L)} \times (1.0 - EPISHD) \quad (1.1)$$

$$IABSL_i = IABS_i \times FL \quad (1.2)$$

$$FGL = SC_i \times NPREDF \times AMAX \times \left(1 - \exp \left[\frac{-EE \times IABSL_i \times 3600}{AMAX \times SC} \right] \right) \quad (1.3)$$

Table 2. Parameters, variables and constants, grouped according to model processes.							
Var/Constant	c/v^a	Value SI	Value Sa	Unit	Description	Source SI	Source Sa
Phenological cycle and development							
FLV(T)	v (tab)	0.190	0.135	Unitless	Fraction of total dry matter increase allocated to leaves as function of DVS	1	2, 3, 4
FST(T)	v (tab)	0.520	0.421	Unitless	Fraction of total dry matter increase allocated to stems as function of DVS	1	2, 3, 4
FRT(T)	v (tab)	0.290	0.444	Unitless	Fraction of total dry matter increase allocated to roots as function of DVS	1	2, 3, 3
DDTMP	v			°C	Daily average temperature (field site)		
DVRV(T)	v (tab)	0.015	0.022	d ⁻¹	Development rate after flowering as function of temperature	Calibr.	Calibr.
DVRR(T)	v (tab)	0.040	0.015	d ⁻¹	DVR prior to flowering as function of temperature	Calibr.	Calibr.
DVS	v			Unitless	Development phase	Calibr.	Calibr.
Wintering, sprouting, sprout elongation							
NPL	c	30	77	m ⁻²	Plant density	5	6
NDTUB	v	101	NA	m ⁻²	Dormant tuber density	7	
INTUB	c	0.765	NA	g DW tuber ⁻¹	Tuber size	8	
RDTU	c	0.014	NA	d ⁻¹	Relative tuber death rate (on number basis)	7	
NTUBD	v			N m ⁻²	Dead tuber number		
NTUBPD	v			N m ⁻²	Dead tuber number previous day		
NGTUB	v			N m ⁻²	Sprouting tuber number		
REMOB	v			g CH ₂ O m ⁻² d ⁻¹	Remobilization rate of carbohydrates		
ROC		0.0576	0.0576	g CH ₂ O g ⁻¹ DW d ⁻¹	Relative conversion rate of tuber/rhizome into plant material	9	9
RCSHST	c	12	NA	m g ⁻¹ DW	Relation coefficient tuber weight-stem length	9	
CRIFAC	c	0.87	0.535	g DW layer ⁻¹ plant	Critical shoot weight per 0.1-m depth layer	1	10
SURPER	c	99	NA	d	Survival period for sprouts without net photosynthesis	11	
TWGTUB	v			g DW m ⁻²	Total dry weight of sprouting tubers		
IWGRIZ	c	40.1	928	g DW m ⁻²	Initial rhizome weight		
RDRIZ	v	0.00042	0.00042	d ⁻¹	Relative rhizome death rate		
CRRIZ	c	10.6	400	g DW m ⁻²	Critical rhizome weight		
Photosynthesis, maintenance, growth, and assimilate partitioning							
SC	c			J m ⁻² s ⁻¹	Solar constant corrected for varying distance sun-earth	12	12
TL	c	0.1	0.1	m	Thickness depth layer		
IABS(i)	v			J m ⁻² s ⁻¹	Total irradiance absorbed by depth layer i		
IABSL(i)	v			J m ⁻² s ⁻¹	Total irradiance absorbed by shoots in depth layer i		
IRZ(i)	v			J m ⁻² s ⁻¹	Total photosynthetically active part of irradiance on top of depth layer i		
SC(i)	v			g DW m ⁻²	Shoot dry matter in depth layer i		
K(T)	v (tab)	0.019	0.00241	m ² g ⁻¹ DW	Plant species specific light extinction coefficient as function of DVS	1	10
EPISHD	V (tab)	0-0.43	0-0.43	Unitless	Fraction of irradiation shaded by epiphytes	13	13
AMX	c	0.0165	0.0165	g CO ₂ g ⁻¹ DW h ⁻¹	Potential CO ₂ assimilation rate at light saturation for under-water shoots	14	14
<i>(Continued)</i>							

Table 2. Continued							
Var/Constant	c/v^a	Value SI	Value Sa	Unit	Description	Source SI	Source Sa
AMX2	c	0.0620	0.011	g CO ₂ g ⁻¹ DW h ⁻¹	Potential CO ₂ assimilation rate at light saturation for above-water shoots	1, 15	4, 15
AMAX	v			g CO ₂ g ⁻¹ DW h ⁻¹	Actual CO ₂ assimilation rate at light saturation for shoots		
EE	c	0.000011	0.000014	g CO ₂ J ⁻¹	Initial light use efficiency for shoots	12	12
NPRAT	v (tab)	6-8	5-8	Unitless	Plant biomass N:P ratio		
NPREDF(T)	v (eq)	0-1	0-1	Unitless	Relative AMX factor to account for nutrient limitation		
REDF(T)	v (tab)	1	1	Unitless	Relative reduction factor for AMX to account for senescence plant parts	User def.	User def.
REDAM	c	1	1	Unitless	Relative reduction factor to relate AMX to water pH and oxygen level	User def.	User def.
REDAM1	v (tab)	0-1	0-1	Unitless	Relative reduction factor to relate AMX to water current velocity		
AMTMP(T)	v (tab)	0-1	0-1	Unitless	Daytime temperature effect on AMX as function of DVS	16	16
FGL	v			g CO ₂ m ⁻² h ⁻¹	Instantaneous CO ₂ assimilation rate per vegetation layer		
GPHOT	v			g CH ₂ O m ⁻² d ⁻¹	Daily total gross assimilation rate of the vegetation		
DMPC(T)	v (tab)	0-1	0-1	Unitless	Dry matter allocation to each plant layer	1	10
ASRQ	v			g CH ₂ O g ⁻¹ DW d ⁻¹	Assimilation requirement for plant dry matter production		
FL(T)	v (tab)	0-1	0-1	Unitless	Leaf dry matter allocation to each layer of shoot as function of DVS	1	2, 3, 4
GLV	v			g DW m ⁻² d ⁻¹	Dry matter growth rate of leaves		
GST	v			g DW m ⁻² d ⁻¹	Dry matter growth rate of stems		
GRT	v			g DW m ⁻² d ⁻¹	Dry matter growth rate of roots		
GTW	v			g DW m ⁻² d ⁻¹	Dry matter growth rate of the vegetation (excl. tubers, rhizomes)		
TWLVG	v			g DW m ⁻²	Total dry weight live leaves		
TWSTG	v			g DW m ⁻²	Total dry weight live stems		
TWRTG	v			g DW m ⁻²	Total dry weight live roots		
TGW	v			g DW m ⁻²	Total live plant dry weight (excl. tubers, rhizomes)		
MAINT	v			g CH ₂ O m ⁻² d ⁻¹	Maintenance respiration rate vegetation		
MAINTS	v			g CH ₂ O m ⁻² d ⁻¹	Maintenance respiration rate vegetation at reference temperature		
Upper biomass limit	c	950	2,250	g DW m ⁻²	Maximum plant biomass	7	2, 17
Flowering, translocation, senescence, and formation of wintering organs							
RTR	c	0.150	NA	g DW tuber ⁻¹ d ⁻¹	Maximum relative tuber growth rate at 20°C	1	
RTRL	v		NA	g DW tuber ⁻¹ d ⁻¹	Relative tuber growth rate at ambient temperature		
CVT	c	1.05	1.05	Unitless	Conversion factor for translocated dry matter into CH ₂ O	12	12
NINTUB	c	2.0	NA	N plant ⁻¹	Tuber number concurrently initiated per plant	1	
TWCTUB	c	45.9	NA	g DW m ⁻²	Total critical dry weight of new tubers	1, 5, 8	
NNTUB	v		NA	N m ⁻²	New tuber number		
RDR(T)	v (tab)	0.030	0.030	d ⁻¹	Relative death rate of leaves as function of DAVTMP (on DW basis)	1	6

