

***An in silico* model of endotoxic shock mediators**

12 March 2012

**Meghan Makley
Jeff Gearhart
Mark Lisanby
Eric Hack**

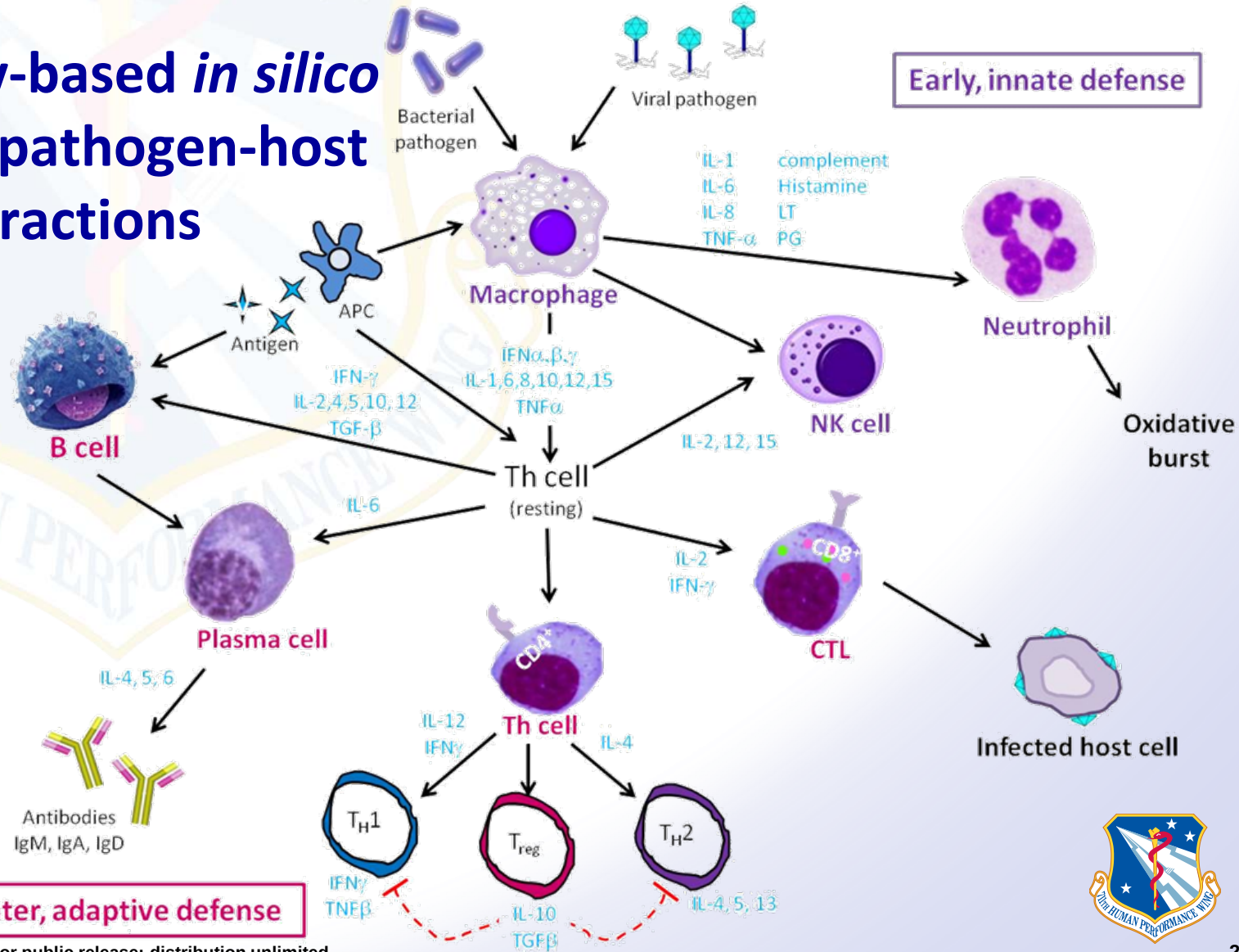


**Henry M. Jackson Foundation
Air Force Research Laboratory
711th Human Performance Wing
Bioeffects Division
Molecular Bioeffects Branch
Wright-Patterson AFB OH 45433**

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
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1. REPORT DATE (DD-MM-YYYY) 12-03-2012		2. REPORT TYPE Interim Tech Report		3. DATES COVERED (From - To) 29 Apr 10 – 12 Mar 12	
4. TITLE AND SUBTITLE An in silico model of endotoxic shock mediators (Briefing Charts)				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER	
				5c. PROGRAM ELEMENT NUMBER 62202F	
6. AUTHOR(S) Makley, Meghan K. ¹ ; Gearhart, Jeffery M. ¹ ; Poepelman, Lee ¹ ; Hack, Charles E. ¹				5d. PROJECT NUMBER ODT	
				5e. TASK NUMBER WP	
				5f. WORK UNIT NUMBER ODTWP005	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) ¹ HJF. 2729 R St. Bldg 837. WPAFB OH 45433-5707				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory 711 Human Performance Wing Human Effectiveness Directorate Bioeffects Division Molecular Bioeffects Branch Wright-Patterson AFB OH 45433-5707				10. SPONSOR/MONITOR'S ACRONYM(S) 711 HPW/RHDJ	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S) AFRL-RH-FS-TR-2012-0021	
12. DISTRIBUTION / AVAILABILITY STATEMENT Distribution A: Approved for public release; distribution unlimited. PA Case # TSRL-PA-0024, cleared 5 Mar 12.					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT Biologically-based <i>in silico</i> models of pathogen-host interactions are being designed in our lab to predict the time-course of pathogenic infection in humans. Macrophages respond to lipopolysaccharides (LPS), including the release of potent lipid autacoids, causing a cascade of events leading to endotoxic shock. However, animals have been shown to vary in response and susceptibility to <i>E. coli</i> endotoxin: guinea pig > hamster > mouse. To establish a sound basis for interspecies extrapolation, a pathogenesis model is being extended to encompass endotoxic shock. Exposing experimental animals to aerosols of LPS elicits bronchoconstriction, activation of alveolar macrophages, and recruitment of inflammatory cells into airways. These effects have been attributed to a potent lipid autacoid, platelet-activating factor (PAF). Species differences in the biomodulatory effects and mechanisms of PAF are similar to those seen with endotoxin. In guinea pigs, PAF (2 ug/kg IV) causes bronchoconstriction and hypotension in seconds and lethality within 25 minutes. In rats, however, 3 ug/kg of PAF had a negligible impact on heart rate. Therefore, a dynamic model for PAF was developed to link a pathogen's kinetics and host response. The current model focuses on kinetics and receptor binding of PAF and its antagonist ginkgolide B (GB). The kinetic models include plasma, red blood cell, lung, heart, and rapidly and slowly perfused tissues, with IV and inhalation exposure routes, and pathways for binding and elimination of PAF. Kinetic parameters were from the literature. The model was used to simulate experimental exposures to PAF and GB, revealing potential explanations for species differences in sensitivity to PAF. Internal dose metrics were generated and correlated with observed signs of infection and lethality in an attempt to identify the most appropriate metrics for predicting adverse effects. This model of pathogen kinetics and these dose metrics help to elucidate mechanisms of host response dynamics and improve cross-species extrapolation of response data.					
15. SUBJECT TERMS <i>In silico</i> model, endotoxic shock, ginkgolide B, kinetics, dosimeter					
16. SECURITY CLASSIFICATION OF: UNCL			17. LIMITATION OF ABSTRACT SAR	18. NUMBER OF PAGES 31	19a. NAME OF RESPONSIBLE PERSON Mark Lisanby
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code) NA



