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Award Number: W81XWH-08-1-0436

TITLE: Production and Characterization of a Novel OX40 Ligand for Clinical Use

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REPORT DATE: August 2012

TYPE OF REPORT: Final

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

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REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
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1. REPORT DATE August 2012		2. REPORT TYPE Final		3. DATES COVERED 15 July 2008 – 14 July 2012	
4. TITLE AND SUBTITLE Production and Characterization of a Novel OX40 Ligand for Clinical Use				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-08-1-0436	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Andrew D. Weinberg, Ph.D. E-Mail: andrew.weinberg@providence.org				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Providence Portland Medical Center Portland, OR 97213				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT The goals of the second year/phase of the DOD funded research, PC073499, have been accomplished. We produced two human versions of the OX40 ligand:trimer:Ig protein. The two protein constructs have different human trimer domains (TRAF2 and Matrilin-4), which confer differing folding patterns within the two-dimension structure of the protein. These two fully human OX40L:Ig fusion proteins were administered to non-human primates to test for in vivo activity. The OX40L:Ig fusion proteins showed potent biologic activity in vivo, but it was clear that the one with the TRAF2 trimer domain was superior. Therefore we are moving forward with OX40L:Ig fusion protein with the TRAF2 trimer domain with the ultimate hope of taking it to clinical trials in patients with prostate cancer.					
15. SUBJECT TERMS T Cells, Co-stimulation, OX40, tumor immunotherapy					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT UU	18. NUMBER OF PAGES 12	19a. NAME OF RESPONSIBLE PERSON USAMRMC
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code)

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PC073499 – Production and Characterization of a Novel OX40 Ligand for Clinical Use.

PI -Andrew Weinberg, PhD

INTRODUCTION: Cancer cells have evolved to evade immune-mediated destruction through several documented mechanisms. Our group has developed a technique to enhance immune function in tumor-bearing hosts through the use of OX40 agonists, which can lead to regression of tumors of various histologies, including prostate cancer (1). In particular, we have produced a human OX40 agonist, termed OX40L:ILZ:Ig that has potent biologic function in vitro and is produced in large quantities by tissue culture cells. The ILZ portion of the chimeric protein was initially a trimerization domain obtained from a yeast sequence. Our goal in the first year of funding was to fully humanize this molecule by incorporating human trimerization domains to replace the yeast sequence, thus lowering the probability of immune-mediated recognition of this recombinant protein by treated cancer patients. This goal was accomplished with two human trimerization domains from the TRAF2 and Matrilin-4 proteins. Now that a fully functional human OX40 ligand protein was developed we proceeded with production and purification of the protein so that it could be tested for in vivo biologic activity in non-human primates. In the second year of funding we found that the biologic activity of the fully human OX40L:Ig protein was as potent as another OX40 agonist Ab that we currently have administered to patients. In the third year we produced a cell line that is GMP compliant and this cell line will be used to produce this protein for toxicity studies and clinical trials. The future goals beyond this funding period is to produce clinical grade human OX40L:ILZ:Ig to test in clinical trials for patients suffering from prostate cancer.

BODY: The first year of funding was spent on perfecting the soluble OX40 ligand molecule. In particular, we proposed to swap the yeast ILZ domain with known human trimerization domains. After modeling the different known human trimerization domains we made a decision to test two types: 1) a non-covalent trimerization domain (from TRAF2 sequence) and 2) a disulfide linked covalent trimerization domain (from matrilin-4 sequence). TRAF2 is an adapter protein that directly associates with the OX40 cytoplasmic tail and ultimately is involved with transmitting a downstream signal to the nucleus. It has been shown that OX40 and OX40 ligand form a trimer at the cell surface and the associated TRAF protein(s) form a trimer within the cytoplasm essentially forming a stacked three protein trimer complex. Hence we hypothesized that the TRAF2 trimer domain might make a perfect spatial fit between the OX40L extracellular domain and the Ig tail. There are only a few of the human trimerization domains that form natural disulfide bonds and the matrilin-4 protein is one of them. We reasoned that a covalently bonded trimerization domain might offer greater stability in vivo and hence might have increased biologic activity when compared to a non-covalent trimer domain. Therefore we produced both OX40L constructs and expressed them in 293 cells as secreted proteins. Protein G column chromatography was used to purify the proteins to >95% homogeneity and we compared their size by native gel electrophoresis. Prior to these experiments we published (2) that our initial recombinant protein, OX40L:ILZ:Ig, containing the yeast trimerization domain folded predominantly as a hexamer as determined by column chromatography. It was clear that the TRAF2 and matrilin-4 trimerization domains gave similar protein products (3 bands) but the matrilin-4

protein appeared less homogenous when compared to the TRAF2 construct. The matrilin-4 domain clearly had more of a higher and lower MW protein, which may or may not correspond to increased biologic activity. The initial bioassay results showed that both protein constructs had potent biologic activity in T cell stimulation assays. While it is hard to pick a clear cut “winner” based on our in vitro assays, both of these proteins were tested side by side in a non-human primate pilot study performed in year 2.