(Continued)

Table 2. Concluded							
Var/Constant	c/v ^a	Value SI	Value Sa	Unit	Description	Source SI	Source Sa
RDS(T)	v (tab)	0.030	0.030	d ⁻¹	Relative death rate of stems and roots as function of DAVTMP (on DW basis)	1	6
TEFF(T)	v (tab)			Unitless	Relative effective temperature function accounting for temperature effect on maintenance respiration, remobilization, maximum tuber growth and death rates as function of temperature	Calibr.	
TRANS	v			g CH ₂ O m ⁻² d ⁻¹	Translocation rate of carbohydrates		
1. K.P. Kenow, unpublished results 2008; 2. Gallagher et al. 1984; 3. Smart and Barko 1980; 4. Best et al. 2008b; 5. Low and Bellrose 1944; 6. Dai and Wiegert 1996b; 7. Clark and Clay 1985; 8. Marburger 1993; 9. Bowes et al. 1979; 10. Morris 1989; 11. Best and Boyd 2001; 12. Penning de Vries and Van Laar 1982a,b; 13. Best et al. 2005; 14. Titus and Adams 1979; 15. Penning de Vries and Van Laar 1982a, b; 16. Best and Boyd 2003; 17. Dai and Wiegert 1996a. ^a A c indicates that the parameter is a constant. A v indicates a variable, eq and tab indicate that the parameter is implemented in the model as an equation and a table, respectively. Abbreviations: SI = <i>S. latifolia</i> ; Sa = <i>S. alterniflora</i> .							

METHODS SIMULATION STUDIES: The generic model is composed by a framework, in which the model applications ARROW and CORDG can be run simultaneously. The model requires daily values of the following environmental variables as inputs: water depth, water transparency, temperature (water or air) and irradiance. Among the required inputs, the data on water depth can be derived from local and regional stage observations obtained from a web-based database. Data on water transparency can be derived from Secchi disk observations also obtained from a web-based database using the relationship of Giessen et al (1990). Thus, the light extinction coefficient (L), required as input for these ecological models, can be derived from measured Secchi disk depths following $L \text{ (m}^{-1}\text{)} = 1.65 / \text{Secchi disk depth (m)}$. The latter relationship is valid for turbid, shallow water only. Both water depth and water transparency data can also be derived from hydrodynamic and sediment transport model results (Best et al. 2008a). Data on irradiance and air temperature can be obtained from local or regional weather stations.

In the present simulation studies the following environmental data were used as inputs: (i) a constant water depth of 0.2 m unless indicated otherwise (Table 3; 0.2 m depth is typical for shallow water bodies such as river pools, but usually daily and seasonal fluctuations occur, as documented by Best and Boyd 2008); (ii) light extinction coefficients, typical for turbid water such as river pools and peat lakes (1.81-2.0 m⁻¹; Table 3); (iii) weather data, either typical for a temperate climate (Mississippi River Pool 8 and Pool 9, near La Crosse, WI, for years 1982 (calibration) and 2006 (validation) in which field data on *S. latifolia* biomass distribution were collected; Sapelo Island, GA, for years 1991 (calibration) and 1972 (validation) in which field data on *S. alterniflora* biomass distribution were collected), or typical for a near subtropical climate where both species also abundantly grow but descriptions of wax, wane, and coexistence are still lacking, i.e., Kenner (near New Orleans), Louisiana, 2006 (exploration; Table 3).

Table 3. Variables and constants, grouped according to field site characteristics and management.							
Var/Constant	c/v ^a	Value SI	Value Sa	Unit	Description	Source SI	Source Sa
Field site characteristics^b							
DPT(T)	v (tab)	0.07-2.09	0.2	m	Water depth (field site)	1, User def.	User def.
WTMP(T)	v (tab)			°C	Daily water temperature as function of day no (field site)	User def.	User def.
L(T)	V(tab)	1.81	2.0	m ⁻¹	Water type specific light extinction coefficient as function of day no (field site)	1, User def.	User def.
WVEL	v (tab)	0-100	0-100	cm s ⁻¹	Water type specific current velocity as function of day no (field site)	User def.	User def.
TGWM(T)	v (tab)			g DW m ⁻²	Total live dry weight measured as function of day no (field site)	User def.	2, 3, User def.
NTM(T) -Va	v (tab)	101	NA	N m ⁻²	Tuber density measured as function of day no (field site)	User def.	
Management (harvesting)							
HAR	c	0 or 1	0 or 1		Harvesting switch (0=off, 1=on)	User def.	User def.
HARDAY	c	1-365	1-365	d	Harvesting day number	User def.	User def.
HARDEP	c			m	Harvesting depth (measured in 0.1-m increments from water surface)	User def.	User def.
1. Clark and Clay 1985; 2. Gallagher et al. 1984; 3. Dai and Wiegert 1996a. ^a A c indicates that the parameter is a constant. A v indicates that a variable and tab indicates that the parameter is implemented in the model as a table. Abbreviations: SI = <i>S. latifolia</i>; Sa = <i>S. alterniflora</i>. ^b Temperate field site: For SI, La Crosse, Wisconsin (lat 43° 10'N, long 91° 30'W); weather file 1982; For Sa, Brunswick, Georgia (lat 31° 15'N, long 81° 28'W); weather file 1991; Near subtropical field site: Kenner, Louisiana (lat 29° 59'N, long 90° 15'W); weather file 2006.							