Biologically-based *in silico* models of pathogen-host interactions





Our ongoing efforts

Immune system response model

- Predict time-course of pathogenic infection in humans
- Quantify systemic response to pathogen exposure
- *F. tularensis* as our case study

Need a 'bridge' to eventual health outcome

- Mediators of shock

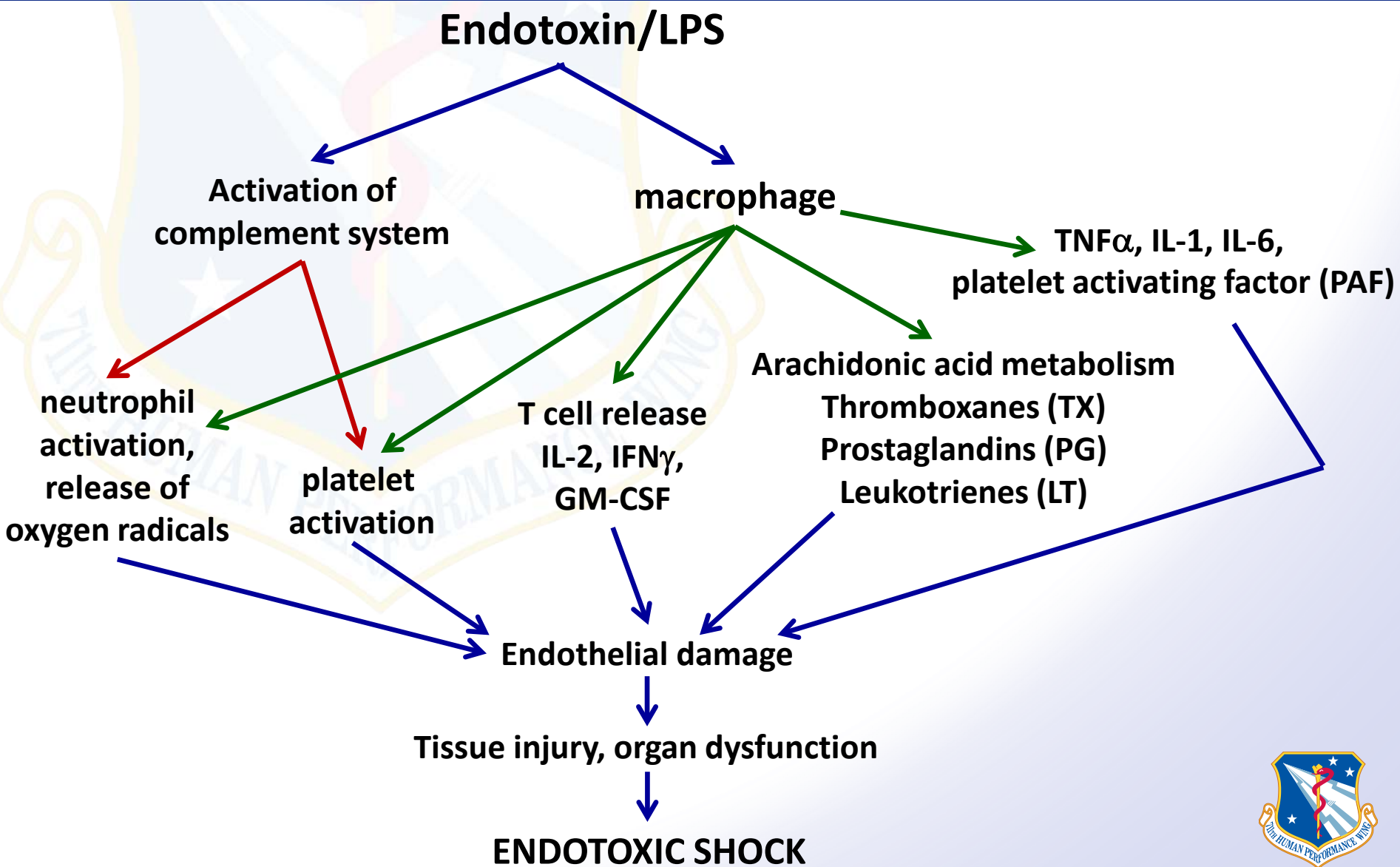
Endotoxin/lipopolysaccharide (LPS)

Principal component of gram-negative bacteria cell wall



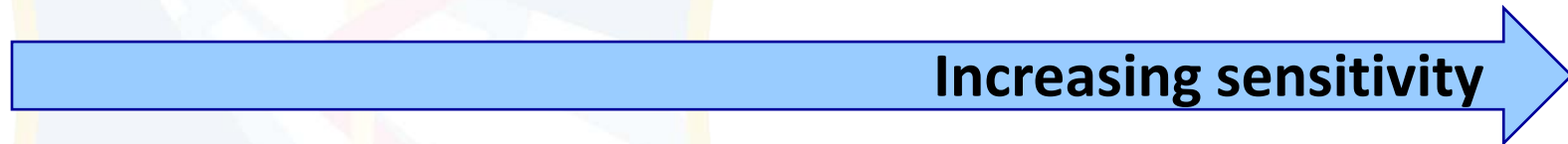


Mediators of endotoxic shock





Species differences in sensitivity to *E. coli* endotoxin



mouse
rat

macaques

human

guinea pig
rabbit

Reason(s) for species-dependent sensitivity to endotoxin?

Hypothesis – Due to differences in mediator kinetics/dynamics?





