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Investigation of a simple visual system for flight control.

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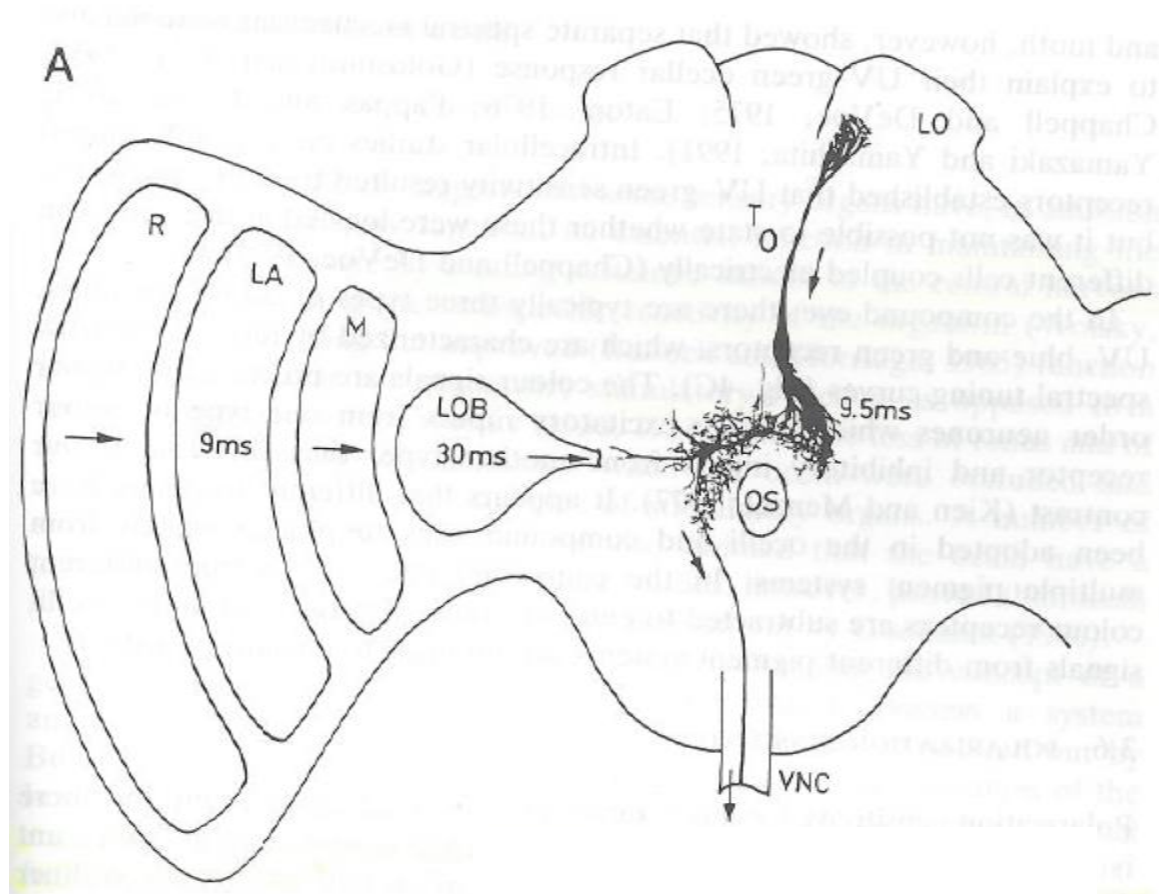
Hypothesis



Characterizing the physiology of ocellar systems and associated behaviors from a wide diversity of insects could ultimately lead to a more complete understanding of insect flight control.



Ocelli are faster than the compound eyes.



<u>Visual system</u>	<u>Latency of pathway</u>	<u>Minimum connections</u>
Compound eyes	25-35 ms	4 interneurons
Ocelli	9ms	1 interneuron



Bug Faces: Ocelli are anatomically diverse.



Mizunami, The diversity of insect ocellar systems, 1994



Identifying good candidates for study.



- Different sensor suites different behaviors
- Same sensor suites similar behaviors
- Different sensor suites similar behaviors
 - Different sensors measure/use same information? Therefore behavior requires that piece of information.
 - Using different information but getting same outcome? Behavior can be produced using different pieces of information.
- Same sensor suites different behaviors
 - Different ecological/environmental demands? Possible to produce the behavior under the right conditions.
 - Using same information differently? Sensors have been co-opted for different purpose.

*Statements assume that “same sensor” means measuring the same information.