SIMULATIONS

***S. latifolia* Base Runs.** ARROW was selected from the framework and run using the nominal parameter values (Table 2), field site variables (Table 3) and weather data (irradiance and air temperature, La Crosse, WI; lat 43° 30'N, long 91° 10'W) as inputs for a 1-year period. The simulated biomass of *S. latifolia* plants (composed by shoots and roots), rhizomes and tubers are shown in Figure 3. Simulated plant biomass augmented with rhizome biomass compared well with the measured 'plant' biomass, in which plants and rhizomes had not been separated during harvesting. Simulated plant biomass reached its maximum 100 days earlier than rhizome biomass, and measured "plant" biomass reached its maximum 10 days after simulated plant biomass. A comparison of simulated and measured rhizome biomass was not possible because rhizome biomass had not been determined in the calibration data set. Simulated tuber number decreased during summer and increased from August onwards to a level that was somewhat greater than initially. The thus-calibrated model simulated a stable *S. latifolia* population that persisted by sprouting from rhizomes as well as from tubers in spring, but did not attain 280 tubers m⁻² as reported by Clark and Clay (1985) in Pool 9 in 1982.

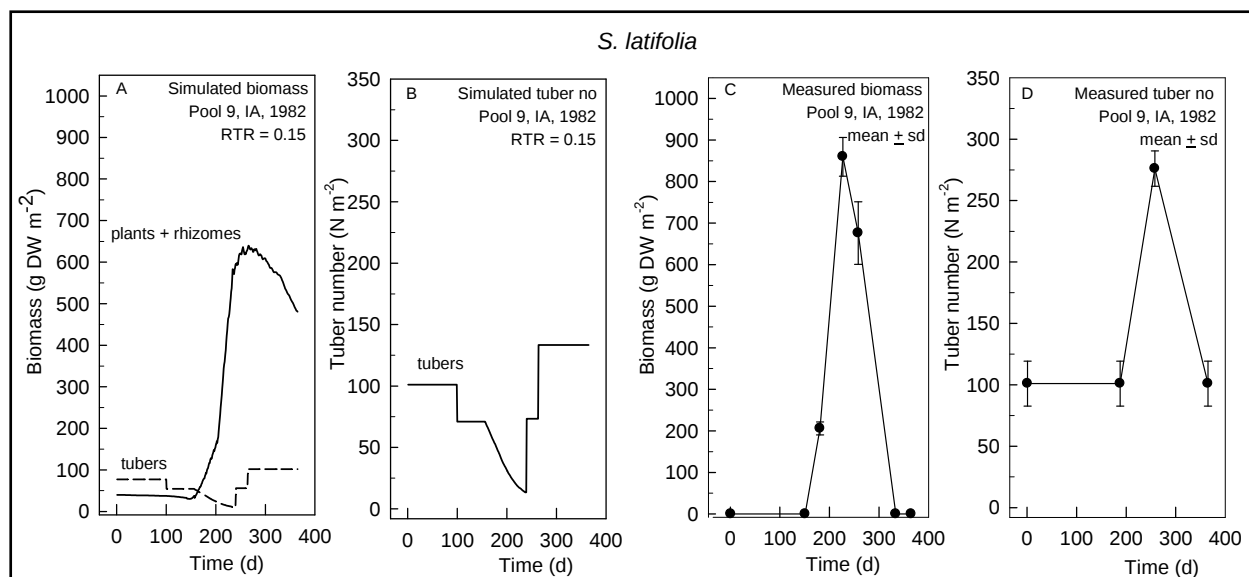


Figure 3. Simulated biomass of plants, rhizomes, and tubers (A), tuber numbers (B), measured plants (C), and measured tuber numbers (D) of *Sagittaria latifolia* in Upper Mississippi Pool 9, IA. Nominal run. Field data 1982 from Clark and Clay (1985); climatological data 1982, La Crosse, Wisconsin (lat 43° 30'N, long 91° 10'W); water depth 0.11 to 2.10 m; light extinction coefficient 1.81.

***S. latifolia* Validation Runs.** Results of the validation run indicated that simulated plant biomass was similar to measured plant biomass, but that in this case simulated tuber numbers greatly exceeded the measured ones (Figure 4).