Vasoactive lipid mediators in endotoxic shock



Eicosanoids can be detected in circulation at high enough concentrations to be responsible for events in endotoxic shock

- High PG levels in circulation of animals subjected to endotoxin
- Increased plasma TXB₂ in humans suffering from severe septic shock
- Endotoxemia and sepsis: Blood PAF levels are elevated





Vasoactive lipid mediators in endotoxic shock



Synthesis inhibitors or receptor antagonists of lipid mediators are capable of modifying the course of endotoxic shock

- Lipoxygenase (LOX) inhibitors protect mice and rats from lethal endotoxemia
- TXA₂ synthetase inhibitors are effective in rat endotoxic shock
- TXA₂ receptor antagonists block development of pulmonary hypertension in endotoxemia
- PAF antagonists (Ginkgolide B, GB) protect rats, mice, pigs, and humans from injurious effects of LPS





Aims



To elucidate wide range of sensitivity to LPS between species

- Looked into differences in kinetics and dynamic responses to downstream mediators (PAF)
- Literature suggests species differences in PAF response in guinea pig, human, and rat (in decreasing order of sensitivity)





Aims



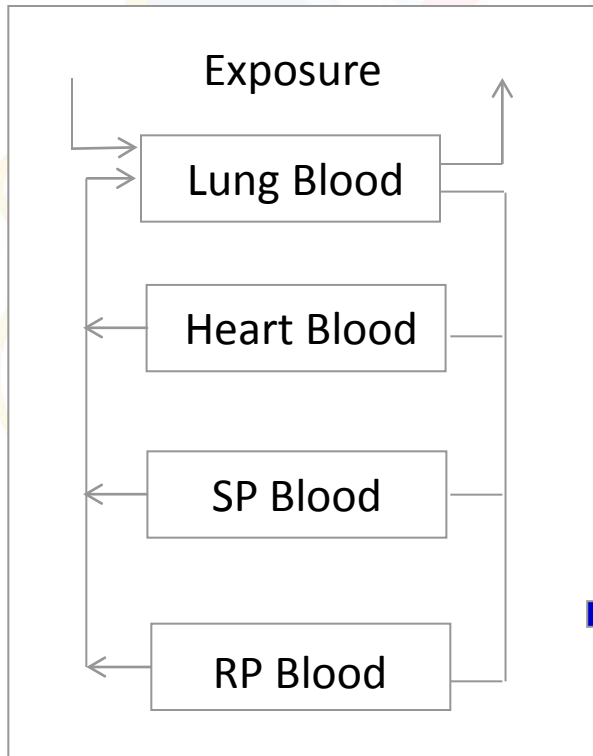
Develop a biologically-based *in silico* model of PAF to give insights into dynamic evolution of endotoxic shock

- Elucidate link between kinetics and biological response
- Simulate kinetic data reported in literature
- Extrapolate animal response data to humans

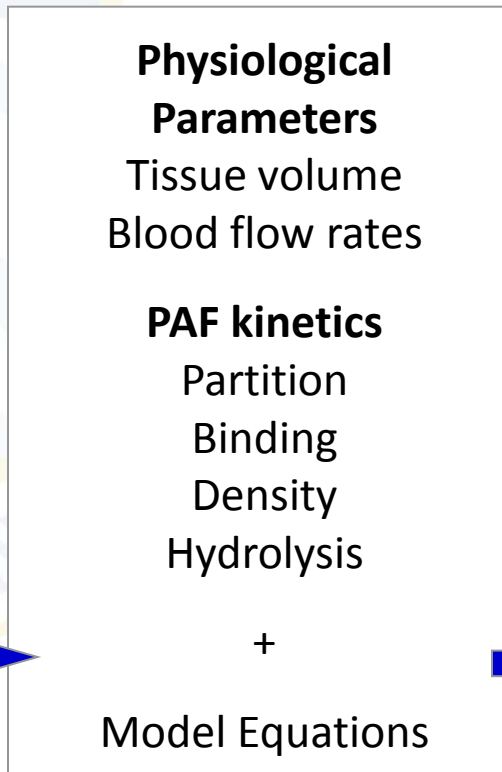
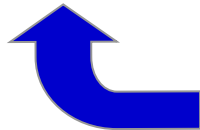




Iterative approach to modeling

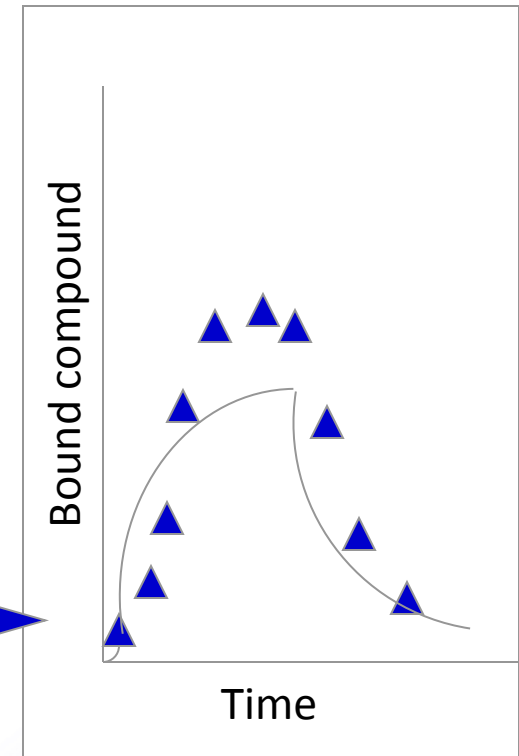


Define Realistic Model



**Collect Data
For Each Species**

Refine Model



Make Predictions





Platelet-activating factor (PAF)

- Autacoid binds to specific PAF receptor sites
 - Affinity and density is dependent on cell, tissue, and species
- Much data in literature focus on characterization of PAF binding to receptors on platelets
- While PAF effects are universal, platelet sensitivity towards PAF receptor varies among species
- **Species-dependent difference at receptor level of platelets**

Increasing platelet PAF receptor density

rat
mouse

Rhesus,
cebus
apella
primate

human
baboon
canine

guinea pig
rabbit





Platelet sensitivity is correlated to in vivo responses to PAF



Increasing platelet sensitivity

mouse

rat

reduced/absent

bronchoconstrictive responses

hemoconcentration at higher doses

dog

primate

human

guinea pig

rabbit

bronchoconstriction

hemoconcentration





Platelet sensitivity is correlated to *in vivo* responses to PAF



We can extrapolate *in vitro* cell line data to whole tissues, and from there to whole animals with PBPK models

- C16-PAF shows slightly higher, but statistically insignificant difference in potency between human lung and platelets
 - **Similarity between PAF receptors in human platelets and lung tissue**
- ***In vitro* platelet binding data were used in our models at tissue level when tissue-specific data were unavailable**