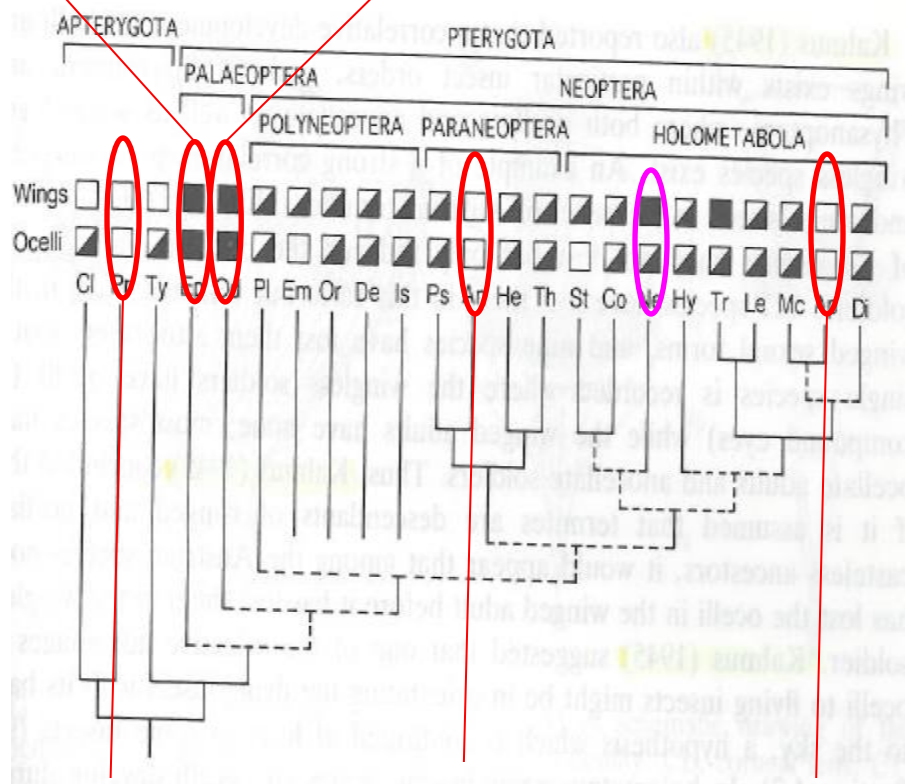
Correlative development of wings and ocelli.



Dragonflies and damselflies

mayflies

Insect orders



All have wings or ocelli



None have wings or ocelli



Some have wings or ocelli

proturans

lice

Fleas

Mizunami, The diversity of insect ocellar systems, 1994



Identifying good candidates for study.



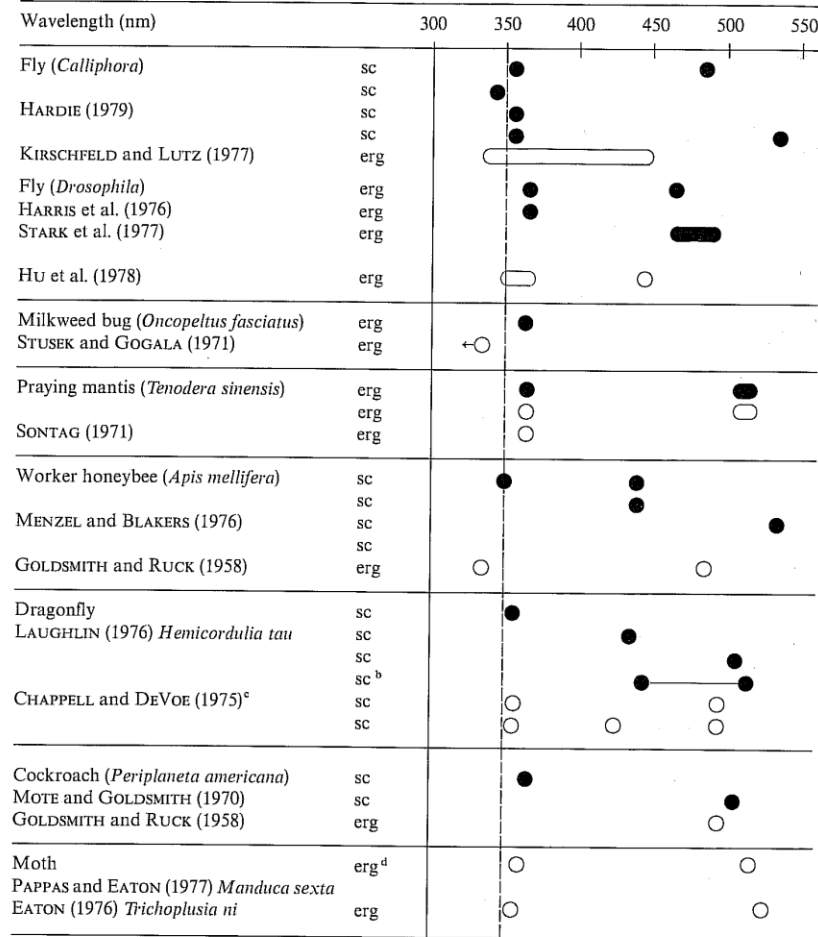
- **Different sensor suites similar behaviors**
 - Similar flight behavior in insects with different sensors (some have ocelli and some do not).
 - Dragonflies vs. nearctic owlflies
- **Same sensor suites different behaviors**
 - Very different flight, but all have the same sensor (all have ocelli).
 - Crane flies vs. robberflies vs. hoverflies



Properties of ocelli have not been widely investigated.



Table 5. Spectral sensitivity of ocellar and compound eye receptor cells ^a



^a Points along any single horizontal line represent spectral sensitivity maxima determined by either single cell intracellular recording (sc) or the electroretinogram (erg). ●, spectral sensitivity maxima for compound eye receptor cells; ○, spectral sensitivity maxima for ocellar receptor cells; ←, peak UV sensitivity possibly at shorter wavelength

^b Range of peak sensitivity for linked pigment cells

^c UV-Green in *Anax junius*, *Libellula pulchella* and *Aeschna tuberculifera*. UV-Blue-Green only in *A. junius*

^d Internal ocellus

Goodman, Organization and physiology of the insect dorsal ocellar system, 1981



Proposed experiments will characterize ocellar properties.