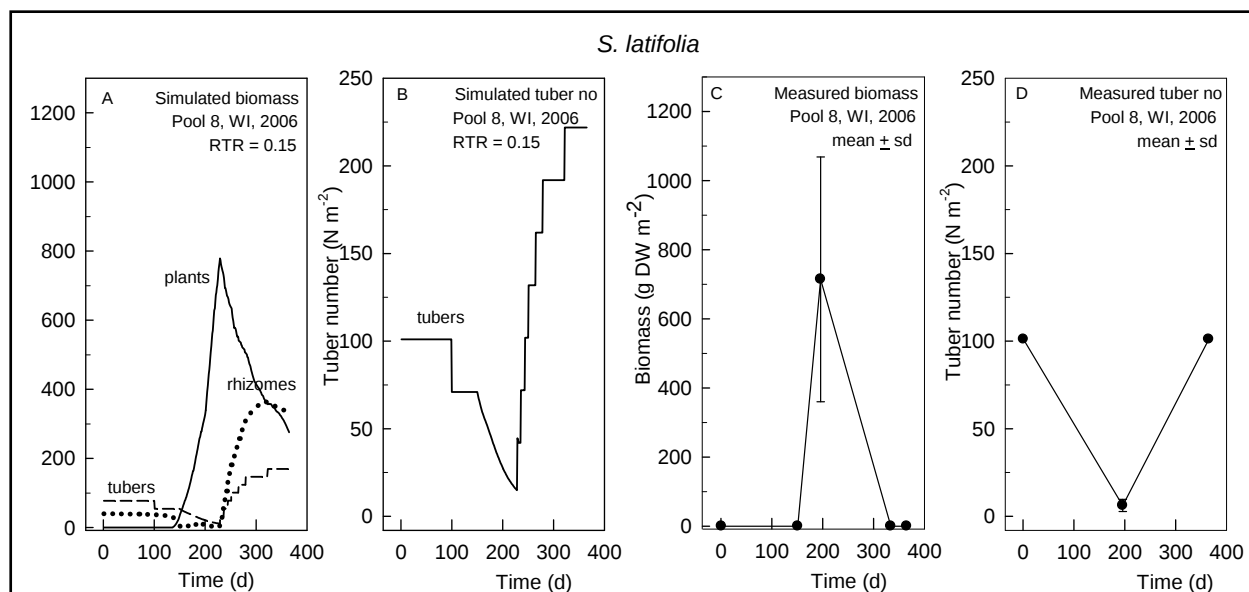


Figure 4. Simulated biomass of plants, rhizomes, and tubers (A), tuber numbers (B), measured plants (C), and measured tuber numbers (D) of *Sagittaria latifolia* in Upper Mississippi Pool 8, WI. Validation run. Field data 2006 from K. P. Kenow (unpublished Long Term Research Management Program, 2006); climatological data 2006, La Crosse, Wisconsin (lat 43° 30'N, long 91° 10'W); water depth 0 to 0.62 m; light extinction coefficient 0.98 to 2.54.

The discrepancies between the simulated and the measured tuber numbers in the calibration and validation runs could be explained in two different ways. On the one hand, the tuber data were extremely scarce and representative for only one or two points in time during the year (i.e., on 30 June and 15 September in 1982, and on 30 June in 2006), making a positive correlation unlikely. On the other hand, since measured tuber density was high at the end of summer in Pool 9 in 1982 and low in Pool 8 in 2006 (the tubers were measured at sites within waterfowl exclosures in which grazing was prevented), it could not be ruled out that plant populations produce tuber numbers which differ greatly between years in a temperate climate, making rhizomes extremely important organs for population persistence at this latitude. The latter possibility was further explored by conducting model runs for four different water level - year combinations (Figure 5). The results of these simulations indicate that simulated plant-rhizome biomass varied by a factor of 1.5, and tuber numbers were sufficient in all cases to enable persistence of the population and sprouting from both rhizomes and tubers, rendering a large variation in tuber numbers between years unlikely.

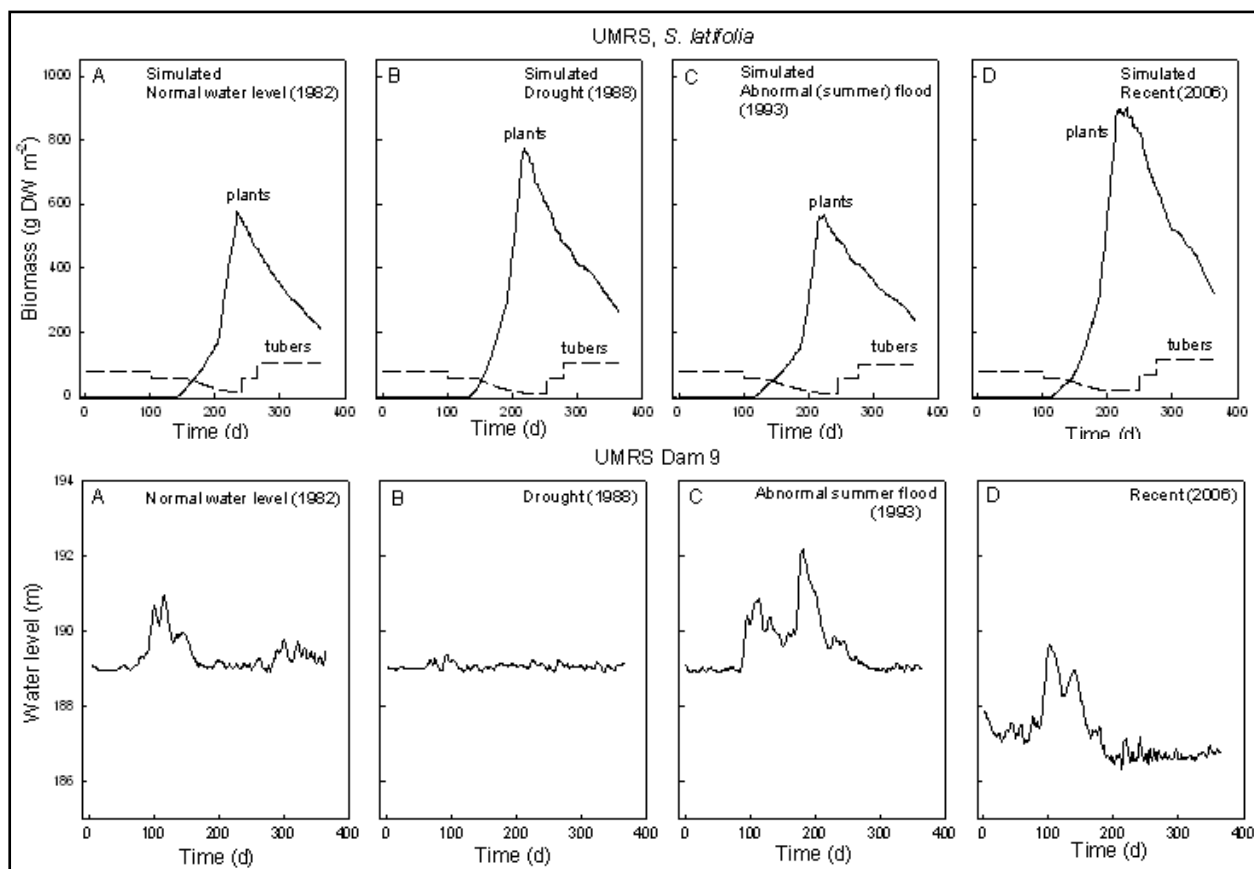


Figure 5. Simulated biomass of plants (including rhizomes) and tubers of *Sagittaria latifolia* (upper) and measured typical water level fluctuations (lower) in Upper Mississippi System Pool 8, Wisconsin. Nominal initial biomass and light extinction coefficient values; climatological data La Crosse, Wisconsin (lat 43° 30'N, long 91° 10'W).