Our initial Statement of Work did not include the non-human primate pilot study discussed above, however in November 2009 we sent a revised statement of work that included an in vivo pilot study and shortly thereafter it was approved. These fully human OX40L proteins needed to be tested in vivo prior to moving forward with making material for toxicity studies and a phase I clinical trial. The human OX40L:Ig proteins do not bind to mouse or rat or other species and therefore had to be tested in non-human primates. The pilot study tested the fully human OX40L:Ig agonists and compared them to the murine anti-human OX40 Ab that has just completed phase I testing in cancer patients. Mouse Ig was used as a negative control in these experiments (see Fig 1 for experimental design). Based on our clinical trial experience and previous OX40 agonist monkey toxicity data (3) we chose a 1 mg/kg dose for all agents and this was administered to four monkeys per group. In Fig 1 we show a schematic of the experimental design, with the OX40-specific agents delivered on days 0, 2, and 4. The monkeys were injected with tetanus prior to the OX40 agonist infusion and pharmacokinetic points were taken pre- and 30 minutes post-infusion for all three infusions. Blood was drawn at multiple time points after infusion (Fig 1) in order to ascertain changes in T cell proliferation through the Ki-67 marker (4) and also assess whether these new OX40 agonists would increase an Ab response to tetanus.

The pharmacokinetics of all three OX40 agonists was assessed as shown in Fig 2. Prior to the monkey study we knew that the OX40L:Ig constructs were shorter lived (from previous mouse experiments) and we found the same was true when they were injected into monkeys. The serum levels of both the TF2 and MT4 constructs increased 30 minutes after infusion and then two days later were barely detectable. This was true for all three infusions. We also found that the peak level of OX40L in the serum was about 5-fold less than the peak OX40 mAb levels. It was also clear that the OX40 mAb lasted for a greater duration, as detectable levels were found 3 days after the last infusion.

Despite the short-lived pharmacokinetics, the OX40L:Ig TF2 fusion protein produced similar levels of biologic activity in the monkeys when compared to the OX40 Ab (CD134 mAb), as assessed by Ki-67 increases over time in peripheral blood T cells and increases in serum tetanus Ab titers (Figs 3-5). Figure 3 depicts a representative monkey from groups A, C, and D showing raw flow cytometry-based data, which were gated on CD4⁺/CD95⁺ T cells and assessing Ki-67 vs CD28 after OX40 agonist stimulation. Similar to what was observed with the mouse anti-human OX40 agonist in our phase I clinical trial (manuscript submitted) there was an increase in Ki-67 staining starting 7 days after the initial OX40 agonist infusion. This increase peaked 14 days after the initial infusion (Figs 2 & 3) and declined thereafter. We performed statistical evaluation at the day 14 time-point and found that all three test groups were significantly increased compared to the negative control (Fig 3B). The majority of the ex vivo Ki-67 examination showed that the OX40L:Ig TF2 protein out performed the MT4 construct in vivo. The TF2 OX40L:Ig construct also showed less molecular heterogeneity when compared to the MT4 construct therefore we picked the OX40L:Ig TF2 protein as our “lead” agent to move forward with. We next examined Ki-67 expression within the central memory CD4 T cell population (as defined by CD28⁺/CCR5⁻ (5)) and found that the OX40L TF2 protein showed a greater increase when compared to the OX40L MT4 protein and

the anti-OX40 Ab at days 7 and 11 post-infusion (Fig 3C). This data suggested that the OX40L:Ig protein may have immune stimulatory properties that are indeed different and may be superior than the mAb in vivo. It should be noted that others have postulated that increases in central memory T cell populations are a good indicator of tumor rejection in vivo (6). We next examined increases in CD8 memory T cell Ki-67 expression. The anti-OX40 Ab showed the greatest increase in CD8 T cell proliferation (Fig 4A). When the statistics were analyzed on day 14 only the OX40L:Ig TF2 protein and the CD134 mAb showed a significant increase in CD8 memory T cell Ki-67 expression when compared to the mIgG control (Fig 4B). Finally, we assessed a tetanus-specific Ab response in the four groups of treated monkeys. We previously found that monkeys treated with an OX40 Ab showed increased titers to immunizing Ags (3). We found that monkeys treated with either anti-OX40 or either OX40L:Ig fusion protein showed similar increases in serum Abs to tetanus. Figure 5 shows tetanus Ab titers for a representative monkey in the OX40L:Ig TF2, CD134 mab, and mouse IgG groups (A, B, and C respectively). As summarized in Fig 5D the average tetanus serum Ab titers were increased approximately 5-fold in both the OX40L:Ig TF2 and CD134 mab groups when compared to the control group (mouse IgG). Thus, this data confirms that the OX40L:Ig TF2 protein had similar levels of biologic activity when compared to the anti-OX40 Ab and in some cases greater activity (CD4 central memory proliferation).