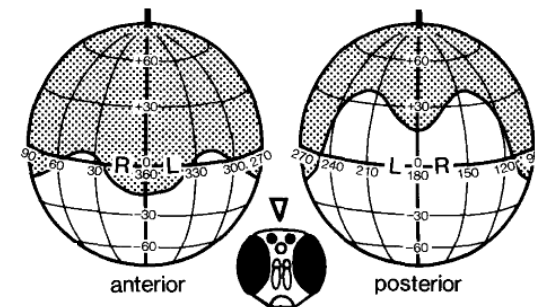
Measured properties:

- Spectral Sensitivity
- Flicker Fusion Frequency
- Field of View & directional sensitivity
- Focal Length and spatial resolution



Techniques:

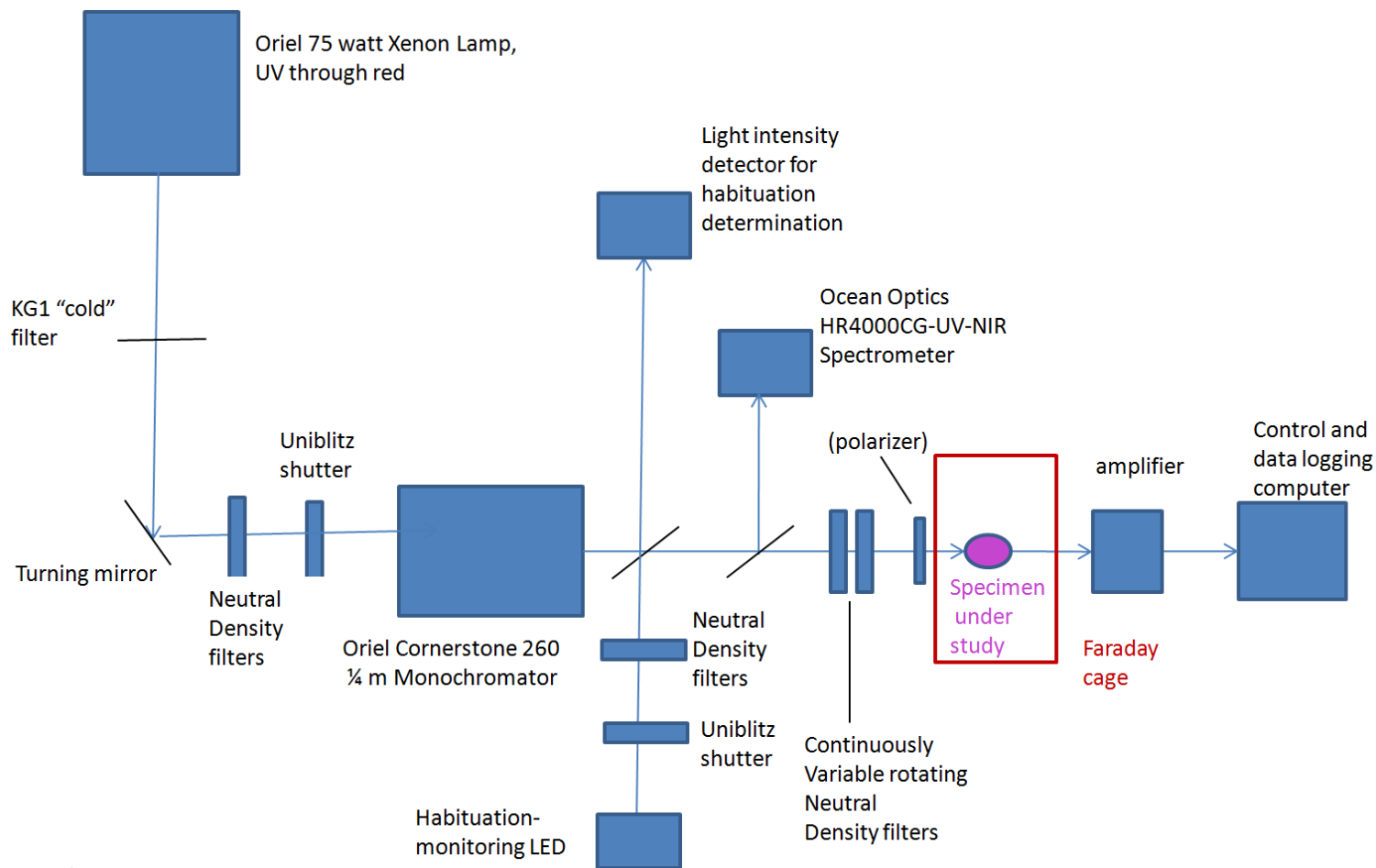
- Electroretinogram
- Extracellular from ocellar nerve
- Intracellular from L neurons



Schuppe and Hengstenber, Optical properties and functional role of the dorsal ocelli 1993