***S. latifolia* Runs in a Subtropical Climate.** To investigate whether ARROW could be used to simulate behavior of a *S. latifolia* community in a subtropical climate, a run was conducted for a more southern site, Kenner, Louisiana (lat 29° 59'N, long 90° 15'W) (Figure 6). Results of this run indicated that simulated plant biomass exceeded plant biomass in a temperate climate by a factor of four, sufficient rhizome biomass and large numbers of tubers were produced. From this information, it can also be concluded that the model simulated a stable *S. latifolia* population that persisted by sprouting from rhizomes and from tubers in spring.

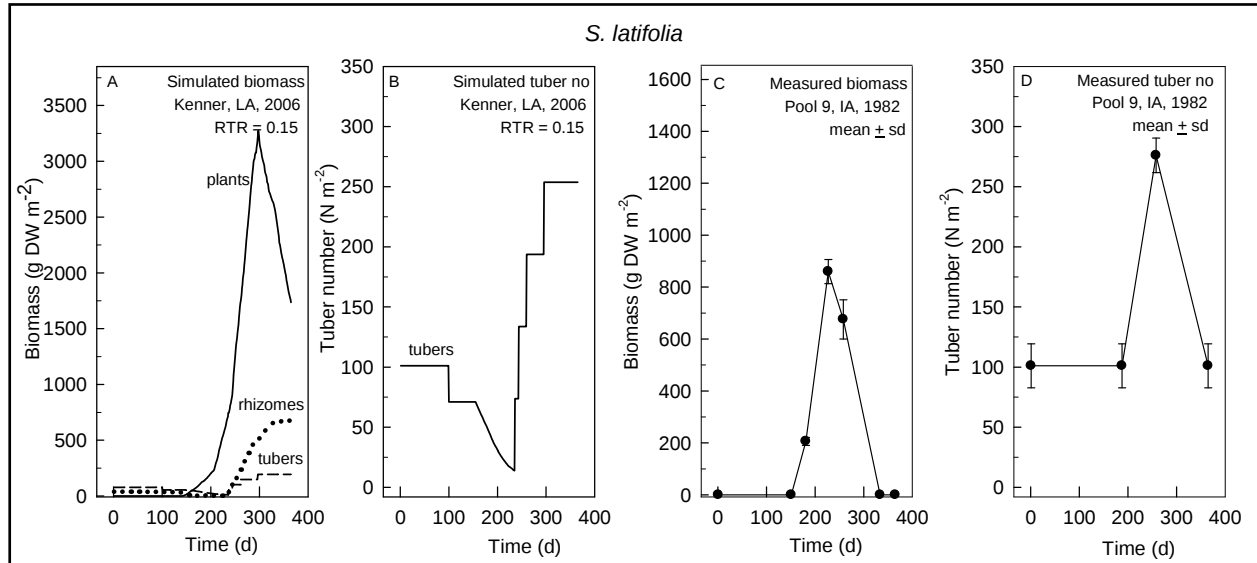


Figure 6. Simulated biomass of plants, rhizomes, and tubers (A), and tuber numbers (B), of *Sagittaria latifolia* in a more southern climate. Nominal initial biomass, light extinction coefficient, and water depth values; climatological data Kenner, Louisiana (lat 29° 59'N, long 90° 15'W). For comparison, measured biomass of plants (C), and tuber numbers (D) in Upper Mississippi Pool 9, IA (near La Crosse, Wisconsin; lat 43° 30'N, long 91° 10'W) provided.

***S. alterniflora* Base Runs.** CORDG was selected from the framework and run using the nominal parameter values pertaining to a *S. alterniflora* population in Sapelo Island, Georgia (Table 2). Field site variables (Table 3) and weather data (irradiance and air temperature, Brunswick, Georgia; lat 31° 15'N, long 81° 28'W) were used as inputs for a 1-year period. CORDG was calibrated after the 'tall' *S. alterniflora* variety, which usually grows relatively close to mean high water level and along creeks. The 'tall' variety differs in that it has a greater plant species characteristic light extinction coefficient for shoots (0.00241 m² g⁻¹ DW; Morris 1989) than the 'short' variety (0.00187 m² g⁻¹ DW; Morris 1989; Dai and Wiegert 1996a), which grows at a higher elevation of the marsh. Simulated biomass of *S. alterniflora* plants (including shoots and roots), roots, and rhizomes is shown in Figure 7. Simulated biomass showed more variation with season than measured biomass, and there was an overall good agreement between both. Simulated plant biomass reached two maxima, one at the end of March and one at the end of October. Measured plant (shoot plus root) biomass showed one maximum at the end of August, was only determined at three points in time (in January, end of August and end of December), and mean values ± standard deviations matched simulated plant biomass values. Measured data on roots and rhizomes were scarce, without replicates, inhibiting unequivocal matching of measured and simulated values – but measured and simulated biomass were in the same order of magnitude as were the trends. The thus-calibrated model simulated a stable *S. alterniflora* population that

persisted by sprouting from rhizomes in spring, with aboveground biomass (composed by shoots) reaching a lower maximum of 900 g DW m⁻² than 1500 g DW m⁻² of belowground biomass (composed by rhizomes and roots), and rhizome mass exceeding plant mass in winter.