Based on the data shown above we picked the OX40L:Ig TF2 as our lead agent and started cell line optimization in the third year of this grant. In order to produce large quantities for clinical trials it was imperative that we developed a cell line that would allow for increased production of this protein. Therefore we contracted with WuXi Apptec to develop a cell line/clone that would produce large amounts of the human OX40L:Ig TF2 protein. We used the OptiVEC vector system for CHO cells and WuXi cloned our sequence into this vector. CHO cells were transfected with the OX40L:Ig TF2 vector and chemical selection for high secreting cells was performed as outlined in Fig 6. After selection the cells were cloned in soft agar and clones that were positive for OX40L:Ig secretion were visualized by an anti-huIg-FITC Ab as depicted in Fig 7. Several 100 clones were picked and further evaluated for protein production. The top four producing clones were grown in 1 liter bioreactors and compared for duration and production of protein on a per cell basis Fig 8. Upon large scale one-liter production, one clone (755-19) was superior to the rest as shown in Fig 8. Hence this clone was chosen to produce our working cell bank and will ultimately be used for GMP production of this protein for future clinical trials.

The final part of the project that fell under the umbrella of this application was to produce a GMP master cell bank. Again this work was contracted out to WuXi Apptec, which has a GMP facility in Pennsylvania. The process to produce a cell bank compliant for GMP protein production is not trivial and involves multiple steps/ tests that are outlined in Fig 9. This work was completed in June of 2012 and we now have 200 vials that are being stored in a GMP compliant facility.

KEY RESEARCH ACCOMPLISHMENTS:

- We produced two novel forms of the human OX40 ligand:trimer:Ig protein that have completely human sequences.
- These proteins were injected into non-human primates and assessed for biologic activity.
- It is clear that the TRAF2 trimer and the Matrilin-4 trimer domains were able to produce biologically active OX40 ligand proteins in vivo, but the TRAF2 construct was superior.

- When the OX40L:Ig proteins were tested side by side with our mouse anti-OX40 antibody there was similar amount of in vivo activity, although the OX40L:Ig fusion protein did show enhanced activity in stimulating proliferation of central memory CD4 T cells.
- The results from the non-human primate pilot study were successful and have generated much enthusiasm for moving the OX40L:Ig TRAF2 protein forward for future clinical trials.
- Cell line development has been performed with a CHO cell that produces the OX40L:Ig TRAF2 protein and a high producing cell line was achieved.
- The cell line was made into a GMP cell bank that can be used to produce clinical grade protein.

REPORTABLE OUTCOMES:

We have yet to publish these results as they somewhat confirmed what we already knew with OX40 agonists in the context of stimulating the murine immune system. However, there were some interesting caveats like the enhanced proliferation of central memory CD4 T cells that have not been previously reported. In collaboration with the Oregon Primate Center we are performing some follow-up studies, where we could incorporate the new findings with the ones presented in this report for a novel manuscript. However, the main goal of this work was to make a cell bank with a potent form of the human OX40L:Ig fusion protein and that was accomplished.

CONCLUSION:

In summary, we have completed the third and final year of research. We performed testing of two human OX40L:Ig fusion proteins and generated sufficient quantities of both proteins and purified them to greater than 95% homogeneity. We compared these immune stimulatory proteins versus our mouse anti-human OX40 Ab that has already been tested in the clinic. The OX40L:Ig fusion proteins showed similar immune stimulatory properties as the mouse monoclonal OX40 Ab. However, it was clear that the OX40L:Ig fusion protein with the TRAF2 trimer domain had greater activity in vivo when compared to the MT4 trimer domain. Therefore we proceeded with further development of the OX40L:Ig TRAF2 trimer protein. In the final year of funding, a cell line was developed that produces large amounts of the OX40L:Ig protein and we were able to show that it had functional activity. This cell line was then made into a GMP cell bank, which ultimately will be used to produce protein for future clinical trials for patients with cancer.

In our phase I clinical trial with the mouse OX40 agonist Ab administered to stage IV cancer patients we have observed some tumor shrinkage in 7/30 patients, however we did not observe a complete response. Our hope is that the fully human OX40 agonists described herein will have increased biologic activity because they can be administered several times without eliciting a strong immune response, unlike the mouse anti-human OX40 Ab. Ultimately, upon completion of this project we will have important preclinical information that will help garner FDA approval of a fully human OX40 agonist to treat prostate cancer patients.