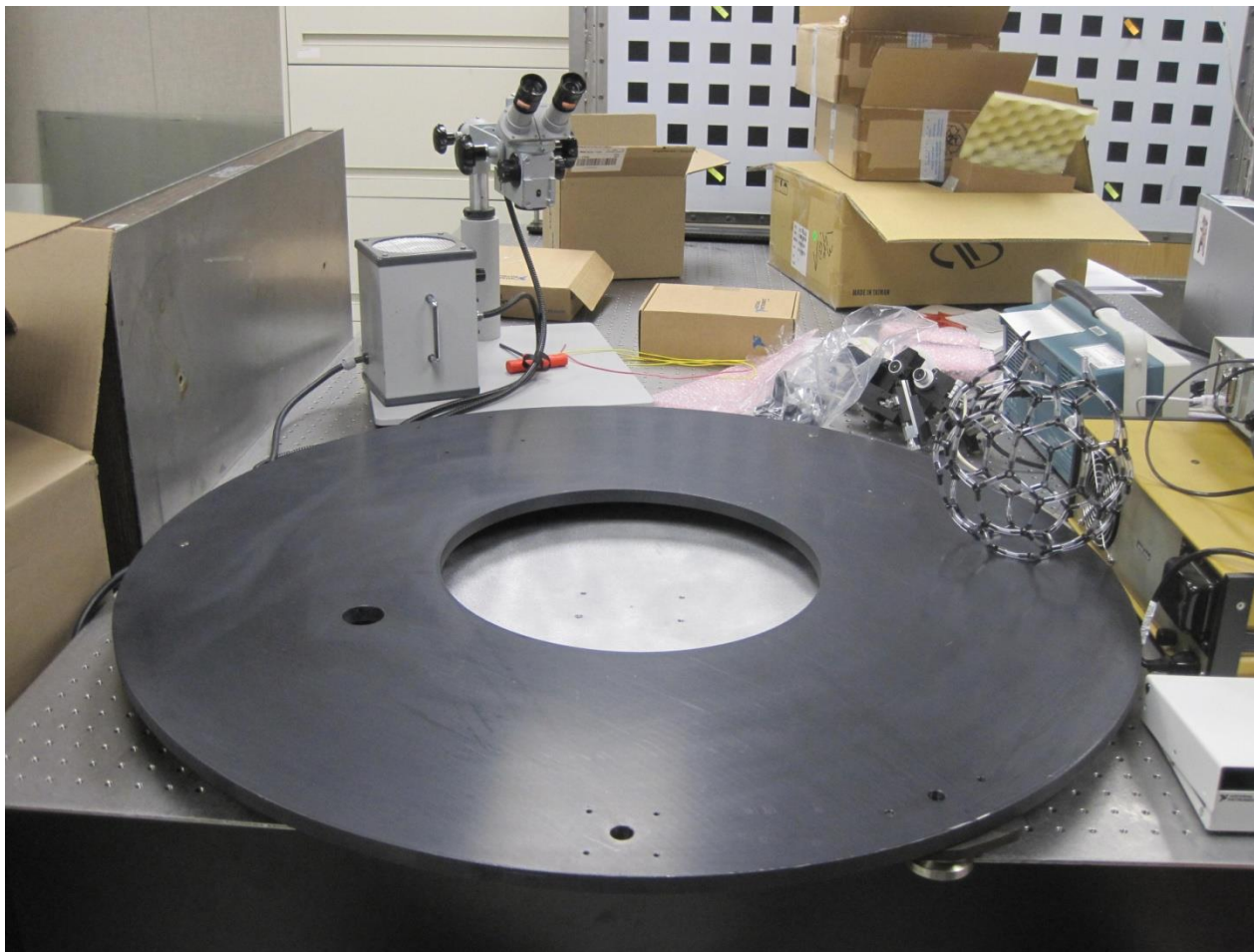


Test spectral responsivity, FFF



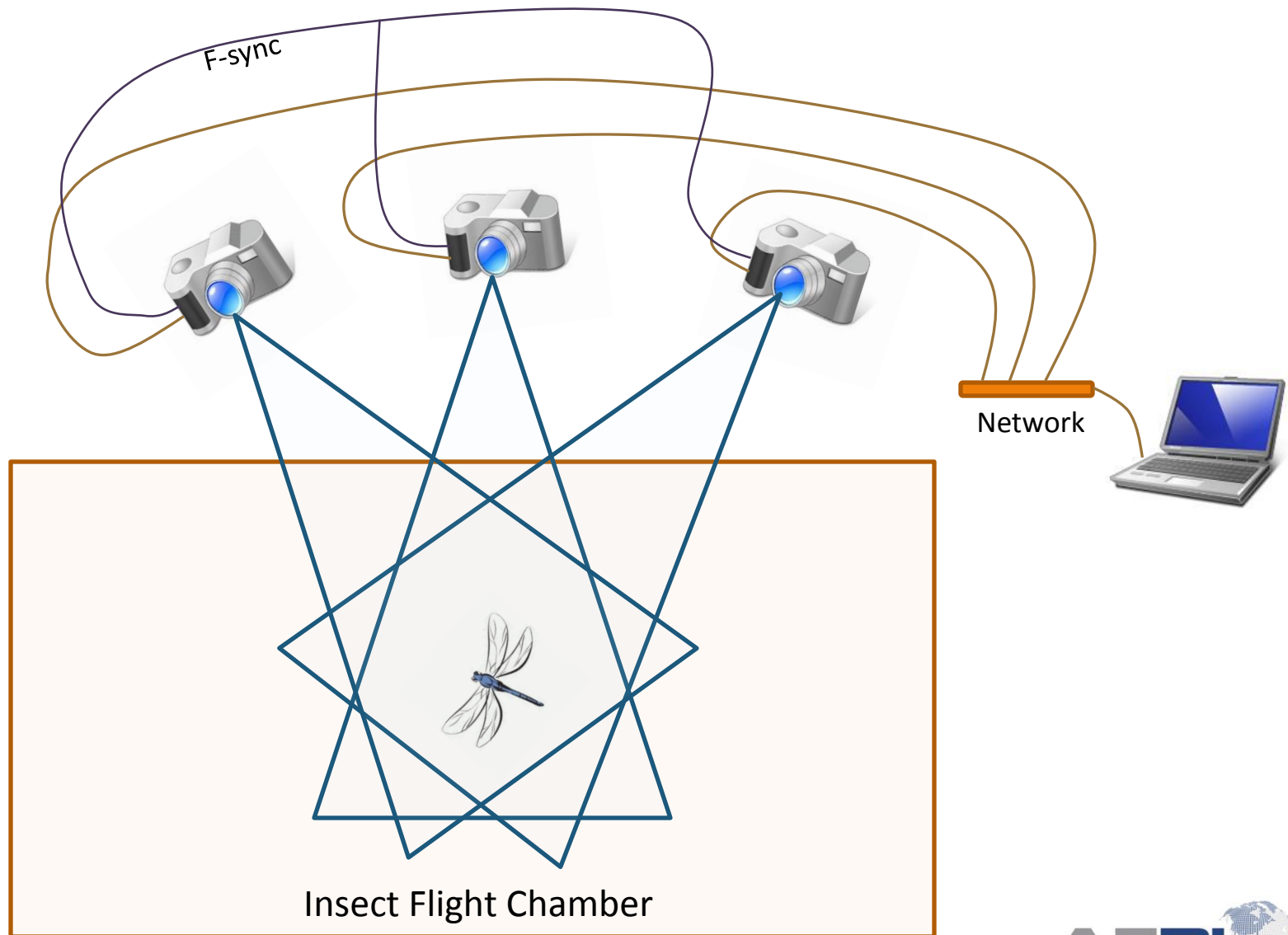


Field of view, directional sensitivity, spatial resolution.





Insect Flight setup, by David Forester.