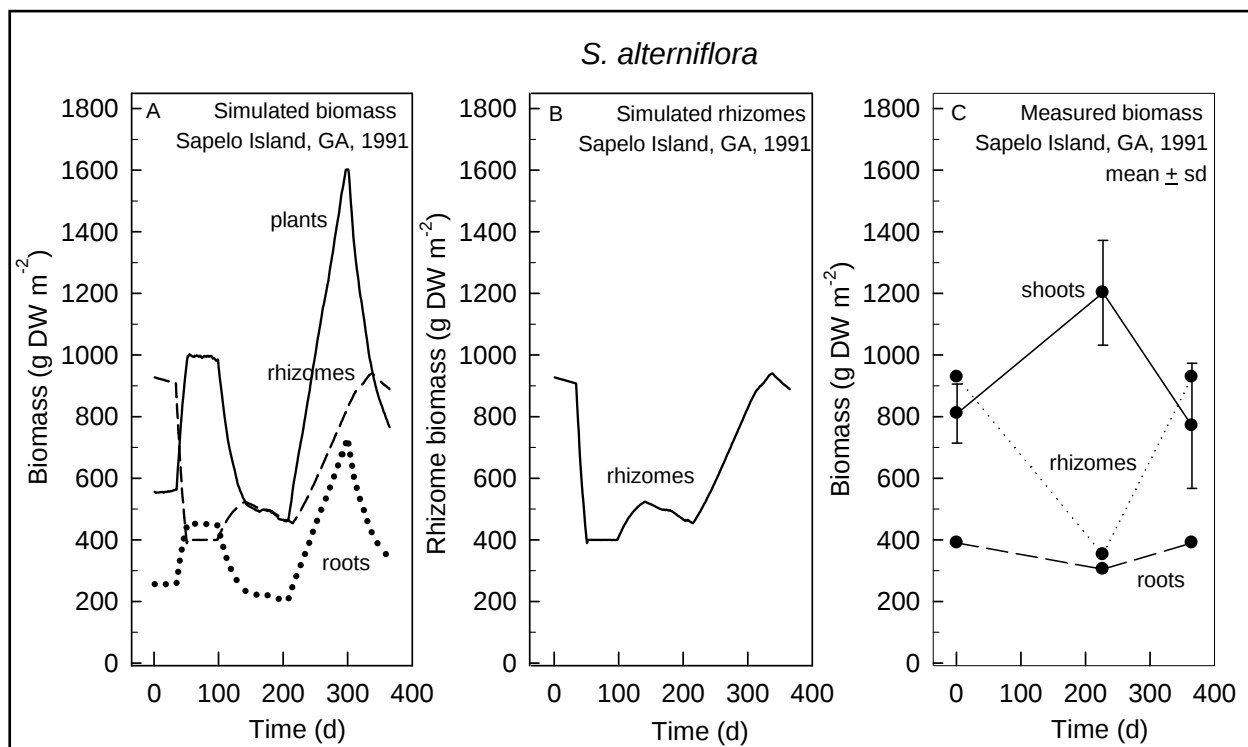


Figure 7. Simulated biomass of plants, roots and rhizomes (A), rhizomes (B), and measured shoots, roots and rhizomes (C) of *Spartina alterniflora* on Sapelo Island, Georgia. Nominal run. Field data 1991 from Dai and Wiegert (1996 a, b); climatological data 1991, Brunswick, Georgia (lat 31° 15'N, long 81° 28'W); water depth 0.20 m; light extinction coefficient 2.0.

***S. alterniflora* Validation Runs.** Results of the validation run indicated that simulated shoot biomass (plants minus roots) was similar to measured shoot biomass (Figure 8). It was not possible to compare simulated to measured belowground biomass, because no belowground biomass was determined in the validation data set.

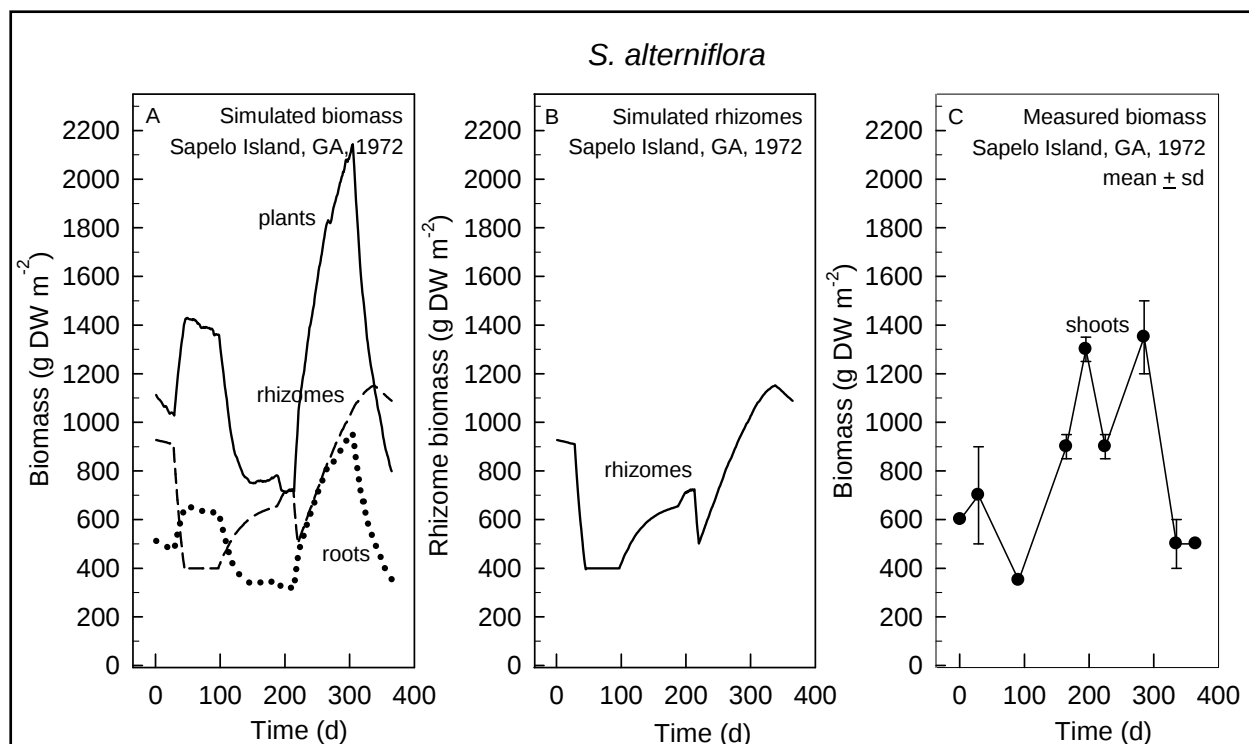


Figure 8. Simulated biomass of plants, roots and rhizomes (A), rhizomes (B), and measured shoots (C) of *Spartina alterniflora* on Sapelo Island, Georgia. Validation run. Field data 1972 from Gallagher et al. (1980); climatological data 1972, Brunswick, Georgia (lat 31° 15'N, long 81° 28'W); water depth 0.20 m; light extinction coefficient 2.0.