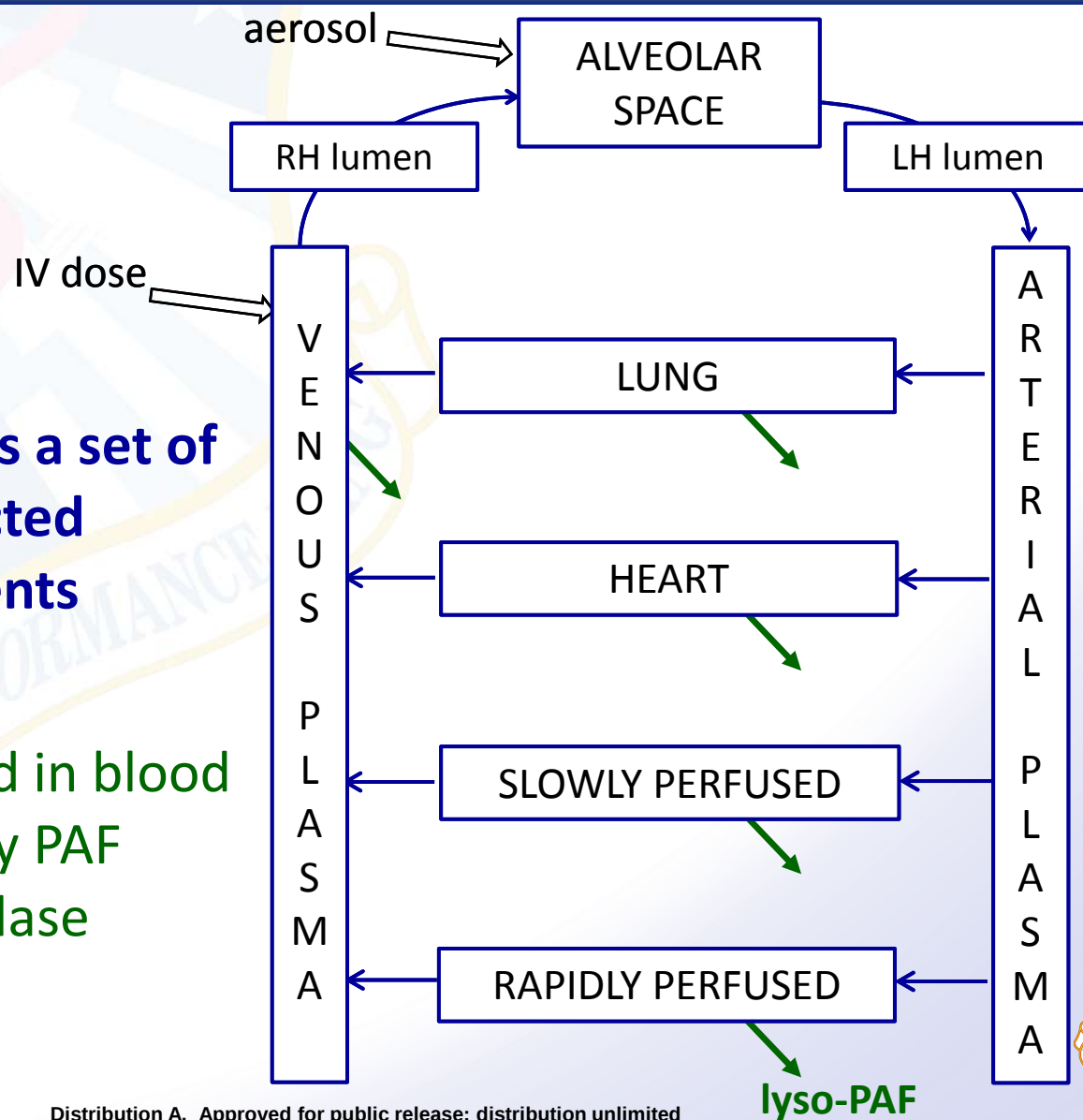


INITIAL PBPK model schematic for endogenous PAF



**Describes body as a set of
interconnected
compartments**

**PAF is metabolized in blood
and tissues by PAF
acetylhydrolase**

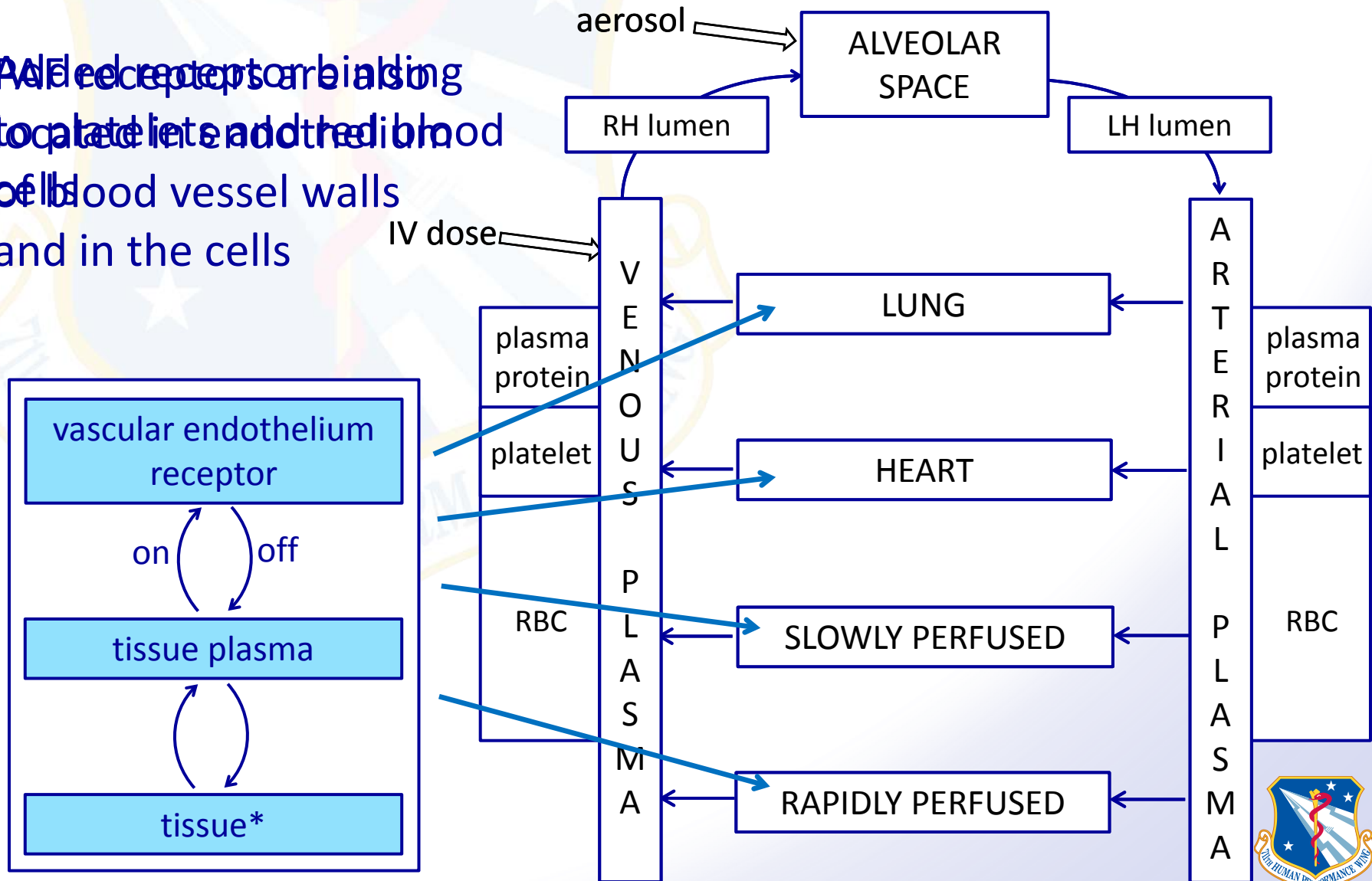




IMPROVED PBPK model schematic for endogenous PAF



AF receptors are binding to platelets and the blood of blood vessel walls and in the cells