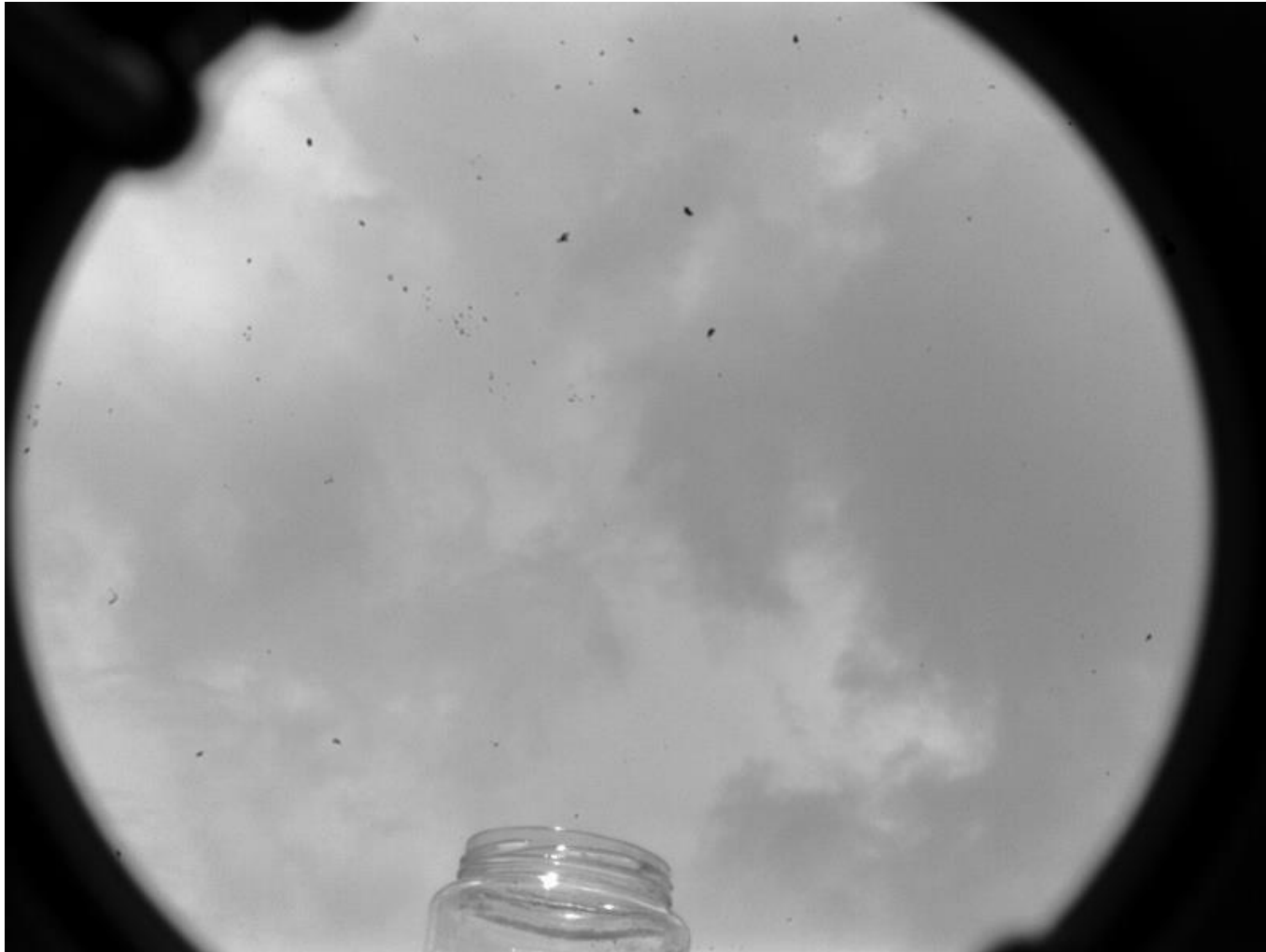


Similar behaving insects, different sensors.





Dragonfly in field





Dragonfly in field, recovers from perturbation.





Owlfly horizontal dorsal flight.





Owlfly vertical flight.





Owlfly vertical backward flight.





Owlfly upside down flight.





screenshot from the tracking software.



clicking_gui_n_cams

Three small thumbnail images are displayed at the top left of the main window:

- Thumbnail 1: A small, dark, cross-shaped object in the center of a light gray background.