***S. alterniflora* Runs in a Subtropical Climate.** To investigate whether CORDG could be used to simulate behavior of a *S. alterniflora* community in a subtropical climate, a run was conducted for the same, more southern site as was used to test ARROW, (i.e., Kenner, Louisiana) (Figure 9). Results of this run indicated that simulated plant, root, and rhizome biomass in Kenner were a factor of 1.4 greater than those on Sapelo Island: the model simulated a stable *S. alterniflora* population that persisted by sprouting from rhizomes in spring. Results of an additional run for the “short” plant variety, which forms a large part of CL marshes, indicated an 18 percent reduction in biomass production compared to production by the “tall” variety by a stable plant population.

Sensitivity Analysis. Sensitivity analysis of a simulation model must be undertaken to assess the parameters likely to strongly affect model behavior. The current analysis is based on the effect of a change in one parameter while all other parameters are kept the same. The parameter under study was changed and 1-year simulations were conducted under nominal environmental conditions. The nominal parameter values (as presented in Table 2) were chosen as a reference level. The results were compared with those of a nominal run. Each parameter was increased once by 20 percent and decreased once by 20 percent. The relative sensitivity (RS) of a parameter was then defined as the relative change in the variable on which the effect was tested divided by the relative change in the parameter (Ng and Loomis 1984). The effects of thirteen parameters

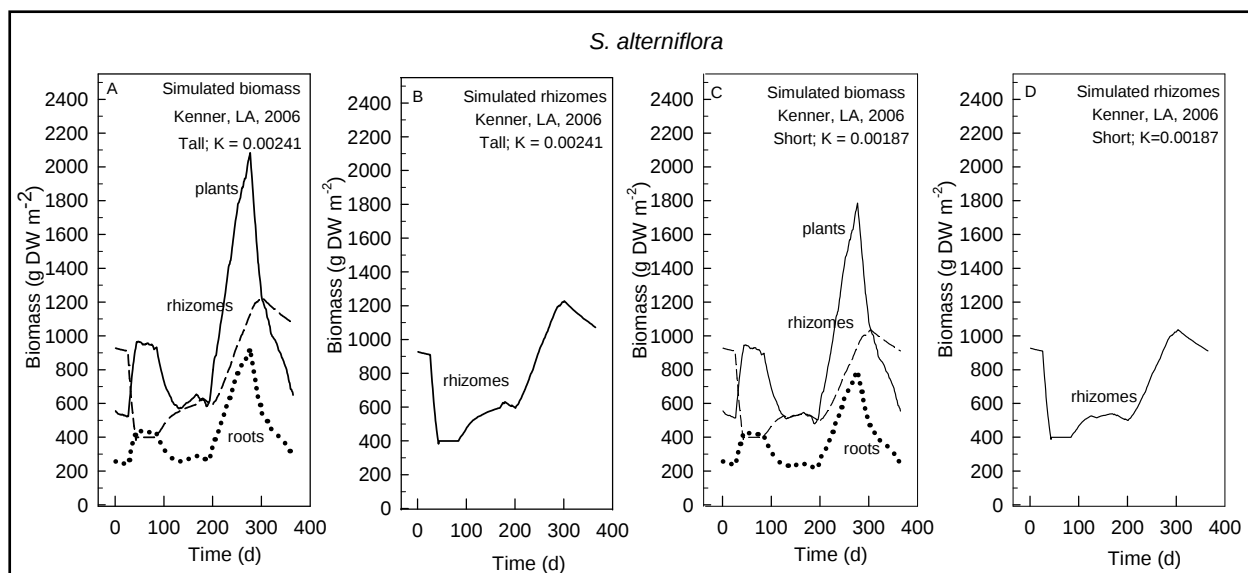


Figure 9. Simulated biomass of plants, roots and rhizomes (A = tall, C = short vegetation), and rhizomes (B = tall, D = short vegetation), of *Spartina alterniflora* in a more southern climate. Nominal initial biomass, light extinction coefficient, and water depth values; climatological data Kenner, Louisiana (lat 29° 59'N, long 90° 15'W).

on three state variables, representing different plant biomass compartments, were tested. A model variable is considered sensitive to a change in the value of a parameter at $RS > 0.5$ and < -0.5 .

$$RS = \frac{(yield_i - yield_r) / yield_r}{(param_i - param_r) / param_r}$$

where

$yield_i$ = value at parameter value i

$yield_r$ = value at reference parameter value r

$param_i$ and $param_r$ = as above

The three state variables, maximum plant biomass, end-of-year tuber number and end-of-year rhizome biomass in the model were sensitive to parameter changes, particularly those affecting carbon capture (*AMX2*-potential CO₂ assimilation rate at light saturation for above-water shoots; and *EE*- initial light use efficiency for shoots; Table 4). In addition, end-of-year rhizome and/or tuber number was sensitive to development rates (*DVRVT*, *DVRRT*) and relative death rate (*RDR*) and in SLAT only to plant density (*NPL*), initial rhizome weight (*IWGRIZ*) and relative tuber growth rate (*RTR*).

Table 4. Relative sensitivity (RS) of selected state variables to deviations in parameter values from their nominal values as presented in Table 2. The RS of a parameter is the relative change in the variable on which the effect was tested divided by the relative change in the parameter. A model variable is considered sensitive to a change in the value of a parameter at $RS > 0.5$ and < -0.5 . Results were obtained in 1-year simulations under nominal conditions. Sensitive values shaded.