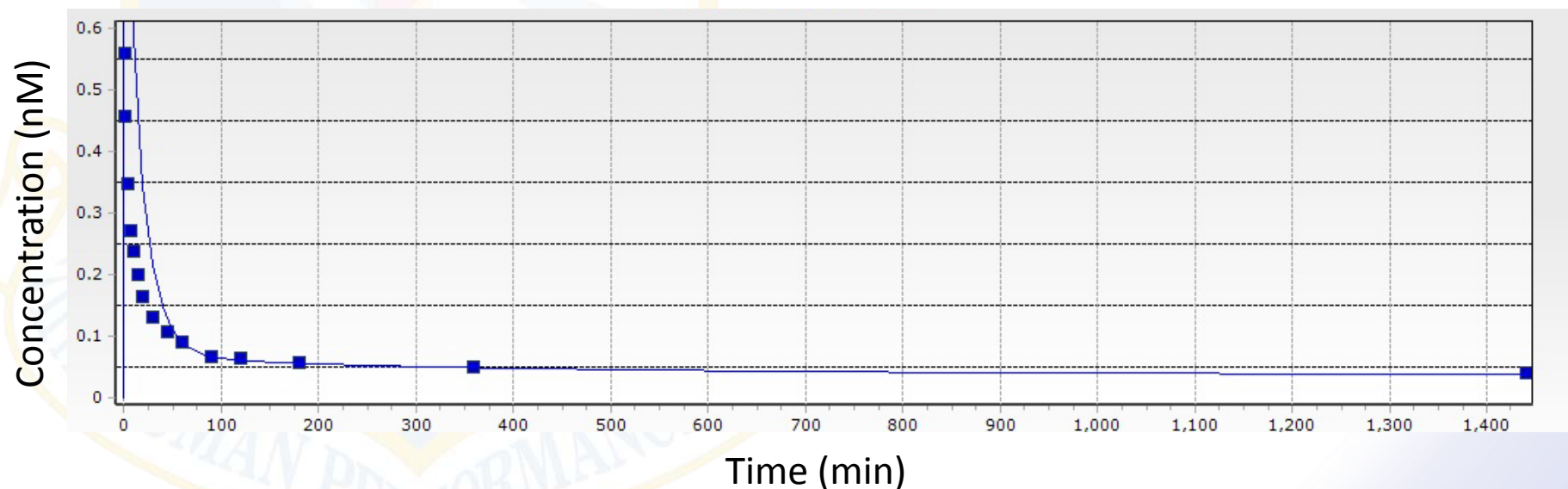


The resulting integrated model is used to simulate pharmacokinetics of PAF after intravenous exposure...





Pharmacokinetics of PAF (1.25 $\mu\text{g/kg}$ IV)



Simulation of [^3H]PAF concentration in venous plasma in rabbit following a 1.25 $\mu\text{g/kg}$ IV exposure compared to data

- This simulation compared with kinetic data show that the model is capable of accurately simulating the experimental data

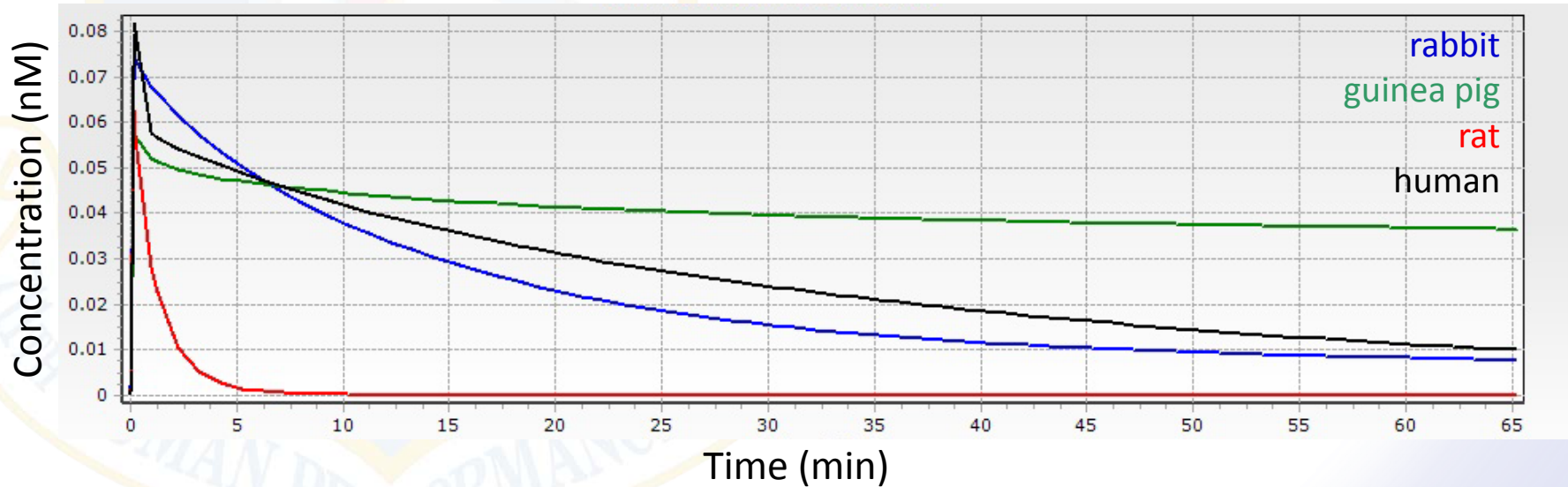
Experimental data: Lartigue-Mattei *et al.*, 1994

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Pharmacokinetics of PAF (1.25 $\mu\text{g/kg IV}$)