- Thumbnail 2: A small, dark, cross-shaped object in the center of a light gray background, with a green border.
- Thumbnail 3: A small, dark, cross-shaped object in the center of a light gray background, with a blue border.

The main window displays a large grayscale image of a mosquito-like insect in flight, with a yellow dashed line indicating its trajectory from the top center towards the bottom right.

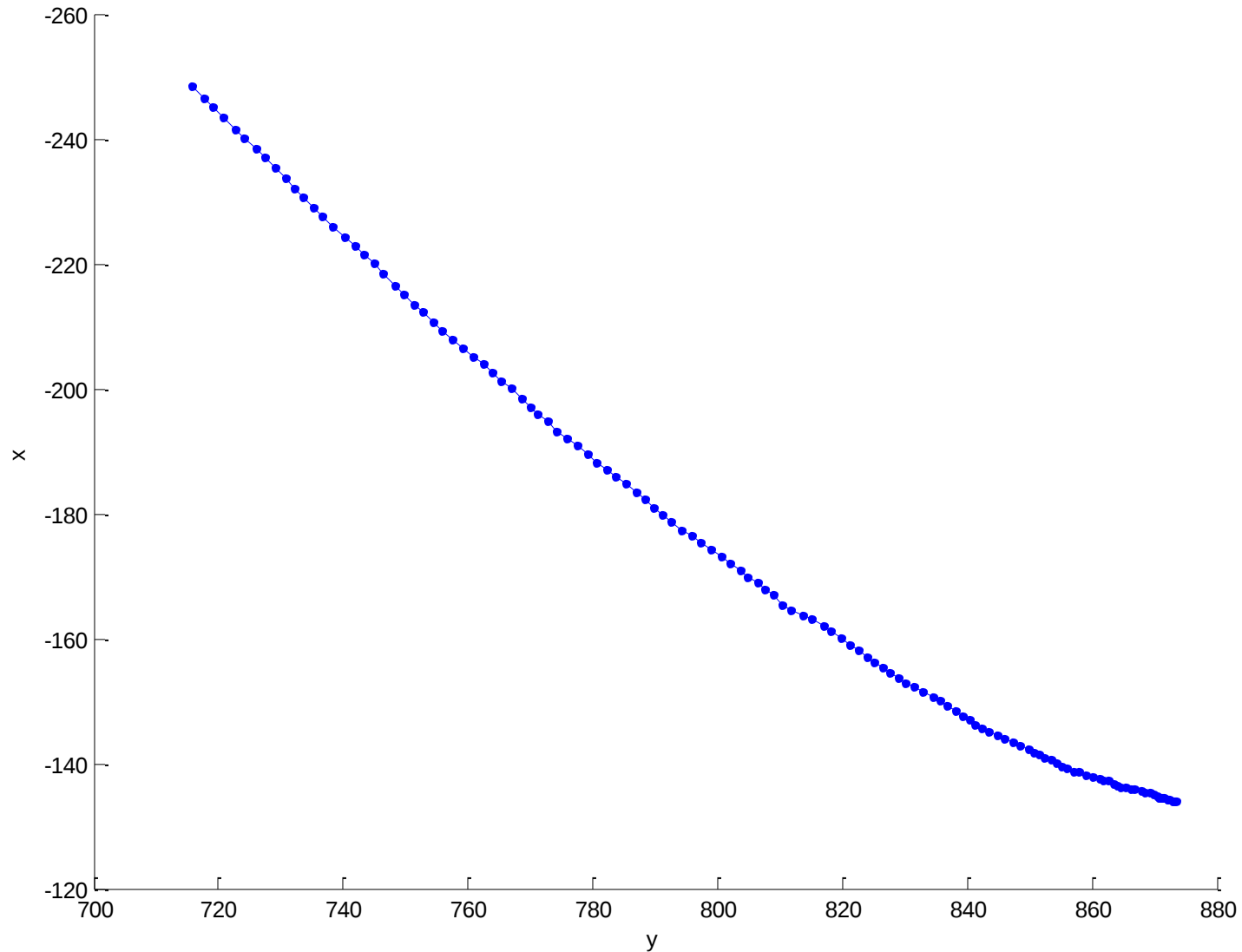
Below the main image, there is a green progress bar and a status bar. The status bar shows "Frame: 1604".