Parameter		Relative sensitivity				
Name	Value	<i>S. latifolia</i>			<i>S. alterniflora</i>	
		Max. plant biomass	EOY ¹ rhiz. biomass	EOY ¹ tuber no	Max. plant biomass	EOY ¹ rhiz. biomass
DVRVT	+20%	0.25	0.48	0.12	-0.11	-0.09
	-20%	0.42	4.99	2.56	-0.07	-0.15
DVRRT	+20%	0.25	0.48	0.12	-0.68	-0.76
	-20%	0.24	2.85	1.46	-1.26	-0.19
NPL	+20%	0.04	0.01	0.85	-0.55	-0.38
	-20%	0.04	0.01	-0.90	-0.04	-0.03
INTUB	+20%	0.04	0.01	0.0	NA	NA
	-20%	0.04	0.01	-2.20	NA	NA
ROC	+20%	0.03	0.00	-0.01	0.05	0.04
	-20%	0.03	0.01	-0.02	0.00	0.00
RTR ¹	+20%	-0.07	-4.99	-0.98	NA	NA
	-20%	-0.07	-0.07	0.0	NA	NA
RDTU	+20%	0.00	0.00	-0.14	NA	NA
	-20%	0.00	0.00	-0.20	NA	NA
IWGRIZ	+20%	0.12	0.02	0.00	0.20	0.20
	-20%	0.17	4.99	2.49	0.02	0.10
TRAFAC	+20%	-0.30	0.14	0.02	-0.13	0.37
	-20%	-0.30	4.99	2.54	-0.33	0.31
AMX	+20%	0.00	0.00	0.00	0.00	0.00
	-20%	0.00	0.00	0.00	0.00	0.00
AMX2	+20%	1.11	0.00	0.00	0.75	0.78
	-20%	0.69	4.99	2.34	2.05	2.69
EE	+20%	0.90	0.92	2.49	1.46	1.42
	-20%	1.02	3.66	1.35	2.04	2.75
RDR	+20%	-0.17	-4.99	-2.48	-1.95	-2.28
	-20%	-0.21	-0.53	0.00	-0.41	-0.98

Note: ¹ EOY - end of year

The sensitivity of maximum plant biomass and end-of-year tuber number/rhizome biomass to changes in environmental factors was assessed by following the same approach as for sensitivity analysis of the model parameters. For this purpose, parameter changes were based on value ranges taken from literature, which sometimes differed more than 20 percent from the nominal parameter values presented in Table 4. Also in this analysis, the model proved to be sensitive (Table 5). All state variables were sensitive to changes in climate. Maximum plant biomass was less sensitive in SLAT than in SPALT. Changes in the light reflection coefficient at the water surface (RC) had no significant effect, while changes in the other tested parameter values had

effects which decreased in the order of the water type's specific light extinction coefficient (*LT*) greater than water depth (*DPTT*).

Table 5. Environmental factor analysis, expressed as relative sensitivity (RS) of selected state variables to deviations in parameter values from their nominal values, as presented in Table 2. The RS of a parameter is the relative change in the variable on which the effect was tested divided by the relative change in the parameter. A model variable is considered sensitive to a change in the value of a parameter at RS > 0.5 and < -0.5. Results were obtained in 1-year simulations under nominal conditions. Sensitive values shaded.						
Parameter		Relative sensitivity				
Name	Value	<i>S. latifolia</i>			<i>S. alterniflora</i>	
		Max. plant biomass	EOY ³ rhiz. biomass	EOY ³ tuber no	Max. plant biomass	EOY ⁵ rhiz. biomass
Climate ¹	Lat 43°30' N					
	Lat 29°59' N	-11.33	-3.31	1.35		
Climate ²	Lat 31°N					
	Lat 29°59' N				-2.05	-1.58
RC ⁴	1.00	-0.06	-0.06	-0.06	-0.03	-0.03
	0.00	-0.06	-0.05	0.00	-0.11	-0.10
LT	+20%	-0.44	-4.99	-2.47	-1.13	-0.93
	-20%	-0.45	-0.28	0.00	-0.62	-0.62
DPTT	+20%	-0.15	-4.99	-2.49	0.00	0.00
	-20%	-0.23	-0.03	0.00	0.00	0.00
¹ Climates at La Crosse, Wisconsin (lat 43° 30' N), Kenner, Louisiana (lat 29° 59' N) ² Brunswick, Georgia (lat. 31° N) ³ EOY – end of year ⁴ Light reflection coefficient at water surface; to enable calculation of the relative sensitivity, a very low value of 0.000001 was used.						

SUMMARY: A dynamic simulation modelling approach to emergent plant biomass formation has been developed to provide a tool for water resource managers. It is now possible to evaluate key environmental conditions in which emergent aquatic vegetation would persist or produce excessive biomass with ensuing consequences for the systems in which they grow, whether they are affected by management measures or not. The generic model is composed a framework, in which the model applications ARROW for *Sagittaria latifolia* and CORDG for *Spartina alterniflora* can be run simultaneously. The model describes major, carbon flow-based ecophysiological processes and biomass dynamics of two common plant species. It also contains unique descriptions of: (1) species-characteristic vertical distribution of shoot biomass which enables the calculation of the fraction of irradiance actually available for absorption by the plant; (2) recalculation procedures of this vertical distribution with daily changes in water level and/or shoot mass removal at various heights and levels within the water column, which enables the evaluation of regrowth potential; and (3) relationships of plant process parameters with site-specific climate which enables the evaluation of effects of different climates. Generally, a good fit was found between simulated and measured biomass in the field. Sensitivity analysis showed that the model is very sensitive to changes in process parameters influencing carbon flow.

PRODUCT DEVELOPMENT AND AVAILABILITY: Aquatic plant growth models are available to both U.S. Army Corps of Engineers (USACE) and non-USACE interested parties. The model can be downloaded from the following URL: <http://el.erdcl.usace.army.mil/products.cfm?Topic=model&Type=aquatic>. Model descriptions and user manuals can be downloaded from the same web-page.

POINTS OF CONTACT: This technical note was prepared by Dr. Elly P.H. Best and William A. Boyd, research biologist and mathematician, respectively, U.S. Army Engineer Research and Development Center, Environmental Laboratory; and Kevin P. Kenow, Upper Midwest Environmental Sciences Center, USGS, La Crosse, Wisconsin. The model was conducted as an activity of the ecological model development work unit of the System-Wide Water Resources Program (SWWRP). For information on SWWRP, please visit <https://swwrp.usace.army.mil/> or contact the Program Manager, Dr. Steven L. Ashby at Steven.L.Ashby@erdcl.usace.army.mil. Questions about this technical note may be addressed to Mr. Boyd at (601-634-3705; William.A.Boyd@usace.army.mil). This technical note should be cited as follows:

Best, E. P. H., W. A. Boyd, and K. P. Kenow. 2011. *A Generic Modeling Approach to Biomass Dynamics of Sagittaria latifolia and Spartina alterniflora*. SWWRP Technical Notes Collection. ERDC TN-SWWRP-11-1, Vicksburg, MS: U.S. Army Engineer Research and Development Center. <https://swwrp.usace.army.mil/>

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