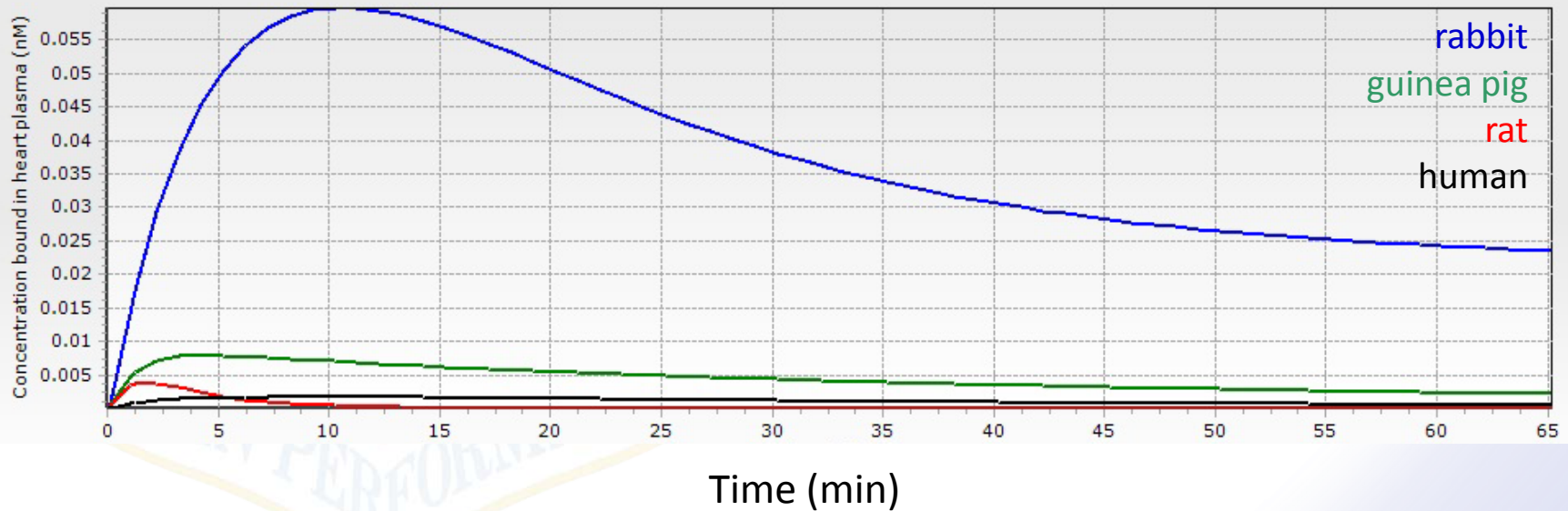
Data show a link between differential sensitivity to LPS and PAF among species and PAF kinetic parameters

- Higher plasma PAF in guinea pig corresponds to its increased susceptibility to LPS





Pharmacokinetics of PAF (1.25 $\mu\text{g}/\text{kg}$ IV)



Simulations demonstrate that bound PAF in heart plasma is higher in rabbit than rat

- Possibly explains why it is more toxic in rabbit than rat



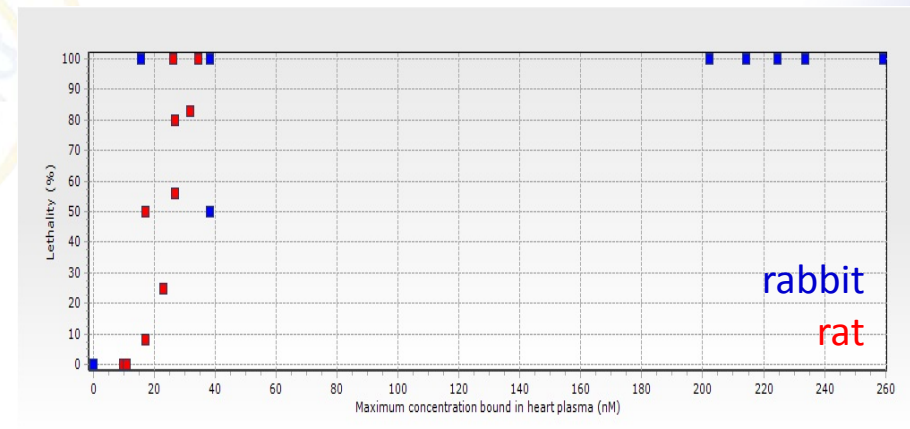
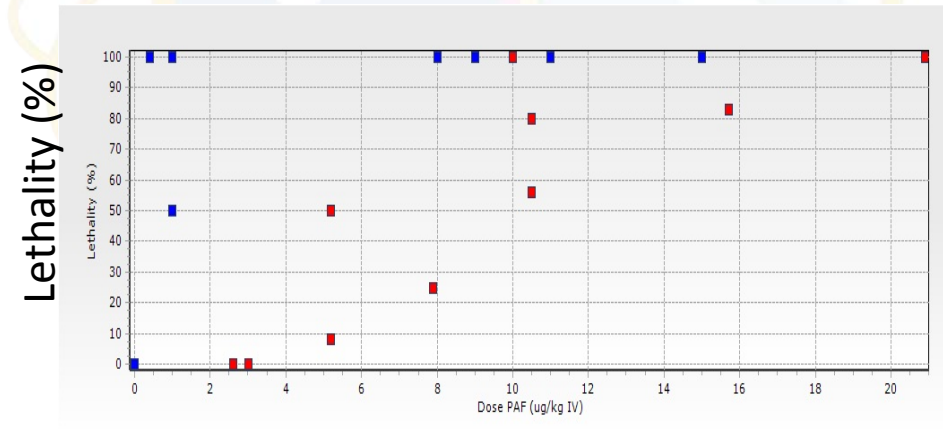


Model predictions of external LD₅₀



- Various model outputs were examined for correlation with observed signs of infection and lethality in an attempt to identify the most appropriate dose metrics for predicting adverse effects