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APPENDICES: Not applicable

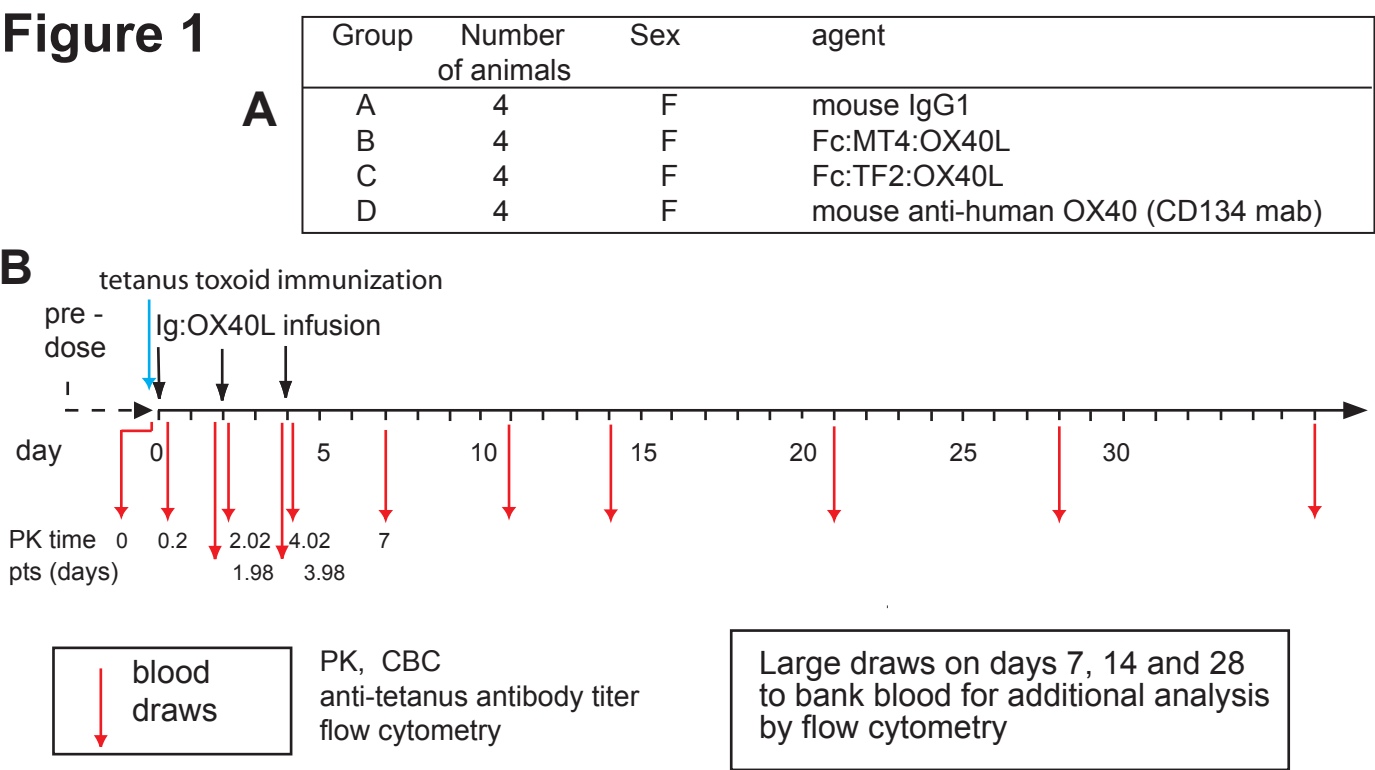


Figure 1. Schematic for testing the activity of recombinant OX40L in non-human primates. **A.** Schematic of treatment groups. Mouse IgG1 (MOPC21) was obtained from BioXpress. Fully human recombinant OX40L fusion protein containing the coiled-coil domain from either matrilin-4 (FcMT4OX40L) or from Traf 2 (FcTF2OX40L) were obtained by expression of these constructs in HEK 293 cells in a hollow-fiber bioreactor (FiberCell). Mouse anti-human OX40 was the same monoclonal antibody used in our phase I clinical trial. All reagents were prepared at 2.0 mg/ml in phosphate buffered saline. **B.** Rhesus monkeys were treated with OX40 agonists or controls on days 0, 2 and 4 by IV infusion at a dose of 1mg/kg. Blood samples were taken at the times indicated. For the analysis of pharmacokinetics, samples were taken 30 minutes before and 30 minutes after each infusion. Whole blood was used for phenotypic analysis by flow cytometry and for CBC. Plasma was prepared for PK analysis and for measuring antibody titers to tetanus toxoid.

Figure 2

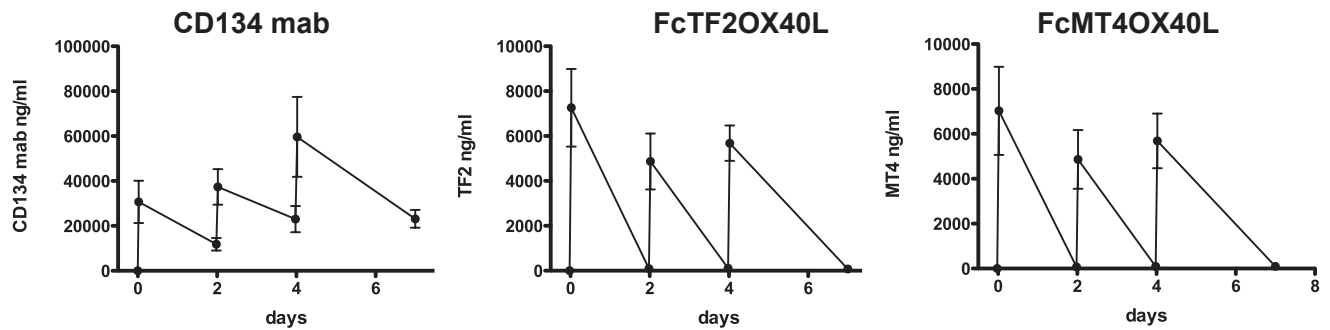


Figure 2. Pharmacokinetics of recombinant OX40L (FcTF2OX40L and Fc MT4OX40L) compared to OX40 agonist mouse monoclonal antibody (CD134 mab) in Rhesus monkeys. Blood samples were obtained according the scheme in figure 15, (30 minutes before and after the I.V. infusions on days 0, 2 and 4 and then on day 7). Values for individual animals were combined to obtain the group means at each time point. The concentrations of circulating FcTF2OX40L and CD134 mab were determined by ELISA. For FcTF2OX40L: anti-rat IgG (Fc γ) capture antibody - rat anti-human OX40L - serum samples - anti-human IgG Fc γ conjugated to HRP. For CD134 mab: anti-mouse IgG Fc γ - serum samples - anti-mouse IgG conjugated to HRP.