At the bottom, there are several control buttons and checkboxes:

- Buttons: Load Calibration, Calculate 3D coordinates, Number of points to display (All), ?
- Checkboxes: Center image after click, Auto advance, Display frame numbers, Draw epipolar lines
- Orientation of first calibration grid: ☒ Horizontal ☐ Vertical
- Missing data points: ☒ NaNs ☐ Clip ☐ Clip ends

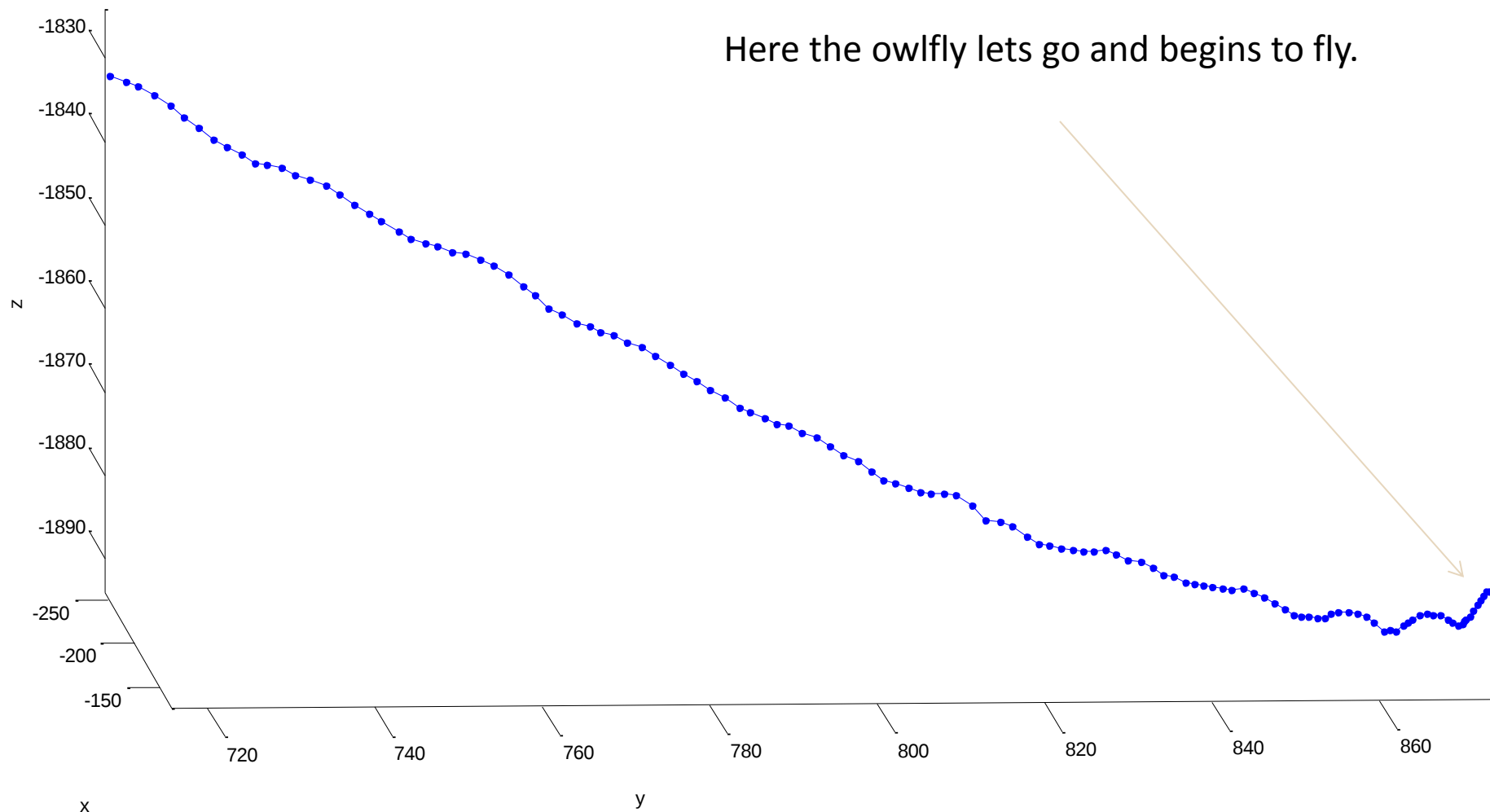


Tracking point on the head – this 2D view is similar to the tracked points shown on the previous slide.