Lethality dose response curve



Peak concentration of bound compound in heart plasma

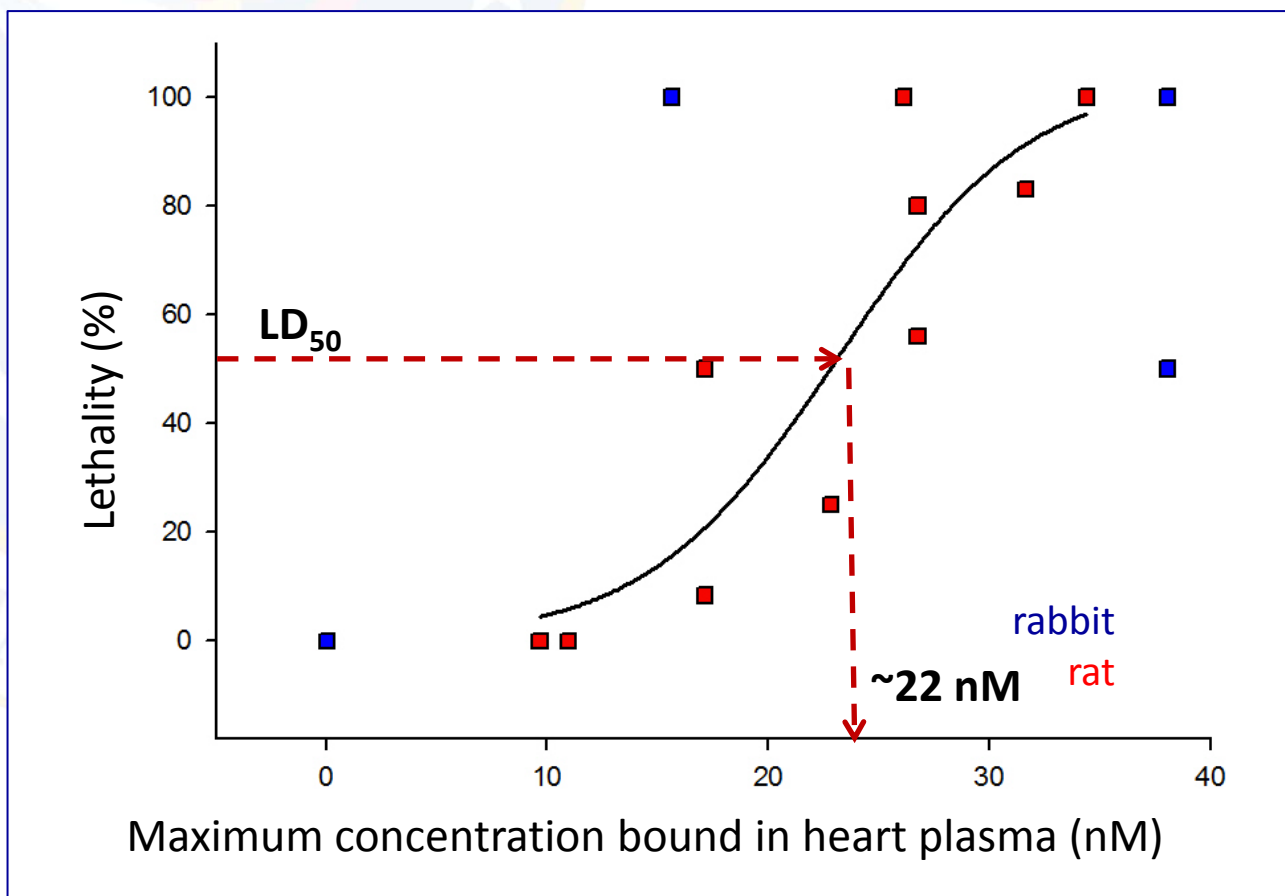
Experimental data: Lefer *et al.*, 1984, Tanaka *et al.*, 1983

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Lethality vs. internal dose metric



Regression of lethality on the internal dose metric results in an internal LD_{50} of $\sim 22 \text{ nM}$ peak bound in heart plasma





Human lethality prediction

- Running the human model shows that the human external dose required to achieve this peak is 900 $\mu\text{g}/\text{kg}$ IV
 - Humans should exhibit intermediate clearance/binding, consistent with intermediate toxicity of PAF in humans

	Rabbit	Human	Rat
External LD50 ($\mu\text{g}/\text{kg}$ IV)	0.57	900	7.5

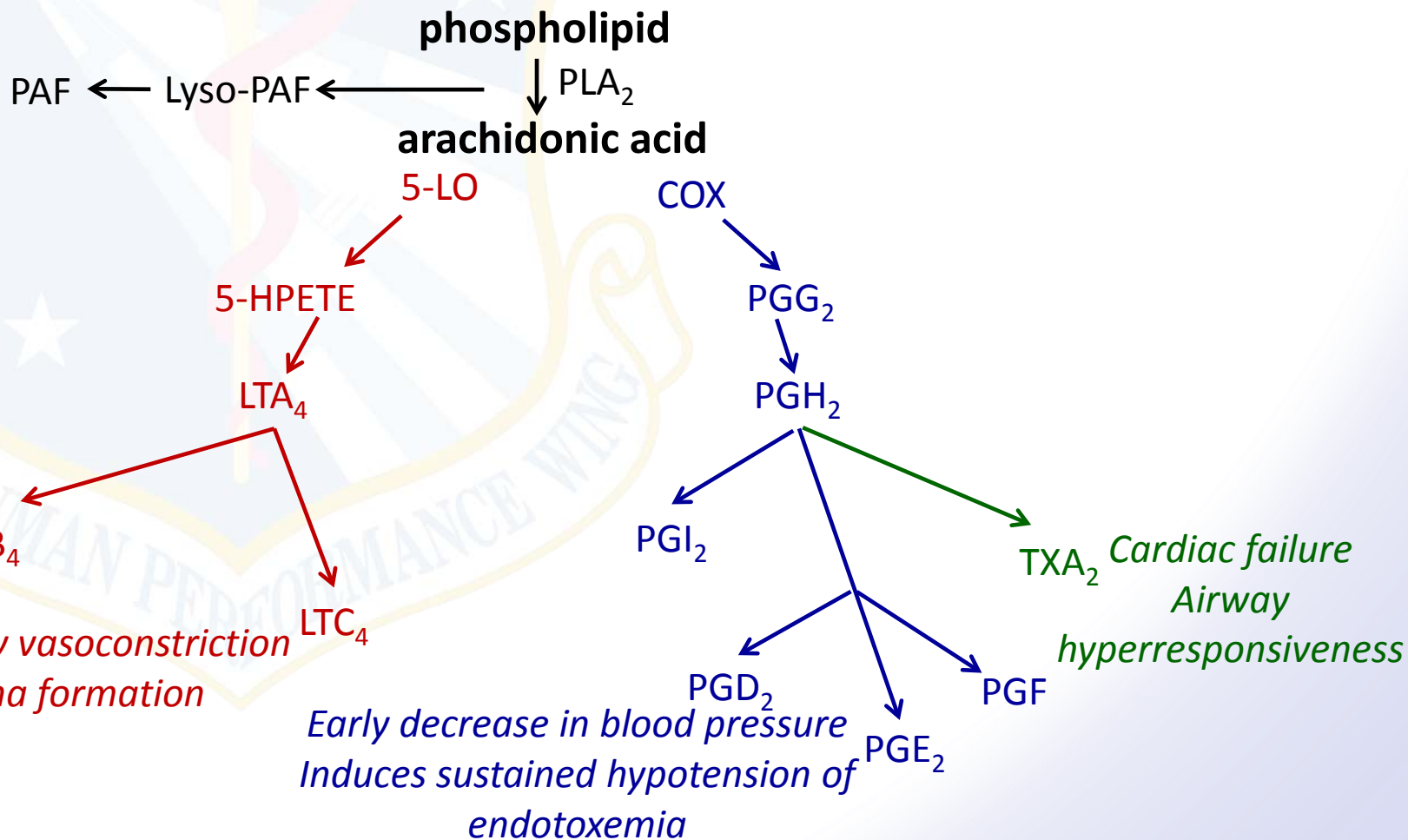
- Wrong dose metric?
- Dynamic differences downstream from receptor binding?
- Future work – *in vitro* experiments to verify value