Figure 3

Total CD4 T cell Memory - gated through CD95/CD4

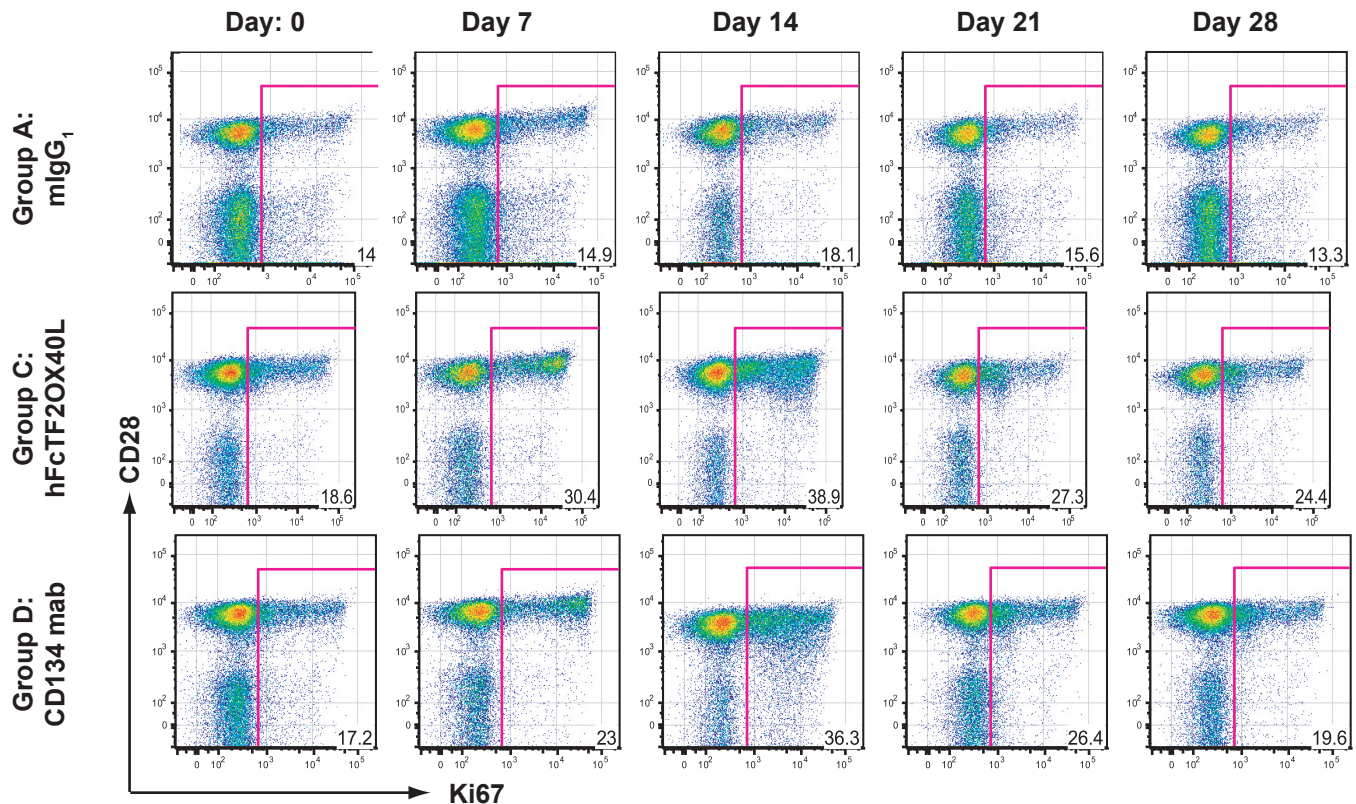


Figure 3. Proliferative response of CD4 T cell memory populations to OX40 engagement using Ki67 as a marker for T cell proliferation. Whole blood assays to assess T cell phenotypes by flow cytometry were performed at the times indicated. The lymphocytes were gated through CD3, CD4, and CD95. The data plots show CD28 vs Ki67 are shown. The number in the bottom right corner is the percentage of Ki67 positive cells, where an isotype control antibody was used to discriminate positive from negative events.