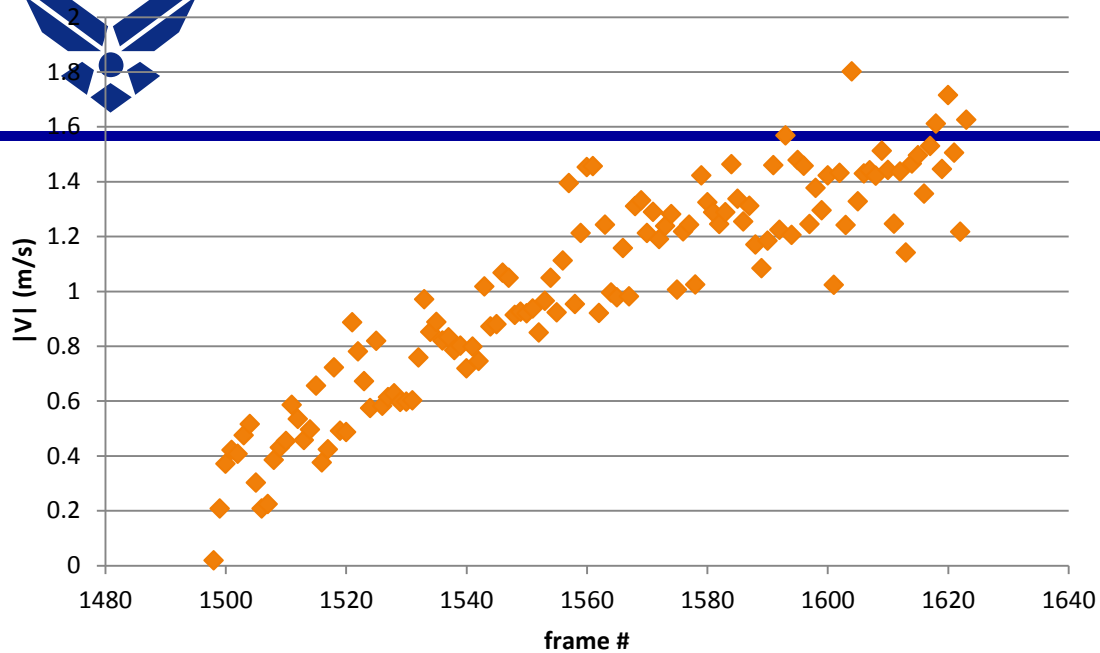
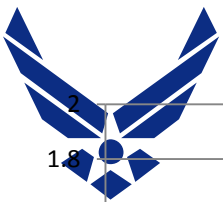




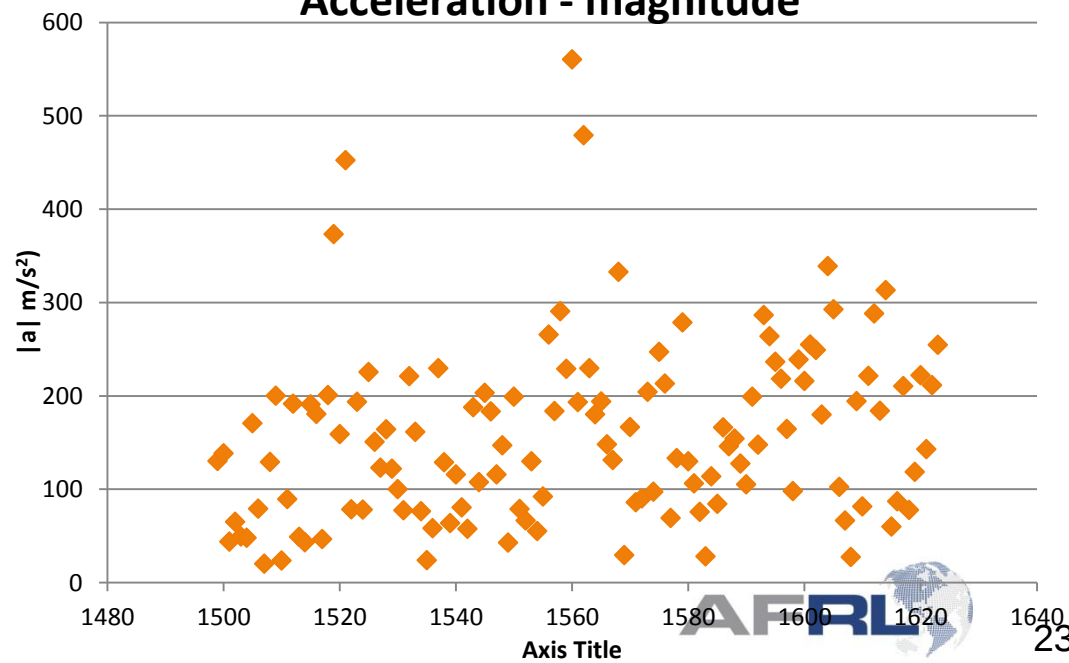
Tracking point on the head – 3D view, like previous plot, but slightly rotated. Showing some interesting features when the owlfly releases and begins to fly.



Velocity - magnitude



Acceleration - magnitude





Way forward.



- Wing beat frequency
- Recovery after perturbation.
 - Normalize to body size?
- Auto tracking vs. manual tracking.
- Manipulate sensors.
- Connect to physiology measured from sensors.