Arachidonic acid (AA) cascade

Species differences



Interspecies differences in eicosanoid involvement highlight the need to consider the AA cascade in interpreting toxicity data





Future work

- Additional comparisons with kinetic and toxicity data in literature
- Confirmation of human parameter values used in human lethality prediction
- Addition of other exposure routes in order to simulate realistic human exposure scenarios (inhalation)





Adapt model to antagonists

PAF antagonists protect against injurious effects of LPS

- GB inhibits PAF-induced platelet aggregation and attenuates airway vascular permeability, hypotension, and lethality, and also AA accumulation
- Preincubation of platelets with GB diminished PAF binding in a specific and saturable manner

Model structures for PAF analogs and antagonists are similar

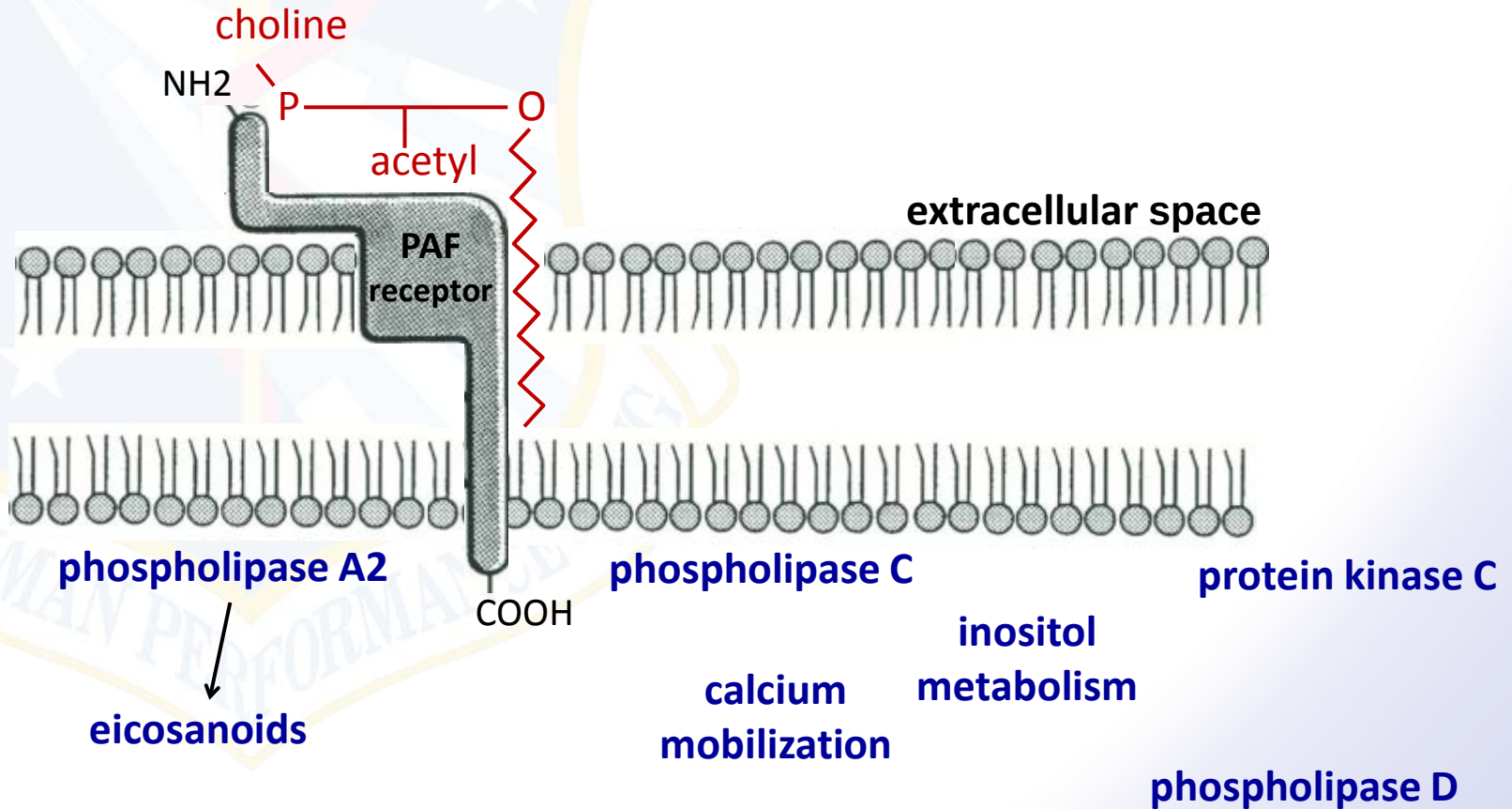
- Different physicochemical and biochemical parameter values

Modeling the kinetics of these antagonists and interactions/competition with PAF for the receptor would evaluate **therapeutic efficacy**





Model refinement



Mechanisms involved in bioactions of PAF are complex probably because receptor activates multiple signaling pathways





Summary

Model of PAF response interactions has been developed

- Describes distribution to tissues, hydrolysis, and binding to receptors in tissue vasculature following IV exposure
- Capable of accurate simulation of kinetic profiles in animals
- Peak PAF bound in heart is indicated as a dose metric for extrapolation of lethality across species

Model development is an iterative process





Acknowledgements



Biological Modeling Group

Tammie Covington
Emily Fleming
Dr. Jeff Gearhart
Eric Hack
Amanda Hanes
Tim Hansell
Capt. Mark Lisanby
Deirdre Mahle
Elaine Merrill
Lee Poeppelman
Dr. Peter Robinson
Chris Ruark
Teri Sterner
Dr. Kyung Yu

Public affairs approval

WPAFB Clearance for Public Release:
2012
Case Number

This work received funding from the
Defense Threat Reduction Agency
(2.G0002_09_AHB_C)





References

- **Chen WD**, Liang Y, Xie L, Lu T, Liu XD, Wang GJ. 2007. Pharmacokinetics of the ginkgo B following intravenous administration of ginkgo B emulsion in rats. *Biol Pharm Bull.* 30(1): 1-5.
- **Lartigue-Mattei C**, Godeneche D, Chabard JL, Petit J, Berger JA. 1984. Pharmacokinetic study of 3H-labeled PAF-acether . II. Comparison with 3H-labeled lyso-PAF-acether after intravenous administration in the rabbit and protein binding. *Agents Actions.* 15(5-6): 643-8.
- **Lefer AM**, Muller HF, Smith JB. 1984. Pathophysiological mechanisms of sudden death induced by platelet activating factor. *Br J Pharmacol.* 83(1): 125-30.
- **Tanaka S**, Kasuya Y, Masuda Y, Shigenobu K. 1983. Studies on the hypotensive effects of platelet activating factor (PAF, 1-O-alkyl-2-acetyl-sn-glyceryl-3-phosphorylcholine) in rats, guinea pigs, rabbits, and dogs. *J Pharmacobiodyn.* 6(11): 866-73.