Figure 4

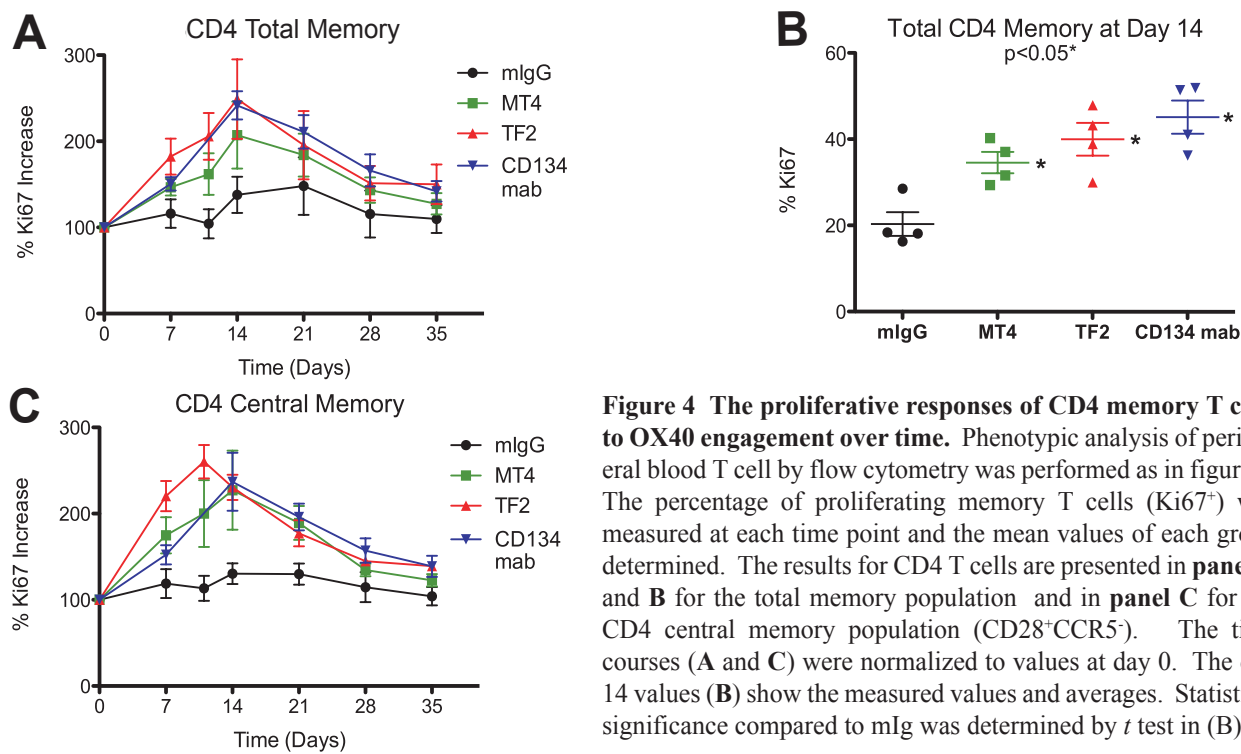


Figure 4 The proliferative responses of CD4 memory T cells to OX40 engagement over time. Phenotypic analysis of peripheral blood T cell by flow cytometry was performed as in figure 3. The percentage of proliferating memory T cells (Ki67⁺) was measured at each time point and the mean values of each group determined. The results for CD4 T cells are presented in **panel A** and **B** for the total memory population and in **panel C** for the CD4 central memory population (CD28⁺CCR5⁺). The time courses (**A** and **C**) were normalized to values at day 0. The day 14 values (**B**) show the measured values and averages. Statistical significance compared to mlg was determined by *t* test in (**B**).

Figure 5

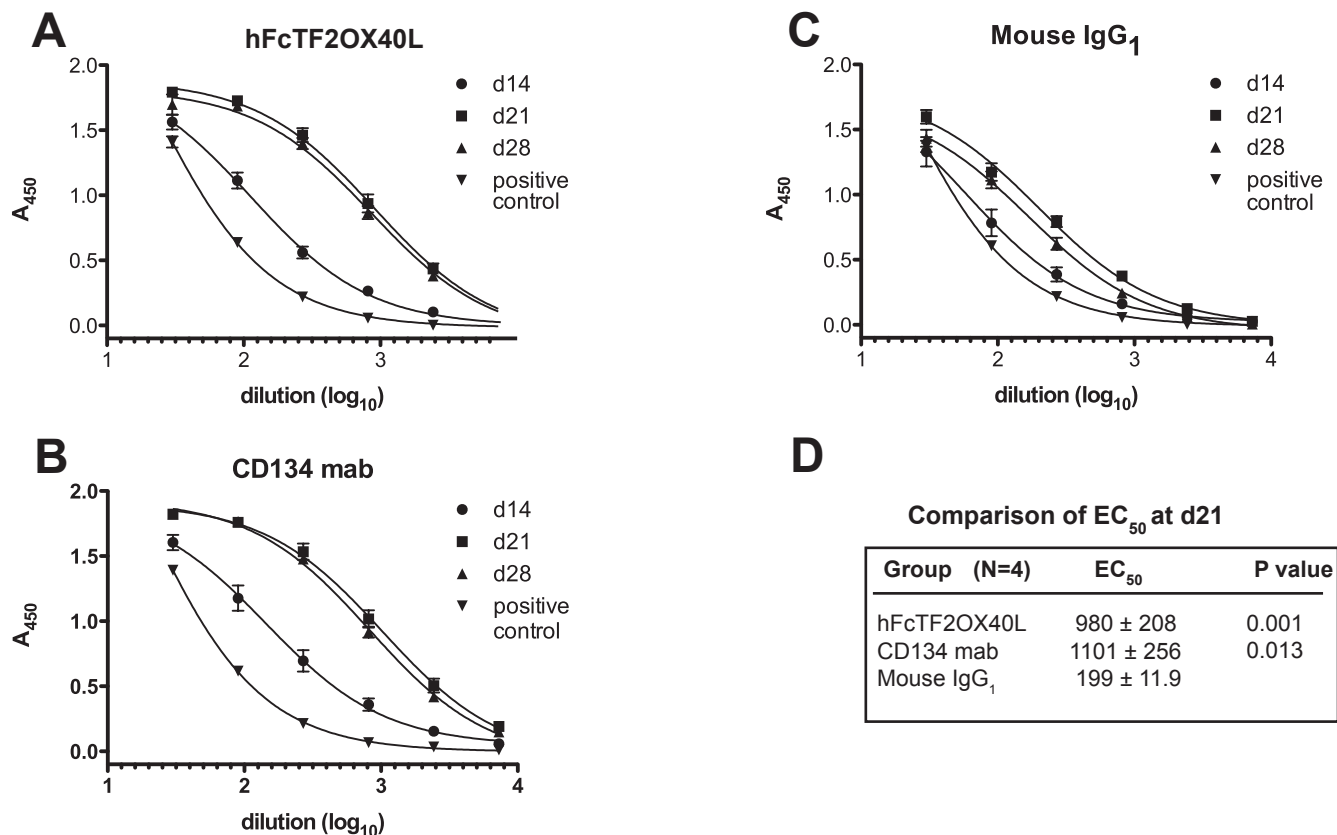


Figure 5. The effect of OX40 engagement on anti-tetanus toxoid antibody titers after vaccination. Plasma was prepared from blood samples taken at days 14, 21 and 28. Anti-tetanus titers were measured by ELISA using an HRP-conjugated secondary antibody that recognizes all classes of monkey antibodies (Rockland). The data for (**A**) OX40L:Ig, (**B**) CD134 mab, and (**C**) mouse IgG are the average curve fit to the data from all four monkey in each group. The titration data was fitted to a sigmoid dose response curve by the least squared method. For statistical comparison (**D**), the EC₅₀ for individual monkeys in each group was determined from the least squares fit. All four monkeys per group were averaged and these values evaluated by t-test for significance vs the mlg control group.

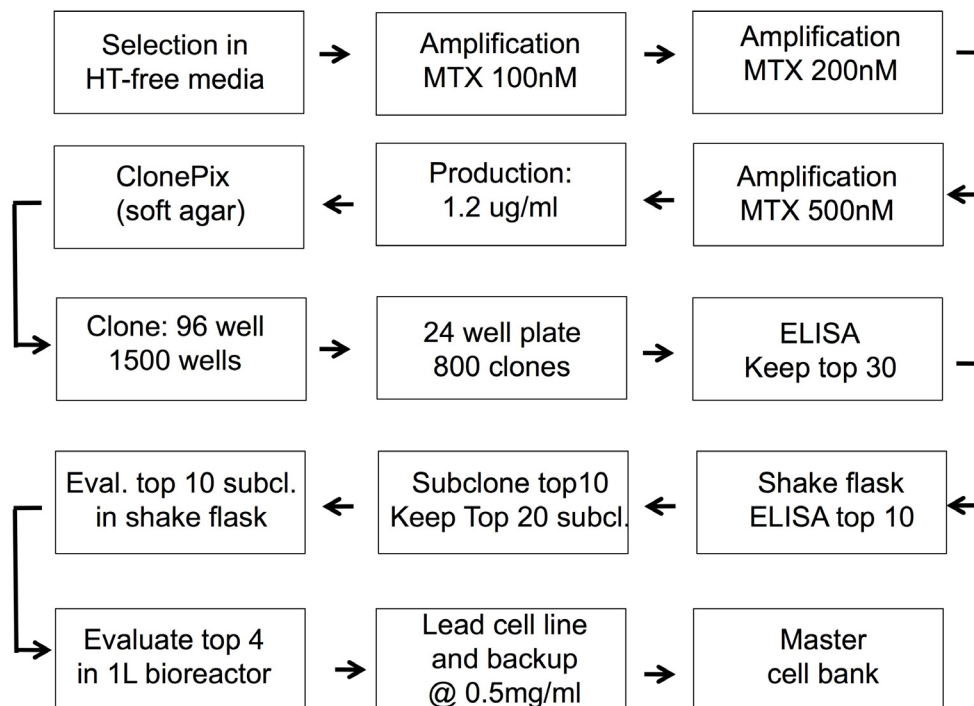
Figure 6

Figure 6. Schematic representation of the development of a Chinese Hamster Ovary (CHO) cell line expressing human OX40L:Ig. CHO cells were transfected with the vector expressing OX40L:Ig and Dihydrofolate Reductase enzyme (DHFR) on the same mRNA under the control of the CMV promoter. DHFR is required for thymidine biosynthesis by the cells. Transformants are selected in the absence of hypoxanthine and thymidine (HT) from the media so the cells are dependent on DHFR supplied by the vector. Methotrexate (MTX) is an inhibitor of DHFR. In the presence of MTX, CHO cells that have duplicated the DHFR (and therefore the OX40L :Ig gene) overcome the inhibition and increase OX40L:Ig expression. Successive increases of the MTX concentration results in further MTX/OX40L:Ig gene amplification.

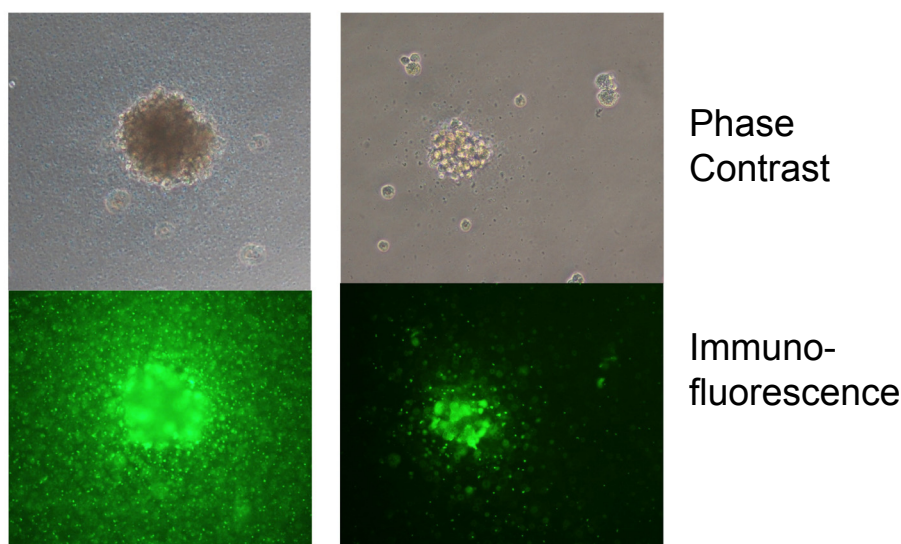
Figure 7**ClonePix example**

Figure 7. Selection of high-expressing amplified clones of CHO cells secreting OX40L:Ig. After amplification, OX40L:Ig-transfected CHO cells were diluted into semisolid agar growth medium to generate a two dimensional array. A FITC labeled anti-human IgG was applied to the cells to identify clones secreting OX40L:Ig. Typically the brighter fluorescence staining corresponds to higher levels of protein synthesis and secretion. The ClonePix instrument permits rapid screening of thousands of clones and recovery of clones for expansion and further analysis. Upper panels are phase contrast micrographs showing clones in soft agar and the lower panels depict the corresponding immunofluorescent labeling of these clones with FITC-conjugated anti-human IgG antibodies.

Figure 8 Summary of 1L Bioreactor Runs

Clone No.	Basal media	Feed	Temp(°C)	Peak CD 10E5 C/ml	IVCD Σ(10E5 C/ml/d)	Duration (d)	Titer (mg/l)	PCD (pg/cell/d)
592-6	OptiCHO	10% Feed B on Day3,7,10	37	55.68	173.35	7	N/A	N/A
755-19	OptiCHO	10% Feed B on Day3,7,10	37	55.49	459.95	13	539.45	11.73
755-1	OptiCHO	10% Feed B on Day3,7,10	37	42.16	397.76	14	N/A	N/A
845-19	OptiCHO	10% Feed B on Day3,7,10	37	59.9	533.21	14	75.17	1.41

Figure 8. Screening of best clones in 1L bioreactors. The top four clones based on cell growth and protein expression in shake flask culture were expanded to 1L fed-batch bioreactors to simulate large-scale protein production. Parameters monitored included Peak Cell Density (peak CD), Integrated Viable Cell Density (IVCD) measured as the viable cell density per day summed over the number of days of the run and finally the amount of OX40L:Ig produced per cell (PCD, picograms per cell per day).

Figure 9

Assessment of Authenticity and Species Identity

Post Cell-Bank Cell Growth for Testing.
Cell Growth for Assays.
Determination of Cellular Morphology and Growth Characteristics.
Cell Line Species Identity by Isoenzyme Electrophoresis.

Sterility and Mycoplasma

Biologics-Sterility Test-Immersion (USP/EP and 21 CFR610.12).
Biologics- B/F Immersion (USP/EP).
Mycoplasma Detection with MycoplasmaStasis (EP and USP).

General Virus Screens

In Vitro Assay for Detection of Adventitious Viral Contaminants.
In Vivo Assay for Viral Contaminants in Guinea Pigs, Adult Mice, Suckling Mice and Eggs.

Specific Virus Tests

Hamster Antibody Production Test.
Detection of MVM DNA by qPCR.

Retroviruses

Thin Section Electron Microscopy: Cell Morphology and Virus Detection.
Co-cultivation of Test Article with Mus dunni Cells and PG4 S+L.
Fluorescent Polymerase Chain Reaction (PCR)-based Reverse Transcription (F-PBRT) Assay.

Adventitious Viruses from Animal Derived Raw Material

Detection of Bovine Polyoma Virus by qPCR.
Detection of Bovine Viruses : Extended Screening for Non-bovine Cell Lines Grown in Bovine Serum or Products.

Figure 9. Testing performed for GMP Master Cell Bank of hOX40L:Ig clone 755-19 cell line. The Master Cell Bank produced for hOX40L:Ig clone 755-19 cell line was tested for cell growth and viability as well as for harboring known pathogens.